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Marine Sponge: Natural Reservoir of a Myriad of Bioactive Compounds



Amitha Kurian¹, Preetham Elumalai^{1, 2}*

¹ Department of Biochemistry, School of Aquatic Food Products and Technology, Kerala University of Fisheries and Ocean Studies, Cochin-682506, Kerala, India

^{2*}Centre for Bioactive Substance, School of Aquatic Food Products and Technology, Kerala University of Fisheries and Ocean Studies, Cochin-682506, Kerala, India

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ABSTRACT

Biodiversity in the marine environment exceeds that of the terrestrial environment. But exploration of marine natural products is still in the infant stage. Among the diverse bioactive compounds which are discovered from marine organisms, sponges have been assigned a high priority as they have been accepted as a promising organism for the discovery of pharmacologically active drug leads. Marine sponges have provided successful examples that prove natural products derived from them had the therapeutic potential to develop as future drugs against different types of cancer, viral diseases, malaria, inflammation etc. All these findings attract the pharmaceutical field to involve in marine sponge research though more research is required to confirm promising results. Today scientists engage in studying the mode of action of the compounds from sponges at molecular level and also their possible role in treating diseases as the fascinating marine life continues to give natural products. This review aims at covering the new developments in the field of marine sponge research and bioactive compound discovery.





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INTRODUCTION

Discovery of marine derived natural products began in the 1950s with the discovery of nucleoside derivatives in the sponge *Tethya crypta* by Bergmann and Feeney. But it was only by the end of 1960, marine organisms were utilized for deriving bioactive compounds. Marine habitat is blessed with a plethora of bioactive compounds that are considered to be a source of potential drugs. 70% of the earth's area is comprised of ocean and 95% of the biosphere is represented by the marine ecosystems. 33-34% animal phyla live in marine environment. Marine bioactive compounds mainly produced by microbes, sponges, gorgonians, soft and hard corals seaweeds, and other marine organisms could be divided into steroids, terpenoids, isoprenoids, norisoprenoids, quinones, brominated compounds, nitrogen heterocyclics, and nitrogen sulfur heterocyclics. It is believed that secondary metabolites, a diverse array of compounds are produced due to factors like self defense, nutrition and competition for space. Secondary metabolites are regarded as communication molecules that are used by animals for intercommunication between themselves and their environment. These communication molecules evolved within the scope of symbiotic interrelationship (Sima and Vetvicka, 2011) have become the new interest of pharmaceutical industry for new drug leads. Bioactive compounds produced by phylogenetically diverse organisms are small molecules up to 3000 Daltons.

In the largely unexplored marine ecosphere, microorganisms form highly specific and symbiotic relationships with sessile, soft bodied and morphologically defenseless filterfeeding organisms like sponges. Microbial symbionts usually produce bioactive natural products. Marine bioactive natural products could be secondary metabolites, enzymes, lipids or heteropolysaccharides (Proksch et al., 2002). Metabolites are mainly of two types, primary and secondary. Primary metabolites that are formed by a limited number of metabolic reactions are considered essential for growth and life in all living systems. When primary metabolites serve as building blocks for the synthesis of macromolecules, proteins, nucleic acids, carbohydrates and lipids, secondary metabolites such as terpenes, polyketides, and alkaloids are not essential for the producing organism and are formed from primary metabolites. Majority of the secondary metabolites play a role in enhancing the survival fitness of the organism and form best examples as chemical weapons used against bacteria, fungi, insects and large animals. Secondary metabolites have captured the attention and interest of pharmaceutical companies. Swift progress has occurred in the field of novel marine natural product discovery from marine organisms (Blunt 2013). Sponges contribute to

nearly 30% of all marine natural products discovered so far. This makes them a prolific producer of bioactive compounds and this is mainly due to the diversity of microbes living inside the sponge body.

Overview of marine sponges, symbionts and bioactive compounds

Marine sponges are multicellular pore bearing invertebrates that belong to the phylum porifera. They have been living on the earth for millions of years and are the simplest and oldest organisms. Regardless of extreme temperatures, they can be found 5-50 meters deep. Sponges have the ability to filter up to 2000 liters of seawater each day and are very efficient in taking up nutrients as well as microorganisms from the seawater. They mainly dominate in many benthic habitats. Sponges are sessile without any tissues or sensory organs but possess different types of cells that help to perform all types of bodily functions.

Complex microbial communities of sponges are renowned for biotechnological and ecological importance. Microbial communities such as bacteria, fungi, microalgae etc. are usually found in sponges comprising more than 40% of sponge body (Wang 2006). Bacteria and fungi are the common symbionts in sponges where bacterial symbionts outnumber fungal symbionts (Taylor *et al*, 2007). The major bacterial phyla that are associated with marine sponge include Cyanobacteria, Bacteriodetes, Actinobacteria, Chloroflexi, Proteobacteria, Nitrospira, Poribacteria, Verrucomicrobia Planctomycetes, Archaea and Acidobacteria. Fungi, microalgae and viruses also inhabit sponge body, though little is known about viruses. Among the microbes associated with marine sponges, it has been identified that the prime producers of bioactive compounds belong to the bacterial phylum Actinobacteria and the fungal division Ascomycota (Kennedy *et al.*, 2009). The three main classes of sponges are Calcarea, Demospongiae and Hexactinellida. Among these, the abounding sources of bioactive compounds have been recognized in the class Demospongiae and the orders Halichondrida, Poecilosclerida and Dictyoceratida.

Many studies have shown that secondary metabolites produced by sponges often have defensive roles of protecting themselves from microbial infections, biofouling, predator attacks, and overgrowth by other sessile organisms. Five compounds isolated from the sponges have successfully used for medicinal purposes, and 13 anticancer compounds are in clinical trial and almost 100 compounds are under preclinical trial till 2014 (Mayer *et al.*, 2010). Marine invertebrates are the prolific producers of natural compounds and among them

phylum porifera occupies the top position due to the sheer amount of novel pharmacologically active metabolites produced, that are of immense use in the treatment of human diseases (Lee *et al.*, 2001) (Jensen 1994) (*Taylor et al.*, 2007). Due to the potential of sponge associated symbionts to produce natural compounds against many diseases like cancer, autoimmune diseases, viral diseases, inflammations and malaria (Molinski *et al.*, 2009) (Simmons *et al.*, 2005) (Gordaliza 2010) (Alcaraz 2006) they have become a major field of interest to scientists from different disciplines.

Bioactive sponge metabolites

Sponges have been considered as the topmost producers of natural products since the chemical diversity of sponge products is remarkable and they include bioactive terpenes, fatty acids, sterols, peroxides, nucleosides and derivatives of amino acids that are halogenated. Some of the major bioactive compounds discovered from marine sponges include anti-tumour, anti-malarial, anti-viral, antibiotic, immunosuppressive, neuro suppressive, anti-inflammatory and antifouling agents (Blunt *et al.*, 2013) and a few important compounds have been described below.

Anti-oxidants and anti-inflammatory compounds

Several bioactive compounds from marine sponges that possess anti-inflammatory, antioxidant and radical scavenging properties have been discovered in the last few years. A recent study on the antioxidant property of *Tedania anhelans* confirmed the antioxidant potential of its associated bacteria *Bacillus* species (D.Balakrishnan *et al.*, 2014).

Table 1: Closest match of *Tedania angels* associated bacteria showing anti-oxidant activity (Reference D. Bal akrishnan *et al.*, 2014)

Strain	Source	Nearest relative	Identity [%]	Phylogenetic affiliation (Phylum)
Bacillus licheniformis KDRSS1	Tedania anhelans	Bacillus licheniformis DSM 13 (CP000002)	97	Firmicutes
Bacillus subtilis KDRSS4	Tedania anhelans	Bacillus subtilis DSM 10 (AJ276351)	97	Firmicutes
Bacillus subtilis KDRSS6	Tedania anhelans	Bacillus subtilis strain 168 (AL009126)	97	Firmicutes

In another study Ageloline A, new antioxidant and antichlamydial quinolone from the marine sponge-derived bacterium *Streptomyces* sp. SBT345 was derived that showed the antioxidant potential of the novel compound (Cheng *et al.*, 2016). Phorbaketal A, isolated from the marine sponge *Phorbas* sp., was shown to inhibit the production of inflammatory mediators via down-regulation of the NF-κb pathway and up-regulation of the HO-1 pathway (Yun-Ji Seo *et al.*, 2015).

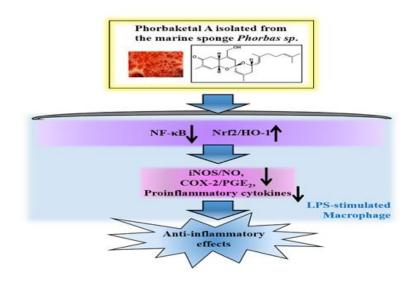


Figure 1. Mode of action of Phorbaketal A (Reference Yun-Ji Seo et al., 2015)

A study on the anti-inflammatory activities of extracts from bacteria associated with marine sponge *Theonella sp* proved the cytotoxicity and anti-inflammatory activity of secondary metabolites produced by these bacteria when studied using macrophage cell lines (Siti Aisha Mohd Radzi *et al.*, 2015).

The extracts of sponges *Callyspongia, Niphates and Stylissa* collected from the Red sea showed potent reducing power and inhibitory effects on oxidative stress (Shaaban M *et al.*, 2012). A scientific study in the sponge *Pandaros acanthifolium* showed cytoprotective as well as antioxidant properties (Berrué F *et al.*, 2012). Another study proved the presence of metabolites like phenolic compounds, alkaloids and polyketides that exhibited antioxidant properties (Longeon A *et al.*, 2011).

Anti-tumour compounds

A derivative of the nucleosides spongothymidine and spongouridine isolated from *Tectitethya crypta* known as Ara-C is considered as the pioneer anticancer agent that can be used against leukemia (Proksch P *et al.*, 2002). It has been currently used in clinical trials against myeloid neoplasms along other anticancer drugs (Feldman *et al.*, 2011). Almost 39 compounds from sponges that can induce apoptosis and that can act as anticancer agents have been identified (Essack *et al.*, 2011).

Renieramycin isolated from the sponge genera *Reniera* has shown to induce apoptosis in lung cancer cells (Halim *et al.*, 2011).

Figure 2. Structure of Renieramycin (Reference Magbubah Essack et al., 2011)

Researchers have proven that a lectin from the sponge *Cinachyrella apion* has the ability to induce cell death in tumor cells (Valeriote *et al.*, 2012).

From the marine sponge *Monanchora pulchra*, a novel polycyclic guanidine alkaloid isolated is Monanchocidin that induced cell death in human monocytic leukemia, human cervical cancer (HeLa) and mouse epidermal cells (Guzii *et al.*, 2010).

Figure 3. Structure of Monanchocidin (Reference Magbubah Essack et al., 2011)

Antitumour effect of cyclodepsipeptides jasplakinolide isolated from the marine sponge *Jaspis splendens* and its analogs neamphamides and geodiamolides are generally accepted as actin-polymerising and actin-stabilising drug and were tested to possess potent cytotoxic activities (Rosa Lemmens-Gruber 2014).

Another study investigated the anticancer properties of marine sponge *Hyattella cribriformis* ethyl acetate (EA) fraction in various cancer and normal cell lines that showed potent anticancer activity by promoting tubulin polymerization as evidenced mitotic arrest and induction of apoptosis (Pazhanimuthu Annamalai *et al.*, 2015).

Guanidine Alkaloids from the marine sponge *Monanchora pulchra* was found to show cytotoxic properties and prevented EGF-Induced neoplastic transformation *in vitro* and this study proved that guanidine marine alkaloids hold potential to eliminate human cancer cells and prevent cancer cell formation and spreading (Sergey *et al.*, 2016). Four new tetracyclic meroterpenes, dysiherbols A–C and dysideanone E isolated from a *Dysidea sp.* marine sponge were found to possess NF-κB inhibitory and cytotoxic activity (Wei-Hua Jiao *et al.*, 2016).

Four triterpenes, isolated and identified as sipholenone A, sipholenol A, neviotine A and sipholenol L from the Red Sea Sponge, *Siphonochalina siphonella* were found to be cytotoxic to MCF-7 and HepG-2 cancer cell lines (SM Al-Massarani *et al.*, 2015). Methanol extracts of *Haliclona species* indicated that it could induce apoptosis via the JNK-p53 pathway and caspase-8 in nonsmall cell lung cancer A549 cells (Woori Bae *et al.*, 2015).

Immunosuppressive compounds

Promising immunomodulating activities have been attributed to marine natural compounds. Immunomodulators may be biological or synthetic substances, that can suppress, modulate or stimulate, the immune system including both adaptive and innate immunity of the immune response. Immune system suppression is desired in cases of hypersensitivity to certain antigens or organ transplantations. It was by the end of 1980s, immunosuppressive compounds from marine sponges were derived. Two compounds with important immunosuppressive activity were discovered in Agelas flabellrformis whose structures determined were found to be effective in suppressing the response of murine splenocytes (Gunasekara et al., 1989). In a study conducted on the marine sponge Callyspongia difusa almost 10 marine bacterial strains were isolated that showed antagonistic activity against clinical bacterial pathogens. This led researchers to suggest that the sponge associated bacterial strain Virgibacillus species may possess the potential to contribute to the discovery of novel antibiotics against infections and also for the production of potential immunomodulators (Kalirajan et al., 2013). Immunomodulatory potential exhibited by the marine sponge Aurora globostellata was proven by oral administration to Wistar rats. The results obtained indicated that extracts possess immunosuppressant activity (Chairman K et *al.*, 2013)

Solomonsterol A, considered as a selective pregnane X receptor (PXR) agonist has been isolated from the marine sponge *Theonella swinhoei*. It has been found to possess anti-

inflammatory activity that could attenuate systemic inflammation and immune dysfunction in mouse models of rheumatoid arthritis (Andrea Mencarelli *et al.*, 2014).

Fig 4.Structure of Solomonsterol A (Ref: Valentina Sepe et al., 2012)

Studies of Mayer *et al.*, 2000; 2004; 2011, has shown that the polyoxygenated sterols identified from Dysidea species possess immunosuppressive ability to block the binding of interleukin 8 (IL-8), to the IL-8 receptor (de Almeida Leone *et al.*, 2000). Likewise, from Mycale species, Pateamine A that can act as the selective inhibitors of the production of (IL-2) has been derived. Interleukin 2 plays a role in antigen-antibody reaction through the activation of B cells and T resting cells thus producing secondary immune response (Pattenden *et al.*, 2004).

Fig 5.Structure of Pateamine (Ref: Gerald Pattenden 2004)

A novel family of closely related N-acyl dopamine glycosides from the Icelandic marine sponge *Myxilla incrustans* were tested for immunomodulating activity in an in vitro dendritic cell model (Einarsdottir E *et al.*, 2016).

Antiviral and antimalarial compounds

Ara-A (vidarabine) isolated from sponge *Tethya crypta* has been considered as the most important antiviral agent as it inhibits DNA synthesis of herpes, vaccinia and varicella viruses. Such compounds are produced by enzyme clusters in sponges (Villa 2010).

The extracts from sponge *Petromica citrina* produced by the *Bacillus sp*ecies gives promising results for the treatment of hepatitis C (Santiago *et al.*, 2013). Psammaplin derivatives identified in the Indonesian marine sponge *Aplysinella strongylata* showed antimalarial activity against *falciparum* malaria parasites (Mudianta *et al.*, 2012). Diterpenes from the sponge *Stylissa* exhibit antimalarial activity (Chanthathamrongsiri *et al.*, 2012).

From the marine sponge *Verongula rigida*, Bromo tyrosine derived compounds were screened for *in vitro* activity against parasitic protozoa such as *Leishmania panamensis*, *Plasmodium falciparum and Trypanosoma cruzi*. Some of the compounds showed potent and selective anti-parasitic activity (Galeano *et al.*, 2011).

A diterpenoid showing potent anti-malarial activity was isolated from the marine sponge *Hymeniacidon sp* (Avilés 2010).

In a study conducted on the marine sponge *Stylissa carteri* alkaloids, debromohymenialdisine (DBH), hymenialdisine (HD), and oroidin exhibited the ability to inhibit HIV-1 replication (Aubrie O'Rourke *et al.*, 2016).

Two novel asteltoxins named asteltoxin E and F, and a new chromone, together with four known compounds isolated from the marine sponge *Callyspongia sp.*, erived fungus, Aspergillus sp. SCSIO XWS02F40 was found to possess antiviral activities against H1N1 and H3N2 (Yong-Qi Tian *et al.*, 2016).

Batzelladine- and Crambescidin-like guanidine alkaloids isolated from *Poecilosclerida* marine sponge were found to exhibit antiviral activity against different viruses such as Human immunodeficiency virus (HIV-1), Herpes simplex virus (HSV-1), and Human hepatitis B virus (HBV) (Estelle Sfecci *et al.*, 2016).

A polyketide endoperoxide, Plakortin, isolated from the sponge *Plakortis simplex* was found to show oxidative stress mediated antiparasitic activity against malaria parasite *Plasmodium* falciparum (Oleksii A. Skorokhod *et al.*, 2015). A compound stachybotrin D, isolated from

a sponge associated fungus *Stachybotrys chartarum*, was found to exhibit anti-HIV activity and this was found to be possible by targeting reverse transcriptase (Ma *et al.*, 2013). Li *et al.* studied the property of another anti-HIV compound from *Stachybotrys chartarum* and found that the compound chartarutine B resulted in 50% inhibition of HIV-1.

Figure 6: Structure of Stachybotrin D (Modified from Anak Agung Gede Indraningrat et al.,2016)

Figure 7: Structure of chartarutine B (Modified from Anak Agung Gede Indraningrat et al., 2016)

Antibiotics

Halogenated alkaloids from the marine sponge *Lotrochota purpurea* exhibited inhibitory activity against diseases related to fungi and bacteria (Shen *et al.*, 2012). Alkaloids isolated from the marine sponge *Agelas mauritiana* showed antifungal activity against *Cryptococcus neoformans*, antileishmanial activity *in vitro* and antibacterial activity against *Staphylococcus aureus* and methicillin-resistant *S. aureus in vitro* (Yang *et al.*, 2012).

Diterpene isonitriles isolated from the marine sponge *Cymbastela hooperi*, and the sesquiterpene axisonitrile-3, isolated from the sponge *Acanthella kletra*, were evaluated in a series of bioassays including anti-algal, anti-photosynthetic, antibacterial, antifungal, and

anti-tubercular. The results showed that the majority of the tested compounds were active in at least two of the applied test systems (Wright *et al.*, 2011).

In another study, marine sponge's sediments and sponge-derived actinomycetes were isolated and tested for bioactive metabolites with antimicrobial and antifungal activity. Nine of the fifteen active extracts were active against multiresistant gram-positive bacteria and/or fungal indicator organisms, including vancomycin-resistant Enterococcus faecium and multidrug-resistant *Candida albicans* (Engelhardt *et al.*, 2010).

In an extensive study using marine sponge, *Neopetrosia exigua* it was concluded that the antimicrobial activities of *N. exigua* fractions evaluated using disc diffusion and microdilution methods showed that the active metabolites were present in n-hexane, CH₂Cl₂, nBuOH, and water fractions and *Staphylococcus aureus* was the most susceptible microbe evaluated (Ibrahim Majali *et al.*, 2015). A new anti-MRSA (Methicillin resistant *Staphylococcus aureus*) compound producing *Streptomyces* strain isolated from a sponge was found to show cell wall destructing property (Appadurai Muthamil Iniyan *et al.*, 2016).

A study on the marine sponge *Dysidea granulose* proved that polybrominated diphenyl ether, 2-(2',4'-dibromophenoxy)-3,5-dibromophenol proved that they exhibit potent and broad antibacterial activity against methicillin resistant *Staphylococcus aureus* (MRSA), methicillin sensitive *Staphylococcus aureus* (MSSA), *Escherichia coli* O157:H7, and *Salmonella* (Shi Sun *et al.*,2015).

Figure 8: Structure of polybrominated diphenyl ether from sponge *Dysidea granulose* (Reference Shi Sun *et al.*, 2015)

CONCLUSION

Since marine invertebrates are found to produce significant number of natural products and secondary metabolites, isolation of new microbes with potential to produce secondary metabolites from the largely unexplored marine environment requires extensive research. Among them, marine sponges exhibit tremendous capability to produce diverse secondary metabolites. Sponge associated marine microorganisms are the prolific producers of natural compounds with potential against different diseases. These products possess anticancer, antimalarial, antiviral, immunosuppressive properties and other medicinal effects. Plethora of bioactive compounds from the marine life is greatly useful for developing new effective drugs with fewer side effects. In this context, marine sponges offer a promising source for developing new drugs that can be expected to benefit the human kind and the society. In this review, the most important and highly cited reviews for marine sponge bioactive compounds have been included.

REFERENCES

- 1. Alcaraz MJ, Payá M. Marine sponge metabolites for the control of inflammatory diseases. Curr Opin Investig Drugs2006; 7: 974-979.
- 2. Andrea Mencarelli , Claudio D' Amore , Barbara Renga , Sabrina Cipriani , Adriana Carino , Valentina Sepe , Elisa Perissutti , Maria Valeria D'Auria , Angela Zampella , Eleonora Distrutti and Stefano Fiorucci . Solomonsterol A, a Marine Pregnane-X-Receptor Agonist, Attenuates Inflammation and Immune Dysfunction in a Mouse Model of Arthritis. Mar. Drugs 2014; 12(1), 36-53.
- 3. Appadurai Muthamil Iniyan, Thankaraj Rajam Jabila Mary, Francis-Joseph Rosemary Sharmila Joseph, Rajaretinam Rajesh Kannan, Samuel Gnana Prakash Vincent. Cell wall distracting anti-Methicillin-resistant *Staphylococcus aureus* compound PVI331 from a marine sponge associated Streptomyces. Journal of Applied Biomedicine 2016.
- 4. Aubrie O'Rourke, Stephan Kremb, Theresa Maria Bader, Markus Helfer, Philippe Schmitt-Kopplin, William H. Gerwick, Ruth Brack-Werner, Christian R. Voolstra. Alkaloids from the Sponge *Stylissa carteri* Present Prospective Scaffolds for the Inhibition of Human Immunodeficiency Virus 1 (HIV-1). Mar. Drugs 2016; 14(2), 28; doi:10.3390/md14020028.
- 5. Avilés E, Rodríguez AD. Monamphilectine A, a potent antimalarial \hat{I}^2 -lactam from marine sponge *Hymeniacidon sp*: isolation, structure, semisynthesis, and bioactivity. Org Lett 2010;12: 5290-5293.
- 6. Bergmann W., Feeney R.J. Contributions to the study of marine products. XXXII. The nucleosides of sponges I. J. Org. Chem. 1951; 16: 981–987.
- 7. Bergmann W., Feeney R.J. The isolation of a new thymine pentoside from sponges 1. J. Am. Chem. Soc.1950; 72:2809–2810.
- 8. Berrué F, McCulloch MW, Boland P, Hart S, Harper MK, *et al.* Isolation of steroidal glycosides from the Caribbean sponge *Pandaros acanthifolium*. J Nat Prod 2012; 75: 2094-2100.
- 9. Blunt JW, Copp BR, Keyzers RA, Munro MH, Prinsep MR. Marine natural products. Nat Prod Rep 2013; 30: 237-323.
- 10. Chairman K, Jeyamala M, Sankar S, Murugan A, Ranjit Sing. Immunomodulating properties of bioactive compounds present in *Aurora globostellata*. Int J Mar Sci 2013; 3: 151-157.
- 11. Chanthathamrongsiri N, Yuenyongsawad S, Wattanapiromsakul C, Plubrukarn A Bifunctionalized amphilectane diterpenes from the sponge *Stylissa* cf. massa. J Nat Prod 2012; 75: 789-792.

- 12. Cheng Cheng, Eman M. Othman, Anastasija Reimer, Matthias Grüne, Vera Kozjak-Pavlovic, Helga Stopper, Ute Hentschel, Usama R. Abdelmohsen. Ageloline A, new antioxidant and antichlamydial quinolone from the marine sponge-derived bacterium *Streptomyces sp.* SBT345. Tetrahedron letters. 2016; Pages 2786–2789.
- 13. Dhivya Balakrishnan, Arulanandu Sabreen Bibiana, Arumugam Vijayakumar, Ramachandran Sarojini Santhosh, Kandasamy Dhevendaran, Paramasivam Nithyanand. Antioxidant Activity of Bacteria Associated with the Marine Sponge *Tedania anhelans*. Indian J Microbiol 2014.
- 14. Einarsdottir E, Liu HB, Freysdottir J, Gotfredsen CH, Omarsdottir S. Immunomodulatory N-acyl Dopamine Glycosides from the Icelandic Marine Sponge *Myxilla incrustans* Collected at a Hydrothermal Vent Site. Planta Med. 2016 Jun; 82(9-10):903-9. doi: 10.1055/s-0042-105877. Epub 2016 May 2.
- 15. Engelhardt K, Degnes KF, Kemmler M, Bredholt H, Fjaervik E, *et al.* Production of a new thiopeptide antibiotic, TP-1161, by a marine Nocardiopsis species. Appl Environ Microbiol 2010;76: 4969-4976.
- 16. Essack M, Bajic VB, Archer JA. Recently confirmed apoptosis-inducing lead compounds isolated from marine sponge of potential relevance in cancer treatment. Mar Drugs2011; 9: 1580-1606.
- 17. Estelle Sfecci, Thierry Lacour, Philippe Amade, Mohamed Mehiri. Polycyclic Guanidine Alkaloids from Poecilosclerida Marine Sponges. Mar. Drugs 2016; 14(4), 77; doi:10.3390/md14040077.
- 18. Feldman EJ, Lancet JE, Kolitz JE, Ritchie EK, Roboz GJ, *et al*. First-in-man study of CPX-351: a liposomal carrier containing cytarabine and daunorubicin in a fixed 5:1 molar ratio for the treatment of relapsed and refractory acute myeloid leukemia. J Clin Oncol 2011; 29: 979-985.
- 19. Fieseler L, Horn M, Wagner M, Hentschel U.Discovery of the novel candidate phylum "Poribacteria" in marine sponges. Appl Environ Microbiol 2004; Jun; 70(6):3724-32.
- 20. Galeano E, Thomas OP, Robledo S, Munoz D, Martinez A. Antiparasitic bromotyrosine derivatives from the marine sponge *Verongula rigida*. Mar Drugs 2011; 9: 1902-1913.
- 21. Gordaliza M. Cytotoxic terpene quinones from marine sponges. Mar Drugs2010; 8: 2849-2870.
- 22. Gunasekera SP, Cranick S, Longley RE. Immunosuppressive compounds from a deep water marine sponge, Agelas flabelliformis. J Nat Prod 1989:52: 757-761.
- 23. Guzii AG, Makarieva TN, Denisenko VA, Dmitrenok PS, Kuzmich AS, *et al.* Monanchocidin: a new apoptosis-inducing polycyclic guanidine alkaloid from the marine sponge *Monanchora pulchra*. Org Lett 2010; 12: 4292-4295.
- 24. Halim H, Chunhacha P, Suwanborirux K, Chanvorachote P. Anticancer and antimetastatic activities of Renieramycin M, a marine tetrahydroisoquinoline alkaloid, in human non-small cell lung cancer cells. Anticancer Res 2011; 31: 193-201.
- 25. Handbook of Anticancer Drugs from Marine Origin 2014; pp 101-111.
- 26. Ibrahim Majali, Haitham N. Qaralleh1, Syed Z. Idid, Shahbudin Saad, Deny Susanti, Osama Y. Althunibat. Potential Antimicrobial Activity of Marine Sponge Neopetrosia exigua. Journal of Basic and Applied Research.2015; 1-13.
- 27. Jensen PR, Fenical W. Strategies for the discovery of secondary metabolites from marine bacteria: ecological perspectives. Annu Rev Microbiol 1994; 559-84.
- 28. Kalirajan A, Karpakavalli M, Narayanan KR, Ambiganandham K, Ranjit Singh AJA, *et al.* Isolation, characterization and phylogeny of sponge-associated bacteria with antimicrobial and immunomodulatory potential. Int J Curr Microbiol App Sci 2013; 2: 136-151.
- 29. Kennedy J, Baker P, Piper C, Cotter PD, Walsh M, Mooij MJ, Bourke MB, Rea MC, O'Connor PM, Ross RP, Hill C, O'Gara F, Marchesi JR, Dobson ADW. Isolation and analysis of bacteria with antimicrobial activities from the marine sponge *Haliclona simulans* collected from Irish waters. Mar Biotechnol 2009; 11:384–396.
- 30. Laport M., Santos O., Muricy G. Marine sponges: Potential sources of new antimicrobial drugs. Curr. Pharma. Biotechnol 2009;10:86–105
- 31. Lee YK, Lee JH, Lee HK. Microbial symbiosis in marine sponges. J Microbiol 2001; 39:254–264.
- 32.Li, Y.; Liu, D.; Cen, S.; Proksch, P.; Lin, W. Isoindolinone-type alkaloids from the sponge-derived fungus Stachybotrys chartarum. Tetrahedron 2014: 70, 7010–7015.
- 33. Longeon A, Copp BR, Quévrain E, Roué M, Kientz B, *et al.* Bioactive indole derivatives from the South Pacific marine sponges *Rhopaloeides odorabile* and *Hyrtios sp.* Mar Drugs 2011;9: 879-888.

- 34.Ma, X.H.; Lo, L.T.; Zhu, T.J.; Ba, M.Y.; Li, G.Q.; Gu, Q.Q.; Guo, Y.; Li, D.H. Phenylspirodrimanes with Anti-HIV activity from the sponge-derived fungus Stachybotrys chartarum MXH-X73. J. Nat. Prod. 2013, 76, 2298–2306.
- 35. Mayer AM, Glaser KB, Cuevas C, Jacobs RS, Kem W, *et al.* The odyssey of marine pharmaceuticals: A current pipeline perspective. Trends PharmacolSci 2010; 31: 255-265.
- 36. Molinski TF, Dalisay DS, Lievens SL, Saludes JP. Drug development from marine natural products. Nat Rev Drug Discov2009; 8: 69-85.
- 37. Mudianta IW, Skinner-Adams T, Andrews KT, Davis RA, Hadi TA, *et al.* Psammaplysin derivatives from the Balinese marine sponge *Aplysinella strongylata*. J Nat Prod 2012; 75: 2132-2143.
- 38.Oleksii A. Skorokhod, Denise Davalos-Schafler, Valentina Gallo, Elena Valente, Daniela Ulliers, Agata Notarpietro, Giorgia Mandili, "Francesco Novelli, Marco Persico, Orazio Taglialatela-Scafati "Paolo Arese, Evelin Schwarzer. Oxidative stress-mediated antimalarial activity of plakortin, a natural endoperoxide from the tropical sponge *Plakortis simplex*. Free Radical Biology and Medicine 2015; Pages 624–637.
- 39. Pattenden G, Critcher DJ, Remuiñán M. Total synthesis of ()-pateamine A, a novel immunosuppressive agent from *Mycale sp.* Can J Chem 2004; 82: 353–365.
- 40. Pazhanimuthu Annamalai, Malini Thayman, Sowmiya Rajan, Lakshmi Sundaram Raman, Sankar Ramasubbu, Pachiappan Perumal. Ethyl acetate extract from marine sponge *Hyattella cribriformis* exhibit potent anticancer activity by promoting tubulin polymerization as evidenced mitotic arrest and induction of apoptosis. Pharmacogn Mag 2015; Apr-Jun; 11(42): 345–355.
- 41. Peter Sima, Vaclav Vetvicka. Bioactive substances with anti-neoplastic efficacy from marine invertebrates: Porifera and Coelenterata. World J Clin Oncol 2011; Nov 10; 2(11): 355–361.
- 42. Proksch P, Edrada RA, Ebel R. Drugs from the seas current status and microbiological implications. Appl Microbiol Biotechnol 2002; 59: 125-134.
- 43. Rosa Lemmens-Gruber. Antitumour Effect of Cyclodepsipeptides from Marine Sponges. 2014.
- 44. Santiago Bastos JC, Konecny Kohn L, Fantinatti-Garboggini F, Aiello Padilla M, Furtado Flores E, *et al.* Antiviral Activity of Bacillus sp. Isolated from the Marine sponge *Petromica citrina* against bovine viral diarrhea virus a surrogate model of the hepatitis C virus. Viruses 2013; 1219-1230.
- 45. Sergey A. Dyshlovoy ,Kseniya M. Tabakmakher, Jessica Hauschild, Regina K. Shchekaleva, Katharina Otte, Alla G. Guzii, Tatyana N. Makarieva, Ekaterina K. Kudryashova, Sergey N. Fedorov, Larisa K. Shubina, Carsten Bokemeyer, Friedemann Honecker, Valentin A. Stonik, Gunhild von Amsberg. Guanidine Alkaloids from the Marine Sponge *Monanchora pulchra* Show Cytotoxic Properties and Prevent EGF-Induced Neoplastic Transformation in Vitro. Mar. Drugs 2016; 14(7), 133; doi: 10.3390/md14070133.
- 46. Shaaban M, Abd-Alla HI, Hassan AZ, Aly HF, Ghani MA. Chemical characterization, antioxidant and inhibitory effects of some marine sponges against carbohydrate metabolizing enzymes. Org Med Chem Lett 2012; 2: 30.
- 47. Shi Sun, Corene B. Canning, Kanika Bhargava, Xiuxiu Sun, Wenjun Zhu, Ninghui Zhou, Yifan Zhang, Kequan Zhou. Polybrominated diphenyl ethers with potent and broad spectrum antimicrobial activity from the marine sponge *Dysidea*. Bioorganic & Medicinal Chemistry Letters 2015; Pages 2181–2183.
- 48. Shen S, Liu D, Wei C, Proksch P, Lin W. Purpuroines A-J, halogenated alkaloids from the sponge Iotrochota purpurea with antibiotic activity and regulation of tyrosine kinases. Bioorg Med Chem 2012; 6924-6928.
- 49. Simmons TL, Andrianasolo E, McPhail K, Flatt P, Gerwick WH.Marine natural products as anticancer drugs. Mol Cancer Ther 2005; 4: 333-342.
- 50.Siti Aisha Mohd Radzia, Yosie Andriania, Habsah M.a, Tengku Sifzizul Tengku Mohamada, Jasnizat Saidina,b. In-vitro anti-inflammatory activities of extracts from bacteria associated with marine sponges. Theonella sp. Jurnal Teknologi 2015.
- 51.SM Al-Massarani, AA El-Gamal, MS Al-Said, SS Al-Lihaibi, OA Basoudan. In vitro Cytotoxic, Antibacterial and Antiviral Activities of Triterpenes from the Red Sea Sponge, *Siphonochalina siphonella*. Tropical Journal of Pharmaceutical Research 2015.
- 52. Taylor MW, Radax R, Steger D, Wagner M. Sponge-associated microorganisms: evolution, ecology, and biotechnological potential. Microbiol Mol Biol Rev 2007; 71(2):295-347.

- 53. Valeriote FA, Tenney K, Media J, Pietraszkiewicz H, Edelstein M, *et al.* Discovery and development of anticancer agents from marine sponges: perspectives based on a chemistry-experimental therapeutics collaborative program. J Exp Ther Oncol 2012; 10: 119-134.
- 54. Villa FA, Gerwick L. Marine natural product drug discovery: Leads for treatment of inflammation, cancer, infections, and neurological disorders. Immunopharmacol Immunotoxicol 2010; 32: 228-237.
- 55. Wang G. Diversity and biotechnological potential of the sponge-associated microbial consortia. J Ind Microbiol Biotechnol. 2006; 33:545–551.
- 56. Wei-Hua Jiao, Guo-Hua Shi, Ting-Ting Xu, Guo-Dong Chen, Bin-Bin Gu, Zhuo Wang, Shuang Peng,Shu-Ping Wang, Jia Li, Bing-Nan Han, Wei Zhang, Hou-Wen Lin. Dysiherbols A–C and Dysideanone E, Cytotoxic and NF-κB Inhibitory Tetracyclic Meroterpenes from a Dysidea sp. Marine Sponge. J. Nat. Prod 2016;79 (2), pp 406–411.
- 57. Woori Bae, Hyun Kyung Lim, Kyoung Mee Kim, Hyosun Cho,Sun Yi Lee, Choon-Sik Jeong,Hyi-Seung Lee, Joohee Jung. Apoptosis-Inducing Activity of Marine Sponge Haliclona sp. Extracts Collected from Kosrae in Nonsmall Cell Lung Cancer A549 Cells. Evidence-Based Complementary and Alternative Medicine.Volume 2015; Article ID 717959, 8 pageshttp://dx.doi.org/10.1155/2015/717959.
- 58. Wright AD, McCluskey A, Robertson MJ, MacGregor KA, Gordon CP, *et al.* Anti-malarial, anti-algal, anti-tubercular, anti-bacterial, anti-photosynthetic, and anti-fouling activity of diterpene and diterpene isonitriles from the tropical marine sponge *Cymbastela hooperi*. Org Biomol Chem 2011; 9: 400-407.
- 59. Yang F, Hamann MT, Zou Y, Zhang MY, Gong XB, *et al.* Antimicrobial metabolites from the Paracel Islands sponge *Agelas mauritiana*. J Nat Prod 2012; 75: 774-778.
- 60. Yong-Qi Tian ,Xiu-Ping Lin, Zhen Wang, Xue-Feng Zhou, Xiao-Chu Qin, Kumaravel Kaliyaperumal, Tian-Yu Zhang, Zheng-Chao Tu, Yonghong Liu. Asteltoxins with Antiviral Activities from the Marine Sponge-Derived Fungus *Aspergillus sp.* SCSIO XWS02F40. Molecules 2016; 21(1), 34; doi:10.3390/molecules21010034.
- 61. Yun-Ji Seo, Kyung-Tae Lee, Jung-Rae Rho, Jung-Hye Choi. Phorbaketal A, Isolated from the Marine Sponge Phorbas sp., Exerts Its Anti-Inflammatory Effects via NF-κB Inhibition and Heme Oxygenase-1 Activation in Lipopolysaccharide-Stimulated Macrophages 2015; Mar. Drugs, 13(11), 7005-7019.

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