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Investigation on Preliminary Phytochemical and Proximate Analysis of *Nelumbo nucifera* Gaertn Seeds



Shukla K*, and Chaturvedi N

Department of Food Science and Nutrition, Banasthali
University, Dist. Tonk, Rajasthan. 304022, India.

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ABSTRACT

Nelumbo nucifera Gaertn (*Nelumbonaceae*) is rhizomatous aquatic plant used in Ayurvedic medicine and its various parts have been employed for the treatment of ailments. The purpose of the study is to evaluate preliminary qualitative phytochemical constituents and proximate composition of *Nelumbo nucifera* seeds. The preliminary qualitative phytoconstituents were alkaloids, flavonoids, glycosides, steroids, tannins and terpenoids and proximate constituents were moisture, ash, crude protein, crude fat, crude fiber, vitamin C, iron and calcium. These parameters were evaluated with standard methods of Association of Official Analytical Chemists. The result revealed that the presence of alkaloids, flavonoids, glycosides, steroids, tannins and terpenoids in aqueous extract of *nelumbo* seeds. The proximate analysis of *nelumbo* seeds showed that moisture: 6.86 ± 0.17 g/100g, ash: 7.9 ± 0.06 g/100g, crude protein: 19.99 ± 1.77 g/100g, crude fat: 2.5 ± 0.28 g/100g, crude fiber: 1.73 ± 0.06 g/100g, carbohydrate: 61.02 ± 0.28 g/100g, vitamin C: 0.22 ± 0.01 mg/100g, iron: 14.37 ± 0.73 mg/100g and calcium: 29.42 ± 0.62 mg/100g. The study may conclude that *nelumbo* seed powder could be incorporated in food formulation as a good source of nutrients and the natural endowments of phytonutraceuticals showed its pharmacological and therapeutic potential apart from its nutritional essence which could be explored to provide affordable medicines to masses.

1. INTRODUCTION

Aquatic plants grow abundantly in lakes and waterways all over the world and categorized as floating submerged and emergent aquatic vegetation¹. For centuries, it has been perceived as a nuisance rather than resource as they block canals, hinder boat traffic and increase water borne diseases². In recent past numerous workers all over the globe have tried to explore the possibilities of using these plants as a source of animal feed and medicinal plants³.

In India, abundant aquatic plants like *Nymphoides*, *Rannunculus*, *Azolla pinnata*, *Nelumbo nucifera*, *Trapa natans* Linn., *Eichhornia crassipes*, *Ludwigia peploides*, *Nymphaea lotus* Linn., Cape Cod Grass, *Vallisneria spiralis*, *Typha latifolia* etc. species are found in different waterways⁴. Among them, *Nelumbo nucifera* is a rhizomatus aquatic which considered as a nutraceutically valued edible as a non-conventional vegetable. It is extensively grown in Australia, China, India, and Japan and accepted as health food for the reason that it contains rich amount of health-promoting phytochemicals, proteins and essential minerals⁵.

The seeds are also used indigenously in Ayurveda, Chinese and folk medicines to treat tissue inflammation, cancer, diuretics and skin diseases⁶. Because of the high nutritional value of fresh lotus, its consumption in recent years has increased rapidly, showing that lotus is a valuable food⁷, that have been used for medicinal properties that include improving learning and memory⁸, hepatoprotective⁹, anti-obesity, anti-HIV activity, anti-tumor effect¹⁰, diuretic activity, antipyretic activity, antidepressant and anti-inflammatory activity¹¹. Different classes of phytoconstituents have been isolated from various parts of *N. nucifera* which include alkaloids, steroids, triterpenoids, flavonoids, glycosides and polyphenols which may be used to treat hyperdipsia, dermatopathy, menorrhagia, leprosy, cancer, skin diseases, poison antidote and fever which have been reported to possess rich antimicrobial properties¹².

Information regarding to nutritional aspects of underutilized seeds will be a basic requirement, before consumption and commercialization. The major aim of the present study was to determine preliminary phytochemicals and nutritional components of *nelumbo* seeds.

2. MATERIALS AND METHODS

2.1. Sample collection and preparation:

Fresh seeds of lotus used in the present study were collected from village Ichoi, Milkipur, Faizabad, Uttar Pradesh. The plant seeds were separated from weeds, washed and rinsed with distilled water. It was sun-dried for 5 days, ground in a mortar with a pestle into coarse powder and packed in an air-tight plastic container for further analysis.

2.2. Preparation of aqueous extract and laboratory analysis:

The aqueous extract of lotus seeds was prepared by soaking 100 g of dried powdered samples in 500 ml of distilled water for 12 h. The filtrate of seeds powder was tested for the presence of various active principles namely flavonoids (Shonoda test), saponins (Froth Test), tannins (FeCl₃Test), glycosides (Molisch's Test), steroids (Libermann- Burchard's Test), terpenoids etc¹³. Determination of proximate composition was carried out in accordance with Association of Official Analytical Chemists¹⁴. This constitutes the different classes of nutrients present in the samples such as ash content was determined by weight difference after sample mineralization at 600°C for 6 h. Moisture content was determined by drying in an oven at 85°C to constant weight. Crude protein was determined indirectly from the analysis of total nitrogen (crude protein= amount of nitrogen×6.25) using Kjeldhal method by Kel Plus analyzer (Pelican, Model: KES-061). Crude fat was determined through Socs Plus system (Pelican, Model: SCS-6) by using petroleum ether. Crude fiber content of seeds was determined by digesting dry sample with 1.25% H₂SO₄, followed by 1.25% NaOH solution in Fibra Plus Fiber analyzer (Pelican, Model: FES-4). Vitamin C was obtained by titrimetric method by using 2-6 dichlorophenol indophenol dye. Carbohydrate content was estimated by subtracting the values of protein, moisture, ash, fiber and fat from hundred. Mineral elements estimations indicate the amount of inorganic elements present in the sample. The determination was carried out using standard procedures. The mineral element determined were Iron (Fe) by Wong's method and Calcium (Ca) by titration against standard potassium permanganate solution (KMnO₄). The data were expressed as Mean±S.E.M.

3. RESULTS AND DISCUSSION

Table1: Phytochemical screening of seed of the *Nelumbo nucifera*

Parameters	<i>Nelumbo nucifera</i> (Seed)
Alkaloids	+ ve
Glycosides	- ve
Saponins	- ve
Tannins	+ ve
Flavonoids	+ ve
Terpenoids	+ ve
Steroids	+ ve

Phytochemicals are biologically active compounds found in small amounts which are not established nutrients but on the other hand contribute significantly to protection against degenerative diseases¹⁵. The qualitative phytochemical analysis of the aqueous extracts of *Nelumbo nucifera* seed powder showed positive results for the presence of alkaloids, tannins, flavonoids, terpenoids and steroids while glycosides and saponins were absent which are shown in Table 1. These obtained results were corroborative with the reports of that its seeds contain the active principles such as terpenoids, steroids, carbohydrates, flavonoids and phenolic compounds which shows an antioxidant, anti-diabetic, anti-obesity, anti-fungal activity^{16,17,18}. Epidemiological studies have shown that phytochemical components possess rich antioxidant properties and have been reported to exert multiple biological effects including antimicrobial, anti-inflammatory activities, free radical scavenger, pain relievers, tranquilizers and metal chelators^{19,20}. They are effective in reducing cardio-cerebrovascular diseases, cancer mortality, hypercholesterolemia and antibiotic properties²¹.

Table 2: Proximate composition of *Nelumbo nucifera* seed powder

Parameters	<i>Nelumbo nucifera</i> (Seed)
Moisture(g/100g)	6.86±0.17
Ash(g/100g)	7.9±0.06
Protein(g/100g)	19.99±1.77
Fat (g/100g)	2.5±0.28
Fiber(g/100g)	1.73±0.06
Carbohydrate(g/100g)	61.02±0.28
Vitamin-C(mg/100g)	0.22±0.01
Iron (mg/100g)	14.37±0.73
Calcium(mg/100g)	30.60±0.62

Data are means ± SE (n=3).

The value of proximate composition of seed powder of *nelumbo nucifera* is presented in Table 2. The table shows the Mean±SEM (g/100g) of moisture, ash, protein, fat, fiber, carbohydrate, vitamin C, iron and calcium content. The proximate data revealed that the moisture content (g/100g) was low (6.86±0.17) which was advantageous for prolonging the shelf life of the seeds. Since higher content could cause decomposition of fatty acids by microbial action. The anticipated value of moisture content (8.20±0.16 to 8.17±0.23) of lotus seed was approximately parallel which was observed through Wu et al.²². The ash content (g/100g) of seeds obtained was 7.9±0.06 which was higher when compared to the study carried out by Sathithon and Yan-bin, (4.02) ash content²³. The quantity of ash in any seed sample assumes importance as it determines the nutritionally important minerals²⁴.

The crude protein content (g/100g) of seeds was 19.99±1.77 which was close agreement with the results of Chouaibi and his coworkers that the crude protein content of *Zizyphus lotus* L. seed was 19.11±0.03g/100g²⁵. The high protein content in lotus seeds emphasizes their value as a vital source of nutrients. Aside contributing to diets, the relative impact of proteins in body system should not be over looked. As chemical compounds they repair and replace worn out cells, form

structural and globular materials that hold the body, form blood proteins, boost immune system etc²⁶. The fat content (g/100g) of lotus seed was 2.5 ± 0.28 . This data was comparable in fat content with a value 2–3% in chestnut^{27,28,29}, in comparison with other nuts such as almonds: 53.9%, hazelnuts: 35.9% and walnuts: 63.4%. For Spanish chestnuts cultivars, the average value was 3.0% with a range from 2.8% to 3.2%³⁰. Generally, fats have many functions aside insulation and conservation of body temperature in organisms, their fatty acid components such as lauric acid, etc. have been reported to improve health³¹.

The fiber content (g/100g) of lotus seed recorded was 1.73 ± 0.06 while comparing to the study of Mohammed et al that the fiber content of *Nymphaea* lotus seeds powder was 1.60 ± 0.20 ³². Dietary fibers alter the colonic environment in such a way as to protect against colorectal diseases. It provides protection by increasing fecal bulk, which dilutes the increased colonic bile acid concentrations that occur with a high fat diet³³. Evidence from epidemiological studies suggest that increased fiber content in the body of organisms could reduce incidence of diseases like diabetes, high blood pressure, piles, digestive disorders etc.^{34,35}. The seeds were abundant in carbohydrate content (61.02 ± 0.28 g/100g), which was comparable with the findings of Parks, white lotus cultivars had range between 65-66 g/100g in carbohydrate content³⁶. Lotus seeds contained a high amount of carbohydrates which might be due to low lipid content. Observed carbohydrates in the investigated sample may be an indication that they could produce energy to provide power to the cells and tissues of the body for consumption and may play an important role in structural materials for the body system³⁷.

The amounts of vitamin C (mg/100 g) present in *nucifera* seeds was 0.22 ± 0.01 . The result suggests that plant seeds are a good source of vitamin C. While comparing our study on vitamin C content of two varieties of water chestnuts (*Trapa sp.*); 1.1 mg/100 g in green variety and 0.9 mg/100g in red variety³⁸. An adequate amount of iron was found to be 14.37 ± 0.73 mg/100g in *nelumbo* seeds as predicted in Table 2. Similar study was observed by Bhat and Shridhar, 2008 that seeds contain 16.4 ± 4.44 mg/100g iron content³⁹. Chaturvedi et al. reported that iron acts as antioxidant and is involved in strengthening the immune system⁴⁰. The present study revealed that the calcium content (mg/100g) in seed powder was 29.42 ± 0.62 which was higher than the observation obtained from Indrayan et al. and Mani et al. (22%) and (25%) respectively^{41,42}. Calcium constitutes a large proportion of the bone, human blood, and extracellular fluid; it is

essential for the normal functioning of cardiac muscles, blood coagulation and milk clotting, and the regulation of cell permeability. Calcium absorbs vitamin B12 from intestinal tract. It also plays an important role in nerve impulse transmission and in the mechanism of neuromuscular system. These elements support human biochemical processes by serving structural and functional roles as electrolytes⁴³.

4. CONCLUSION

The results of the present study confirmed the folkloric usage of *Nelumbo nucifera* seeds that possess certain compounds with bioactive principles that can be used as active agents in new drugs for treating various metabolic diseases. From the nutritional composition data analysis, it was found that seed can be successfully utilized as an important source of protein and carbohydrates for humans, livestock and also contains appreciable concentrations of phytochemicals which may provides a scientific rationale for the use of lotus seeds in the traditional medicine for the treatment of metabolic disorders.

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6. REFERENCES

1. Gupta OP. In: Aquatic weed management. IVF. Pub. Co. Ltd. New Delhi; 2001.
2. Banerjee A, Matai S. Composition of Indian aquatic plants in relation to utilization as Animal Forage. J Aquatic Plant Manage. 1990; 28:69-73.
3. Deepa PK, Usha PTA, Chandrasekharan Nair AM, Prasannakumari KT. Antipyretic activity of seeds from Red and White type of lotus (*Nelumbo nucifera*) in Albino rat. J of Veterinary World. 2009; 2: 213-214
4. Bhat NA. Some common wild flowers of Srinagar. Makoff printers Delhi; 2002.
5. Sridhar KR, Bhat R. Lotus: a potential nutraceutical source. J Agri Technol. 2007; 3:143-155.
6. Liu CP, Tsai WJ, Lin YL, Liao JF, Chen CF, Kuo YC. The extracts from *Nelumbo nucifera* suppress cell cycle progression, cytokine genes expression, and cell proliferation in human peripheral blood mononuclear cells. Life Science. 2004; 75:699–716.
7. Yen GC, Duh PD, Su HJ, Yeha CT, Wu CH. Scavenging effects of lotus seed extracts on reactive nitrogen species. Food Chem. 2006; 94:596–602.
8. Gong YS, Liu LG, Xie BJ, Liao YC, Yang EL, Sun, Z.D. Ameliorative effects of lotus seed pod proanthocyanidins on cognitive deficits and oxidative damage in senescence-accelerated mice. Behav. Brain Res. 2008; 194: 100-107.
9. Sohn DH, Kim YC, Oh SH, Park EJ, Li X, Lee BH. Hepatoprotective and free radical scavenging effects of *Nelumbo nucifera*. Phytomedicine. 2003; 10:165–169.

10. Duan Y, Zhang H, Xie B, Yan Y, Li J, Xu F, Qin Y. Whole body radio protective activity of an acetone-water extract from the seed pod of *Nelumbo nucifera* Gaertn. Seedpod. Food Chem. Toxicol. 2010; 48: 3374-3384.
11. Sugimoto Y, Furutani S, Nishimura K, Itoh A, Tanahashi T, Nakajima H, Oshiro H, Sun S, Yamada J. Antidepressant-like effects of neferine in the forced swimming test involve the serotonin1A (5-HT1A) receptor in mice. Eur. J. Pharmacol. 2010; 634(1-3):62-67.
12. Bramwell R, Costa D, Giese N, Kang S, Seamon E, Ulbricht CW, Weissner W. (eds) Lotus (*Nelumbo nucifera*). Natural Standard Monograph (www.naturalstandard.com). Cambridge, MA, USA; 2008.
13. Harborne JB. Phytochemical methods. Guide to modern techniques of plant analysis, 2nd ed.; Chapman and Hall: India, 1991; 653.
14. AOAC. (Association of Official Analytical Chemists-International). *Official Methods of Analysis*, 18th ed., W Hortwitz and GW Latimer (ed.), AOAC-Int. Gaithersburg, MD, USA; 2005.
15. Dreosti IE. Recommended dietary intake levels for phytochemicals: Feasible or Fanciful. Asia Pacific J Clin Nutr. 2000; 9: 119-122.
16. Chakravarthi PV, Gopakumar N. Evaluation of Analgesic Activity of Lotus seeds (*Nelumbo nucifera*) in Albino Rats. Veterinary World. 2009; 2(9):355-357.
17. Rai S, Wahile A, Mukherjee K, Saha BP, Mukherjee PK. Antioxidant activity of *Nelumbo nucifera* (sacred lotus) seeds. J Ethnopharmacol. 2006; 104:322-327.
18. Nagarajan DC, Subhan AG, Nyeen P. Antioxidant and hepatoprotectant action of crude alcoholic extract of *Wrightiata mentosa*. Orient Pharm Exp Med. 2008; 81:241-248.
19. Nakayama T, Yamada M, Osawa T, Kawakishi S. Suppression of active oxygen induced cytotoxicity by flavonoids. Bio Chem Pharmacol. 1998; 45:265-267.
20. Cook NC, Samman S. Flavonoids-chemistry, metabolism, cardio protective effects and dietary sources. Nutri Biochem. 1996; 7:66-76.
21. Panthari P, Kharkwal H, Joshi DD. Investigation on Myrica Nagi leaves: Phytochemical screening and physicochemical evaluation. World J Pharm Pharm Sci. 2013; 2(5):2867-2873.
22. Wu, Zheng YB, Chen TQ, Yi J, Qin LP, Rahman K, Lin WX. Evaluation of the quality of lotus seed of *Nelumbo nucifera* Gaertn from outer space mutation. Food Chem. 2007; 105:540-547.
23. Sathithon P, Yan-bin X. Effect of sprouting on the chemical and nutritional qualities and phenolic alkaloid content of lotus (*Nelumbo nucifera* Gaertn.) seeds. Afr J Food Sci. 2010; 6(7):204-211.
24. Vadivel V, Janardhanan K. Agrobotanical traits and chemical composition of *Cassia obtusifolia* L.: a lesser-known legume of the Western Ghats region of south India. Plant Food Hum Nutr. 2002; 57:151-1643.
25. Chouaibi M, Mahfoudhi N, Rezig L, Donsi F, Ferrari G, Hamdi S. Nutritional composition of *Zizyphus lotus* L. seeds. J Sci Food Agric. 2012; 92: 1171-1177.
26. Olusanya JO. Proteins, In: Essentials of food and nutrition. Apex Books Limited, Lagos 2008; 13-21.
27. Breisch H. Cha[^]tagnes et marrons. CTIFL, Paris, 1995.
28. McCarthy MA, Meredith FI. Nutrient data on chestnuts consumed in the United States. Econ Bot. 1988; 42 (1):29-36.
29. Ensminger AH, Ensminger ME, Konlande JE, Robson JRK. The Concise Encyclopedia of Foods and Nutrition, second ed. CRC Press, BocaRaton, 1995.
30. Pereira-Lorenzo S, Ramos-Cabrer AM, Díaz-Hernández MB, Ciordia-Ara M, Rios-Mesa D. Chemical composition of chestnut cultivars from Spain. Scientia Horticulturae. 2006; 107, 306-914.
31. Fite B. The Healing miracle of coconut oil. Piccadilly Books Ltd, Health wise Publications Colorado Springs, Co 2000; 1-4.
32. Mohammed HA, Uka UN, Yauri YAB. Evaluation of nutritional composition of water lily (*Nymphaea lotus* Linn.) from tatabu flood plain, North- central, Nigeria. J Fish Aquat Sci. 2012; 1-4.
33. Dillard CJ, GermanEnig M. Coconut: In support of good health in the 21st century. 1990; 1-27.
34. SACN. Draft SACN position statement in dietary fibre and health and the dietary fibre definition. 2008.
35. Walker ARP. The relationship between bowel cancer and fibre content in the diet. Am J clin Nutr. 1978; 31: 245-251.

36. Park YS, Towantakanit K, Kowalaka T, Jung ST, Ham KS, Heo BG, Cho JY, Yun JG, Kim HJ, Gorinstein S. Bioactive Compounds and Antioxidant and Antiproliferative Activities of Korean White Lotus Cultivars. *J Med Food*. 2009; 12 (5): 1057–1064.
37. Voet DJ, Voet JG, Pratt CW. *The Principles of Biochemistry*. 3rd Edn., John Wiley & Sons 111 River Street, Hoboken:2008; 74-219.
38. Faruk MO, Amin MZ, Sana NK, Shaha RK, Biswas KR. 2012. Biochemical Analysis of Two Varieties of Water Chestnuts (*Trapa Sp.*). *Pak J Biol Sci*.2012; 15: 1019-1026.
39. Bhat R, Sridhar KR. Nutritional quality evaluation of electron beam-irradiated lotus (*Nelumbo nucifera*) seeds. *Food Chem*. 2008; 107:174–184.
40. Chaturvedi VC, Shrivastava R, Upreti RK. Viral infections and trace elements: A complex interaction. *Curr. Sci*. 2004; 87:1536–1554.
41. Indrayan AK, Sharma S, Durgapal D, Kumar N, Kumar M. Determination of nutritive value and analysis of mineral elements for some medicinally valued plants from Uttaranchal. *Curr. Sci*.2005; 89: 1252-1255.
42. Mani SS, Subramanian IP, Pillai SS, Muthusamy K. Evaluation of hypoglycemic activity of inorganic constituents in *Nelumbo nucifera* seeds on streptozotocin-induced diabetes in rats. *Biol Trace Elem Res*. 2010; 138: 226-37.
43. Nelson DL, Cox MM. *Lehninger Principles of Biochemistry*. 5th ed.; W.H. Freeman &Company: Madison Avenue, New York, 2008; 343.

