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Preliminary Phytochemical and GC-MS Profiling of Ethanolic Extract of Leaves of *Calotropis gigantea Linn*. (Asclepiadaceae)



Beena Thomas, A.J Chacko, Reshma Thampy*

Department Of Pharmaceutical Sciences, Centre For Professional and Advanced Studies Kottayam, Kerala, India

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ABSTRACT

Calotropis belong to the Asclepiadaceae family. It is also known as Akada, Aak, Mandar, Aakh etc. It has two species procera and gigantea. Here we study about Calotropis gigantea. The roots and leaves of Calotropis gigantea are used traditionally for the treatment of abdominal, tumors, skin diseases, wound, insect bites. The leaves of the plant contain primary and secondary metabolites. Quantitative analysis showed in the presence of alkaloids, glycosides, terpenoids, saponins, cardiac glycosides, flavonoids. Calotropis gigantea in a small dose is also useful in the treatment of cold, cough, asthma inflammatory diseases and loss of digestive and analgesic property. The medicinal properties of this plant represent it as a valuable source of a medicinal compound. This summarizes that information concerning phytochemical constituent present in the ethanolic extract of Calotropis gigantea in these constituents may be responsible for the pharmacological activity.

INTRODUCTION

Herbals plants are an effective source of traditional and modern medicines, used for primary health care. Plants are the richest source of bioactive organic chemicals on earth. The active metabolites like Phytochemicals from the medicinal plants were under exploration for the development of novel and biodegradable effective drugs as an alternative to the ineffective contemporary medicine. Calotropis gigantea has great medicinal importance to treat indigestion, rheumatism, diabetes, cancer cold, cough, cardiotonic, asthma, scabies etc^[1]. The most important of these chemically active constituents of plants are alkaloids, tannin, flavonoid and phenolic compounds. Many of these indigenous medicinal plants are also used for medicinal purposes [2, 3]. Natural products, which come out from medicinal plants are important for pharmaceutical research and for drug development as a source of therapeutic agents. At presents, the demand for herbal or medicinal plant products has increased significantly. In the recent past, there has been a growing interest in exploiting the biological activities of different Ayurvedic medicinal herbs, owing to their natural origin, costeffectiveness and lesser side effects [4]. Medicinal plants are an expensive gift from nature to human. The approval of traditional medicine as an alternative form of health care and the improvement of microbial resistance to the existing antibiotics has lead researchers to scrutinize the antimicrobial compounds ^[5].

Nowadays synthetic drugs are very expensive to develop and whose cost of development ranges from 0.5 to 5 million dollars. In contradiction to this many medicines of the plant, origin had been used for a long time without any adverse effects. Exploring the healing power of plants is an ancient concept. For centuries, people have been trying to alleviate and treat disease with different plant extracts and formulations ^[6, 7]. Because of the present situation, there is a need for essential efforts that should be made to introduce new medicinal plants to develop cheaper drugs. Plants still represent a largely untapped source of structurally novel compounds that might serve as the lead for the development of novel drugs ^[8]. Gas chromatography-mass spectrometry (GC-MS) is a method that combines the features of gas-liquid chromatography and mass spectrometry to identify different substances within a test sample. Applications of GC-MS include drug detection, fire investigation, environmental analysis, explosives investigation, and identification of unknown samples. GC-MS can also be used in airport security to detect substances in luggage or on human beings. However, fewer reports are available with respect to the pharmacological properties of the plant ^[9].

Keeping this in view, the present study has been undertaken to investigate the phytochemical

constituents present in the ethanol extract of Calotropis gigantea leaf.

MATERIALS AND METHODS

Collection of medicinal plant

The Indian medicinal plant Calotropis gigantea was collected from the medicinal garden of

DPS CPAS Puthuppally Kottayam., Kerala, India.

The plant was authenticated by Rogimon P.Thomas, Assistant professor Department of

Botany, CMS College Kottayam (Kerala).

Preparation of Plant extract

The ethanolic extract of leaves of *Calotropis gigantea* was used in the study. The leaves were

separated, freed from adhering moisture, dried in sunshade and powdered. The powdered

material (32gm)^[10] was packed in soxhlet apparatus and extraction was done using 450 ml of

ethanol (60-70°C) 56 hours. The extracts were filtered using filter paper (No.1) while hot,

concentrated in vacuum under reduced pressure using a rotary flask evaporator, and dried in a

vacuum. The ethanolic extract yielded a dark greenish solid residue. The extract was then

kept in the sterile bottle, under refrigerated conditions, until further use. The dry weight of the

plant extract was obtained by the solvent evaporation and used to determine the concentration

in mg/ml. The extract was preserved at 2 to 4° and sent an aliquot quantity for GC-MS

analysis.

PHYTOCHEMICAL ANALYSIS OF THE EXTRACT:

A small portion of the dry extracts was subjected to the phytochemical test using Harbourne's

(1983) methods to test for alkaloids, tannins, terpenoids, steroids, flavonoids, and glycosides.

1. Test for alkaloids:

About 0.2 g extract warmed with 2% H₂SO₄ for two minutes, filtered and few drops of

Dragendroff's reagent added orange-red precipitate indicates the presence of alkaloids. And

or filtrates were treated with Wagner's reagent (Iodine in Potassium Iodide). Formation of

brown/reddish precipitate indicates the presence of alkaloids.

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2. Test for glycosides: The extracts hydrolyzed with HCl solutions and neutralized with

NaOH solutions. A few drops of Fehling solution A and B were added. Red precipitate

indicates the presence of glycoside. Another test used was Benedict's test, in which the

filtrates were treated with Benedict's reagent and heated gently. Orange-red precipitate

indicates the presence of reducing sugars.

3. Test for tannins: Small quantity of extracts mixed with water, heated, filtered and ferric

chloride added. A dark green solution indicates the presence of tannins.

4. Test for steroids: 2ml of the extract was taken in a test tube and 2 ml of chloroform and 2

ml of concentrated H₂SO₄ was added, the formation of the reddish-brown ring at the junction

of two liquids, shows the presence of steroids

5. Test for flavonoids: Extract of about 0.2 g of the extracts shaken with 5ml of distilled

water and then a few drops of 10% lead acetate solution is added. A yellow or dirty white

precipitate shows the presence of flavonoids. [11]

RESULTS AND DISCUSSION

The preliminary phytochemical screening of the ethanolic extract showed the presence of

alkaloids, glycosides, tannins, steroids, flavonoids.

GC-MS Profiling

Working procedure:

Central instrument unit Thrissur Kerala carried out GC-MS

One-microliter sample was subjected for the study. The Instrument used, Varian, CP-3800

Saturn 2200 GC/MS/MS with factor four VF-5MFcolumn.Oven temperature maintained at

1000C for 1.5 minutes, the temperature gradually increased to 2700 c at 50 c per minute, and

1µlitre sample was injected for analysis. Helium gas 99.9 % was used as the carrier gas, the

flow rate of carrier gas was1ml per minute sample injected temperature was maintained at

250° C and the split ratio is 20 throughout the experiment period. The ionization mass was

done with 70 eV. The mass spectra were recorded for the mass range 40-600 m/z for 60

minutes. Identification of compound was based on a comparison of their mass spectra. As the

compound separated, on elution through column were detected in electronic signals. The m/z

obtained was calibrated through graph obtained, which was called the mass spectrum graph, which is the fingerprint of the molecule. The identification of the compound was based on the comparisons of their mass spectra with Nist library.

Applications of GC -MS

- 1) Very minute amounts of a substance can be measured.
- 2) Various temperature programs can be used to make the readings more meaningful; for example to differentiate between substances that behave similarly during the GC process.
- 3) Gas Chromatography is used in the separation and analysis of multi-component mixtures such as essential oils, hydrocarbons, and solvents.
- 4) Intrinsically, with the use of the flame ionization detector and the electron capture detector (which have very high sensitivities) gas chromatography can quantitatively determine materials present at very low concentrations.
- 5) The most important application area is in pollution studies, forensic work and general trace analysis. [12]

GC-MS chromatogram (Calotropis gigantea)

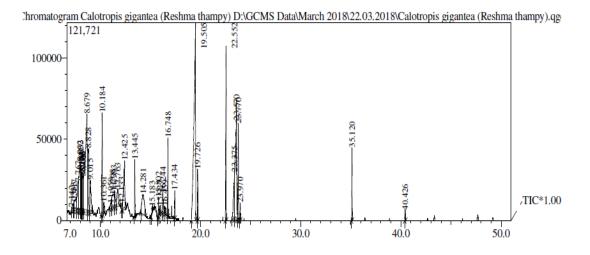


Table number: 1

Peak Report

| Peak | R. time | Area | Area % | Height | Height | Name | Base m/z |
|------|------------|--------|-----------|--------|--------|---|-------------|
| 1 | 7.142 | 27160 | 0.40 | 1628 | 0.15 | 1-BUTANOL | 56.00 |
| 2 | 7.333 | 16666 | 0.25 | 4131 | 0.38 | 1-PYRROLIN-N-OXIDE | 85.05 |
| 3 | 7.417 | 108879 | 1.61 | 6149 | 0.56 | PENTANE, 2,2-DIMETHYL | 57.05 |
| 4 | 7.767 | 177458 | 2.62 | 17849 | 1.63 | 1,3-DIOXOLANE,2,4,5- TRIMETHYL- | 101.00 |
| 5 | 8.002 | 273175 | 4.03 | 30439 | 2.78 | N-(tert-butyl)UREA | 58.00 |
| 6 | 8.033 | 138157 | 2.04 | 30526 | 2.79 | CYCLOBUTANONE,2 METHYL-4-HYDROXY | 58.00 |
| 7 | 8.108 | 48861 | 0.72 | 25623 | 2.34 | PROPANENITRILE, 3- (METHYLTHIO)- | 61.00 |
| 8 | 8.142 | 35190 | 0.52 | 24267 | 2.22 | GLYCERALDEHYDE | 61.00 |
| 9 | 8.175 | 46881 | 0.69 | 23750 | 2.17 | PENTABORANE(11) | 62.05 |
| 10 | 8.206 | 66295 | 0.98 | 24256 | 2.22 | ETHYNE, DIFLUORO | 62.00 |
| 11 | 8.300 | 154202 | 2.28 | 22008 | 2.01 | D1-METHANAMINE, N- METHOXY- | 62.05 |
| 12 | 8.679 | 890088 | 13.14 | 60500 | 5.53 | 2-OXOPENTANEDIOIC SCID | 101.00 |
| 13 | 8.828 | 273761 | 4.04 | 39379 | 3.60 | GLYCERIN | 61.00 |
| 14 | 9.015 | 146089 | 2.16 | 20159 | 1.84 | BENZOFURAN, 2,3-DIHYDRO | 120.05 |
| 15 | 10.184 | 227746 | 3.36 | 63068 | 5.77 | 2-METHOXY-4-VINYLPHENOL | 150.05 |
| 16 | 10.361 | 38099 | 0.56 | 7435 | 0.68 | L-LYXONIC ACID, 5-DEOXY-, GAMMALACTONE | 70.05 |
| 17 | 11.050 | 32751 | 0.48 | 3069 | 0.28 | BUTANE,2-NITRO | 57.00 |
| 18 | 11.208 | 109592 | 1.62 | 8004 | 0.73 | OXIRANE,(BUTOXYMETHYL)- | 57.00 |
| 19 | 11.393 | 83414 | 1.23 | 10191 | 0.93 | ACETALDEHYDE, SEMICARBAZONE | 60.00 |
| 20 | 11.763 | 119017 | 1.76 | 10769 | 0.99 | PROPANENITRILE,2,2- DIMETHYL | 84.05 |
| 21 | 12.133 | 21667 | 0.32 | 4524 | 0.41 | NITROUSACID, BUTYL ESTER | 60.00 |
| 22 | 12.425 | 280763 | 4.14 | 30293 | 2.77 | 2-PIPERIDINEMETHANOL, ALPHA-ETHYL- | 84.05 |
| 23 | 13.445 | 84311 | 1.24 | 35420 | 3.24 | 3',5'- DIMETHYLACETOPHENONE | 180.00 |
| 24 | 14.281 | 20989 | 3.10 | 12468 | 1.14 | BETA-D- GLUCOPYRANOSE,1,6 | 60.00 |

| | | | | | | ANHYDRO | |
|----|--------|---------|--------|---------|--------|--|--------|
| 25 | 15.183 | 12852 | 0.19 | 3867 | 0.35 | 2-PENTENE,3-METHYL-,(Z)- | 69.05 |
| 26 | 15.802 | 32054 | 0.47 | 12307 | 1.13 | NONANOIC ACID | 73.00 |
| 27 | 15.927 | 13413 | 0.20 | 4827 | 0.44 | TRIPROPYLBORANE | 97.05 |
| 28 | 16.244 | 64886 | 0.96 | 12539 | 1.15 | 2,6,6-TRIMETHYL- BICYCLO(3,1,1)HEPT-3- YLAMINE | 98.05 |
| 29 | 16.459 | 29042 | 0.43 | 5342 | 0.49 | DIMETHYL(3-TERT- BUTYLACETOXY)-3- METHYL-2- OXOBUTYL)PHOSPHONATE(1 8)O | 124.05 |
| 30 | 16.748 | 87956 | 1.30 | 48324 | 4.42 | NEOPHYTADIENE | 68.05 |
| 31 | 17.434 | 29558 | 0.44 | 17436 | 1.60 | 3-NONEN-1-OL,(Z) | 55.00 |
| 32 | 19.505 | 133430 | 19.70 | 121721 | 11.14 | HEXADECAOIC ACID | 73.00 |
| 33 | 19.726 | 75720 | 1.12 | 31585 | 2.89 | HEXADECANOIC ACID, ETHYL ESTER | 88.05 |
| 34 | 22.552 | 342102 | 5.05 | 107105 | 9.80 | PHYTOL | 71.05 |
| 35 | 23.357 | 175223 | 2.59 | 29246 | 2.68 | 1,1'-BICYCLOPENTYL | 67.05 |
| 36 | 23.570 | 621755 | 9.18 | 61313 | 5.61 | 9,12,15-OCTADECATRIENAL | 79.05 |
| 37 | 23.770 | 190371 | 2.81 | 59451 | 5.44 | 9,12,15-OCTADECATRIENOIC ACID(Z,Z,Z)- | 79.05 |
| 38 | 23.970 | 35177 | 0.52 | 10779 | 0.99 | PENTACHLOROBROMOBENZ ENE | 55.05 |
| 39 | 35.120 | 97074 | 1.43 | 44867 | 4.10 | UNKNOWN TERPENE IN GRAPES | 69.05 |
| 40 | 40.426 | 22345 | 0.33 | 6451 | 0.59 | DI-ALPHA-TOCOPHEROL | 165.05 |
| | | 6773775 | 100.00 | 1093065 | 100.00 | | |

Activity of bioactive compounds identified in ethanolic extract of *Calotropis* gigantea^[13,14]

Table number: 2

| Serial number | Compound name | Activity | | |
|------------------|--|---|--|--|
| 1 | Hexadecanoic acid, Ethyl ester | Hemolytic, Antioxidant, Flavor, Alpha reductase inhibitor | | |
| 2 | Phytol | Anti-inflammatory, Anticholesteremic | | |
| 3 | 9,12,15 – Octadecatrienoic acid, (z,z,z) | Anti-arthritic, Anti-inflammatory, Anti eczemic, Anti acne, Cancer preventive | | |
| 4 | di-alpha-Tocopherol | Vitamin E | | |
| 5 | Pentachlorobromobenzene | Inflammation, Wounds, Psoriasis | | |
| 6 | 1,1-Bicyclopentyl | Eczema, Psoriasis, Seborrheoeic dermatitis | | |
| 7 | 3-Nonen-1-ol,(z)- | Murine leukemia | | |
| 8 | Acetaldehyde semicarbazone | Antiviral, Anticancer | | |
| 9 | Neophytadiene | Antiseptic, Anti-inflammatory. | | |
| 10 | Beta-D-Glucopyranose, 1,6-anhydro | Anti-microbial | | |
| 11 | Nonionic acid | Anti-microbial | | |
| 12 | Oxirane, (butoxymethyl)- | Anti-viral | | |
| 13 | Benzofuran,2,3-dihydro | Anti-inflammatory, Anti-tumor, Anti- coagulant, Alzimers disease inhibitor | | |
| 14 | 2-oxopentanedioic acid | Immunomodulator | | |
| 15 | 2-Methoxy-4-vinylphenol | Anti-cancer, Anti- diabetic, Anti-oxidant | | |
| 16 | Unknown terpenes | Anti-inflammatory, Expectorant | | |

GC -MS analysis has been found to be an ideal technique for the analysis of volatile and semi-volatile bioactive compounds. The total number of the compound identified in the ethanolic extract was 40. The GC-MS retention time and percentage peak of the individual compounds were demonstrated. It contains terpenoids, saponins, flavonoids, alkaloids, steroids. The major Phytoconstituents present in ethanolic extract was 9,12,15 –

octadecatrienoic acid(z,z,z)- retention time 23.570 peak area 9.18%, Hexadecanoic acid, Ethyl ester retention time 19.505 peak area 19.70%, Phytol retention time 22.552 peak area 5.05%, Unknown terpenes retention time 35.120 peak area 1.43%, Oxirane (butoxymethyl)-retention time 11.208 peak area 1.62%, di-alpha-Tocopherol retention time 40.426 peak area 0.33%, Pentachlorobromobenzene retention time 23.970 peak area 0.52%, Neophytadiene retention time 16.748, peak area 1.30%.

CONCLUSION

Now a day the identification of bioactive compounds from the medicinal plant has increased. A large number of medicinal plants and their purified constituents have shown beneficial therapeutic potentials. The ethanolic extract of *Calotropis gigantea* frequently used as a natural remedy for many diseases. The identification of this study is based on the peak area of the compound (which represents the percentage of that compound). In the present study, 40 compounds were identified from leaves extract of *Calotropis gigantea* using ethanol as a solvent. However, the isolation of individual phytochemical constituents may proceed to find a novel drug. Therefore, the GC-MS method is a direct and fast analytical approach for identification of terpenoids, glycosides, alkaloids, and steroids and only a few grams of plant material is required. The results reveal that the extract has a quite number of chemical constituents, which may be responsible for many pharmacological activities. Further studies are needed on this extracts in order to isolate, identify, characterize and elucidate the structure of these compound.

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