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An Observational Study to Determine the Organism/ Organisms That Colonize the Central Lines and To Study the Sensitivity of Those Organisms



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ABSTRACT

Aims and objectives: This is the first time that 100% of central lines inserted in a hospital setup have been studied for colonization. This study aimed to determine organism/organisms that colonize the central lines, to study the temporal relationship of the identification of the colonizers, and to study the relationship to variables like antibiotic use and Acute Physiology and Chronic Health Evaluation II score. Materials and methods: This is a prospective observational study conducted on 200 patients in whom central lines were inserted at S. L. Raheja Hospital (A Fortis Associate), Mumbai. Tips of all central lines that were removed were cultured. Simultaneous blood cultures were also done. Patient demographics, Acute Physiology and Chronic Health Evaluation II score scores, and length of stay were noted to ascertain the relationship between severity of illness, length of stay in ICU, and colonization of central line. Results: The rate of colonization as per the results of our study was 29%. Grampositive organisms make 43.1%, gram-negative 41.7%, and fungus in 13.7% of cases. There was no relationship to severity of illness, duration of the length of central line days, or comorbidities with the rate of colonization. Conclusion: Very few studies exist regarding the colonization rate of central lines. The data of true colonization rate from our single-center reveal that central lines do get colonized with gram-positive and gram-negative organisms in almost equal proportion. This data also seems to suggest that comorbidities, the severity of illness, or the length of time that the catheter is in situ does not change the colonization rate. This data may thus help us to make protocols for clinical management and in devising materials for making catheters.

INTRODUCTION

Intravascular catheters are frequently used in modern-day medical practice mainly to assess and monitor critically ill patients. It is used primarily to monitor central venous pressure or to administer fluid, medications, or total parenteral nutrition, or obtain blood samples.

Central venous catheters (CVC) act as a portal of entry of bacteria that colonize the skin adjacent to the entry point or may serve as foreign bodies that harbour micro colonies leading to multiple complications¹.

Approximately 87% of bloodstream infections are associated with the presence of some type of intravascular device². Catheter-Related Blood-Stream Infections (CRBSI) is the most life-threatening of all healthcare-acquired infections².

The use of central venous catheters puts patients at risk for local and systemic infectious complications. It includes local site infection, CRBSI, septic thrombophlebitis, endocarditis, and other metastatic infections (e.g., lung abscess, brain abscess, osteomyelitis, and endophthalmitis)³.

Infections associated with the use of intravascular catheters represent around 10-20% of all nosocomial infections and cause substantial morbidity and mortality^{4;5}. More than 250,000 intravascular catheter-related bacteremias and fungemia occur annually in developed countries like the USA with an attributable mortality of 12-25%⁴.

Microbial contamination leads to the development of complex fungal or bacterial biofilm communities which are a potential source of BSI⁶. The most problematic feature of mature biofilms in case of catheter-related infection is an increased ability of biofilm cells to survive antimicrobial agents and the host immune system⁷.

Central venous catheter infections most commonly occur in one of three ways: colonization of the catheter tip during insertion, contamination at the catheter hub with routine use, and infection from another source within the body that spreads through the bloodstream and attaches to the lumen. Rarely CVCs can become infected from contaminated infusions⁸.

CVC contamination causes an increase in hospitalization costs, morbidity, mortality, and duration of hospitalization; thus, prevention of these infections can be effective in reducing these outcomes⁹.

CVC-related bloodstream infections also prolong hospitalization by an average of 6.5 days¹⁰.

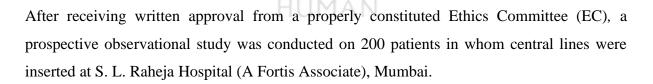
Increased mortality rates may also be attributed to the effects of CVC infections. Probably the most significant problem created by a central venous catheter infection is the negative impact one can have on a patient's quality of life and leads to an economical burden¹¹.

To overcome these problems, there is a need for a study regarding colonization patterns and factors affecting colonization.

Furthermore, determining which antibiotic is appropriate for the treatment of a CRBSI can be quite difficult. Antibiotic resistance is becoming increasingly more common creating a barrier to successful eradication of the organism causing the infection. Hence, studying the antibiotic sensitivity profile of the particular organism will decrease morbidity resulting in better patient management.

Thus, data obtained from the study will have a huge impact on the future both on patient management and guiding the catheter manufacturing industry, particularly in the Indian scenario.

MATERIALS AND METHODS



Inclusion criteria:

1. Patients older than 14 years, in whom Central line (multi-lumen/single /Hemodialysis line/jugular sheath insertion) needs to be removed unrelated to the central line-associated infection.

2. Those patients where there was suspicion of Central Line-Associated Blood Stream Infections (CLABSI), and the line is removed for that purpose.

3. All patients with other sources of infection who had central lines inserted.

Exclusion criteria:

1. Patients who were admitted to SL Raheja hospital with central lines inserted outside.

2. Central lines accidentally removed,

3. Proven cases of CLABSI and/or cases in which the central line bundle was not followed were excluded.

Central line insertion had been carried out by the trained ICU registrar after all aseptic precautions and has been audited by infection control nurse/assistants. The central line had been removed by the researcher himself for this purpose.

At the time of removal of the central line, the terminal 5 cm of the central line had been cut and taken for culture using an aseptic technique and put in a sterile container. The labelling of the container included giving a serial number from 1 to 200. It had been written down in the format "CLS/2017/SLRH/Sr. no.' with the hospital ID of the patient and the patient initials. Then the sample was transferred to the microbiology laboratory by the infection control nurse and personally handed over to the microbiology technician to ensure processing within 2 hours of removal of line.

In CLABSI suspected cases as per standard protocol, simultaneous central line and peripheral blood cultures were taken and catheters were not removed for such purposes.^{3, 16}

Cultures had been processed as per standard methods and observations had been done by both Maki's method and subculture method of the lumen. Then the colony counts were determined.³ the readings and interpretation had been taken by the microbiologist. Processing had been done by microbiology technicians under the guidance of a technical supervisor.

The growth of ≥ 1 microorganism in a quantitative or semiquantitative culture of the catheter tip, had been defined as colonization.³ All organisms isolated had been tested for sensitivity to study epidemiology. Organism isolation and susceptibility testing had been done as per CLSI (Clinical And Laboratory Standard Institute) guidelines 2017. The disposal and sample retention policy of the study material has been as per SLRH Laboratory policy. Statistical analysis was carried out by using IBM SPSS software version 2015.

Ethical policies: Hospital Ethics committee approval was taken before the start of the study.

Ethical Guidelines: NA

Humane considerations: - NA

Conflict of interests-NIL

Funding - NIL

RESULTS

The study involved a total of 200 patients of which 125 (62.5%) of the cases were males and 75 (37.5%) were females. The ages of the cases were ranging from 15.00–91.00 years with the average age being 60.79 years. The mean Acute Physiology and Chronic Health Evaluation II score (APACHE) score was 13.57. The mean duration of the central line in situ was 9.66 days. (as shown in Table 1)

Table No. 1: 1Demographical Data

Parameters	
No. of Cases	200
Age (yrs)	60.79
Mean SD	15.44
Range	15.00 – 91.00 yrs
Sex (%)	125(62.5)
Male Female	075(37.5)
Mean APACHE Score	13.57
Mean Duration of Central Line in Situ	9.66

The total no. of cases with catheter colonization was 58(29%). Out of which 24% patients showed growth in Makis method and 26% patients showed growth in the subculture method (Table 2).

The analysis shown in Table 3 and Graph 1 states that 27.7% of the cases had colonization that belongs to Apache Score < 15 which was comparable with 31.4% of the cases with Apache Score >15. 28.2% of the cases had colonization that belongs to Nutrition Risk Score (NRS 2002) < 4 which was comparable with 30.0% of the cases with NRS (2002) > 4 (Table 4, Graph 2). 28.5% of the cases had colonization that belongs to Nutric Score < 5 which was comparable with 31.4% of the cases with Nutric Score >5(Table 5, Graph 3). 42.9% of the cases had colonization that belongs to Nutric Score as compared to 29.1% of the cases with Leucocytes (> 11000) (Table 6, Graph 4). According to Table 7 and

Graph 5, 23.1% of the cases had colonization that belongs to Duration of Central Line < 7 which was less as compared to 33.9% of the cases with Duration of Central Line > 7.

However, the differences observed between the two groups studies among the above-stated variables as the APACHE score, NRS, Nutric Score, Leucocyte status, duration of the central line in situ were not found statistically significant.

Table No. 2: Profile of Organisms Colonized among Study Cases

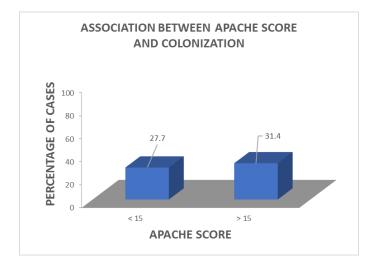
Organisms	No. of Cases (N = 200)	Percentage
Total No. of Cases with Colonized	58	29.0
Makis	48	24.0
Subculture	26	13.0

Table No. 3: Association Between Apache Score and Colonization

Apache Score	No. of Cases	Cases with colonization No. %
< 15	130	36 27.7
> 15	070	22 31.4

By Chi Square Test P > 0.05 Not Significant

Graph 1



NRS (2002)	No. of Cases	Cases with colonization No. %
<u><</u> 4	110	31 28.2
> 4	090	27 30.0

Table No. 4: Association Between Nrs (2002) and Colonization

By Chi Square Test P > 0.05 Not Significant

Graph 2

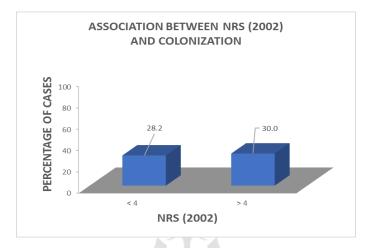
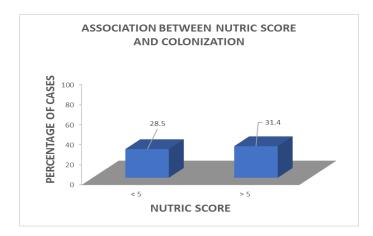


Table No. 5: Association Between Nutric Score and Colonization

Nutric Score	No. of Cases	Cases with colonization No. %
< 5	165	47 28.5
> 5	035	11 31.4

By Chi Square Test P > 0.05 Not Significant

Graph 3



WBC Count	No. of Cases	Cases withColonization No. %
Neutropenia (≤ 4000)	007	03 42.9
Normal (4000 – 11000)	081	21 25.9
Leucocytes (≥ 11000)	110	32 29.1

Table No. 6: Association Between Wbc Count and Colonization

By Chi Square Test P > 0.05 Not Significan

Graph 4

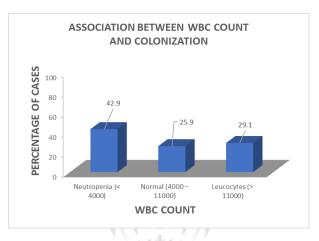
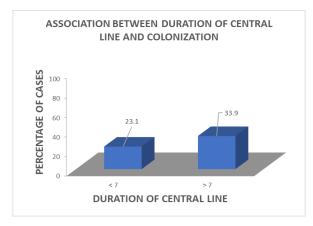


Table No. 7: Association Between Duration of Central Line and Colonization

Duration of central line	HUMAN No. of Cases	Cases with colonization No. %
≤7	091	21 23.1
> 7	109	37 33.9

By Chi Square Test P > 0.05 Not Significant

Graph 5

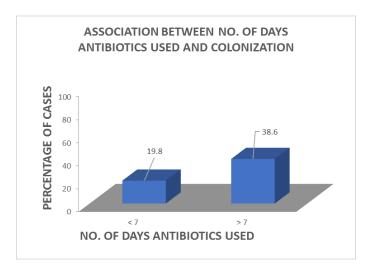


According to Table 8 and Graph 6, 19.8% of the cases that had colonization used Antibiotics for < 7 days which was significantly less as compared to 38.6% of the cases that used Antibiotics > 7 days.

No. of Days Antibiotics Used	No. of Cases	Cases with colonization No. %
<u><</u> 7	096	*19 19.8
> 7	101	39 38.6

By Chi Square Test *P < 0.05Significant

Graph 6



Out of 200 processed catheters, 58 of them showed significant growth. Out of which 27 showed growth of gram-positive organisms, 24 showed growth of gram-negative organisms, 9 cases showed fungal growth.

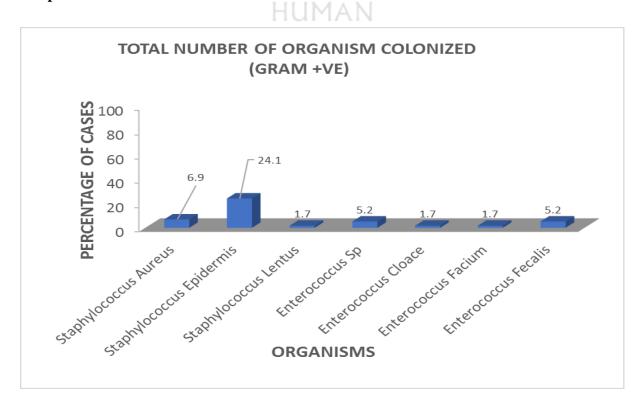
Among gram-positive organisms, the most observed growth was Staphylococcus Epidermis (24.1%) followed by Staphylococcus Aureus (6.9%) followed by Enterococcus Faecalis (5.2%). The growth of all three organisms i.e. Enterococcus Cloacae and Enterococcus Faecium and Staphylococcus Lentus were reported to be 1.7%. Other Enterococcal species showed a total growth of 5.2% (Table 9, Graph 7). Among gram-negative organisms, Pseudomonas and Klebsiella pneumonia showed maximum growth in 13.8% cases each. Serratia and Acinetobacter showed growth in 5.2% and 3.4% cases respectively. Proteus

species, Enterobacter aerogenes, and Burkholderiacepecia showed growth in 1.7% cases each (Table 10, Graph 8). Most common fungus grown was Candida Albicans (6.9%) followed by Candida tropicalis (3.4%). Candida Auris and Candida ciferri showed growth of 1.7% cases each. (Table 11, Graph 9)

Table No. 9: Total Number of Organism Colonized (Gram +Ve	al Number of Organism Colonize	d ($Gram + Ve$)
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Organisms	No. of Cases Colonized (N = 58)		
	No. of Cases	Percentage	
Staphylococcus Aureus	04	06.9	
Staphylococcus Epidermis	14	24.1	
Staphylococcus Lentus	01	01.7	
Enterococcus Sp.	03	05.2	
Enterococcus Cloacae	01	01.7	
Enterococcus Faecium	01	01.7	
Enterococcus Faecalis	03	05.2	

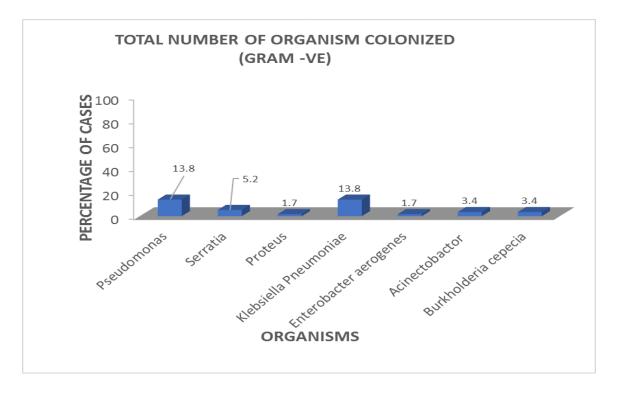
Graph 7



	No. of Cases Colonized (N = 58)			
Organisms	No. of Cases	Percentage		
Pseudomonas	08	13.8		
Serratia	03	05.2		
Proteus	01	01.7		
Klebsiella Pneumoniae	08	13.8		
Enterobacter aerogenes	01	01.7		
Acinectobacter	02	03.4		
Burkholderiacepecia	02	03.4		

Table No. 10: Total Number of Organism Colonized (Gram -Ve)

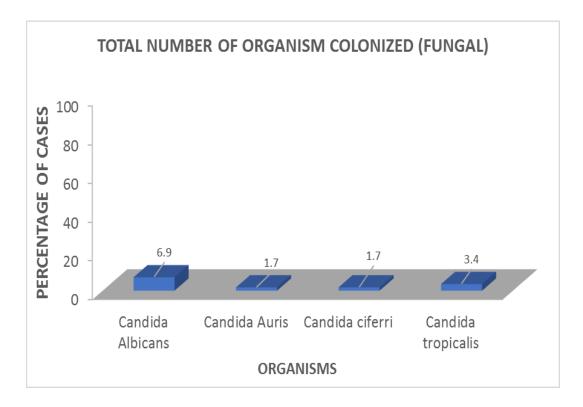
Graph 8



	No. of Cases Colonized (N = 58)		
Organisms	No. of Cases	Percentage	
Candida Albicans	04	06.9	
Candida Auris	01	01.7	
Candida ciferri	01	01.7	
Candida tropicalis	02	03.4	

Table No. 11: Total Number of Organism Colonized (Fungal)

Graph 9



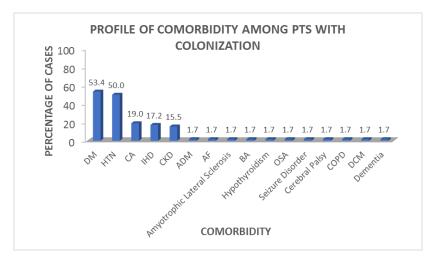
Cases with Diabetes and Hypertension had colonization of 50.0 -53.4% and 19.0% for cancer patients. (Table 12, Graph 10)

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	Cases with Colonization (N = 58)			
Comorbidity	No. of Cases	Percentage		
DM	31	53.4		
HTN	29	50.0		
CA	11	19.0		
IHD	10	17.2		
CKD	09	15.5		
CVA	01	01.7		
AF	01	01.7		
Amyotrophic Lateral Sclerosis	01	01.7		
BA	01	01.7		
Hypothyroidism	01	01.7		
OSA	01	01.7		
Seizure Disorder	01	01.7		
Cerebral Palsy	01	01.7		
COPD	01	01.7		
DCM	HOUMAN	01.7		
Dementia	01	01.7		

Table No. 12: Profile of Comorbidity Among Pts With Colonization

Graph 10



According to Table 13 and Graph, 11,89.5% of colonized cases with Staphylococcus were sensitive for antibiotics like Linezolid, 84.2% for Vancomycin and Teicoplanin whereas 78.9% of colonized cases with Staphylococcus were resistant to antibiotics like Penicillin, 52.6% for Levofloxacin, and 21.5% for Cotrimoxazole and Tetracycline. Among the colonized cases with Enterococcus, 87.5% were sensitive to antibiotics like Teicoplanin, 75.0% for Linezolid, Vancomycin, and Tigecycline whereas 50.0% of colonized cases with Enterococcus were resistant to antibiotics like Penicillin and Levofloxacin and 25.0% for Tetracycline. (Table14, Graph12)

According to Table 15 and Graph 13, 64.0% of colonized cases with Gram-negative were sensitive for antibiotics like Colistin and Minocycline, 56.0% for Amikacin whereas 44.0% of colonized cases with Gram-negative were resistant to antibiotics like Ciprofloxacin, 40.0% for Meropenem and Piperacillin/Tazobactam. 87.5% of colonized cases with Fungal were sensitive for antibiotics like Caspofungin and Micafungin, 37.5% for Flucytosine whereas 75.0% of colonized cases with Fungal were resistant to antibiotics like Voriconazole, 62.5% for Amphotericin – B and Fluconazole. (Table 16, Graph 14)

The cases with 4 co-morbidity had colonization of 10.0% which was less as compared to 34.7% cases with 3 co-morbidity, 20.0% cases with 2 co-morbidity, and 37.3% cases with 1 co-morbidity but the difference was statistically insignificant. (Table 17, Graph 15)

	No. of Cases Sensitive/Resistant cases (N = 19)				
Antibiotics	No. of Sensitive Cases	Percentage	No. of Resistant Cases	Percentage	
Penicillin	02	10.5	15	78.9	
Cotrimoxazole	13	68.4	04	21.5	
Levofloxacin	09	47.4	10	52.6	
Clindamycin	13	68.4	04	21.1	
Linezolid	17	89.5	-	-	
Vancomycin	16	84.2	01	05.3	
Tigecyclin	15	78.9	-	-	
Teicoplanin	16	84.2	-	-	
Tetracyclin	11	57.9	04	21.5	
Daptomycin	13	68.4	-	-	

 Table No. 13: Antibiotic Sensitivity for Staphylococcus Cases



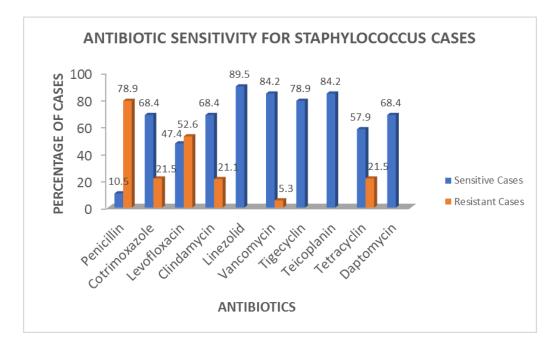


Table No. 14: Antibiotic Sensitivity for Enterococcus Cases

Antibiotics	No. of Cases Sensitive/Resistant cases (N = 8)					
	No. of Sensitive Cases	Percentage	No. of Resistant Cases	Percentage		
Penicillin	03	37.5	04	50.0		
Cotrimoxazole	02	25.0	-	-		
Levofloxacin	01	12.5	04	50.0		
Clindamycin	-	-	-	-		
Linezolid	06	75.0	-	-		
Vancomycin	06	75.0	-	_		
Tigecyclin	06	75.0	-	-		
Teicoplanin	07	87.5	-	-		
Tetracyclin	02	25.0	02	25.0		
Daptomycin	03	37.5		-		



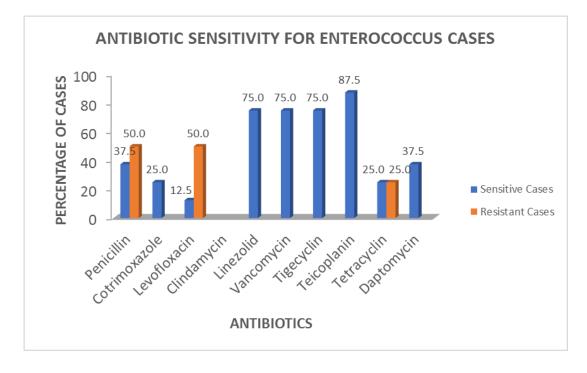


Table No. 15: Antibiotic Sensitivity for Gram Negative Cases

	No. of Cases Sensitive/Resistant cases (N = 25)				
	No. of		No. of		
Antibiotics	Sensitive A	Percentage	Resistant	Percentage	
Antibiotics	Cases	I el centage	Cases	rercentage	
Amikacin	14	56.0	08	32.0	
Amoxycillin/Clavulanic Acid	04	16.0	08	32.0	
Cefepime	10	40.0	09	36.0	
Cefoperazone + Sulbactam	11	44.0	06	24.0	
Ceftriaxone/Sulbactam/EDTA	20	80.0	-	-	
Ciprofloxacin	11	44.0	11	44.0	
Colistin	16	64.0	-	-	
Cotrimoxazole	12	48.0	05	20.0	
Imipenem/Cilastatin	09	36.0	07	28.0	
Meropenem	12	48.0	10	40.0	
Minocycline	16	64.0	02	8.0	
Piperacillin/Tazobactam	09	36.0	10	40.0	
Tigecycline	10	40.0	08	32.0	



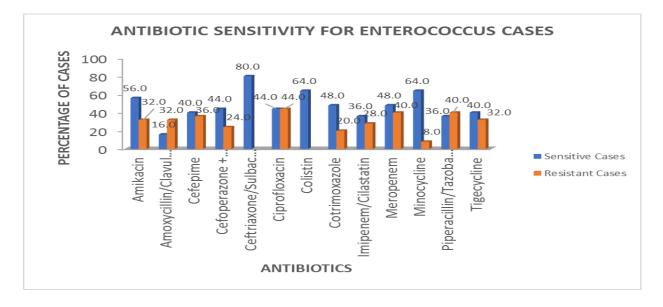


Table No. 16: Antibiotic Sensitivity for Fungal Cases

	No. of Cases Sensitive/Resistant cases (N = 8)				
Antibiotics	No. of Sensitive Cases	Percentage	No. of Resistant Cases	Percentage	
Amphotericin – B	02	- 25.0 AN	05	62.5	
Fluconazole	02	25.0	05	62.5	
Flucytosine	03	37.5	04	50.0	
Voriconazole	02	25.0	06	75.0	
Caspofungin	07	87.5	-	-	
Micafungin	07	87.5	-	-	

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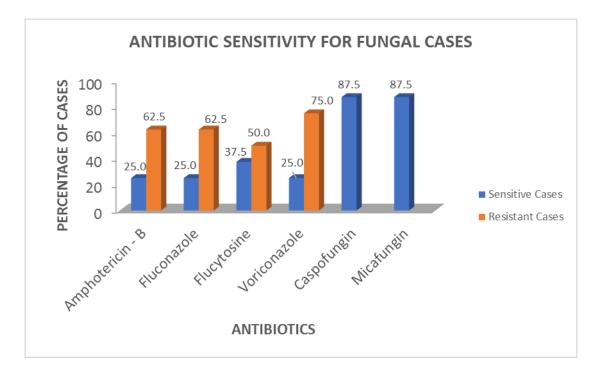
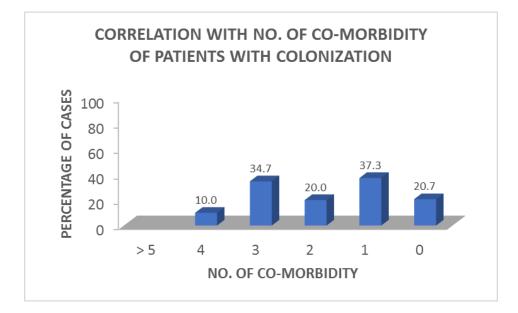


Table No. 17: Correlation With No. Of Co-Morbidity Of Patients With Colonization

	Cases with colonization No.		
No. of Co-Morbidity	No. of Cases	%	
> 5	-		
4	10	01 10.0	
3	49	17 34.7	
2	45	09 20.0	
1	67	25 37.3	
0	29	06 20.7	

By Chi Square Test P > 0.05 Not Significant





DISCUSSION

Central venous catheters (CVCs) though indispensable in current medical and intensive care treatment, also puts patients at risk of a catheter-related infection (CRI) resulting in increased morbidity and mortality. The rate of CVC-BSI across Indian hospitals ranges from 4.01/1000 catheter days to 9.26/1000 catheter days. CVC-associated complications like CLABSI, sepsis, septic thrombophlebitis, endocarditis result in significant morbidity, increased duration of hospitalization, and additional medical costs. Catheter colonization precedes catheter-related infections. Hence, there is a necessity of citing the organisms colonizing the catheter in critically ill patients and governing the antibiotic policies for effective management. The study was conducted on 200 patients admitted to SLR hospital fulfilling all inclusion and exclusion criteria. The mean age of the patients included in the study was 60.79 years with male predominance i.e. 62.5%. The total duration of catheter days was 1930. In the present study, out of 200 patients with a central line, 58 showed colonization with the incidence of colonization being 29%. The colonization rate in our study was found to be 30.03 per 1000 catheter days. The incidence rate of central line catheter colonization ranges from 18.3% to 78% in different studies¹⁵⁰⁻¹⁵⁴. According to MaríaJesúset al, colonization was found in 18.3% cases¹⁵⁵. According to Hagau N et al, colonization was found in 25.88% cases¹⁵⁶ which was similar to the observations found in our study. Patil et al reported the colonization rate of 31.34 per 1000 catheter-days¹⁵⁸. Gahlot et al reported a colonization rate of 36.4 per 1000 catheter days¹⁵⁷. These findings support the findings in our study. Organisms were

identified by the semiquantitative method i.e. Makis method for extraluminal growth and by the quantitative method for intraluminal growth. 24% of catheter tip cultures displayed colonization by the Makis method, and 13% colonization was displayed by the subculture method. The Makis method was used for tip culture by various studies including Harshal Shah *et al* ¹⁵⁹, Gahlot *et al*¹⁵⁷, Patil *et al* ¹⁵⁸. In our study, to define the severity of disease, the APACHE score was used as a parameter. THE mean APACHE score was 13.57. It was found that 27.7% of the cases had colonization with a reported Apache Score of < 15 which was comparable with 31.4% of the cases with an Apache Score of> 15. The difference found was not statistically significant. A similar finding was reported in a study conducted by Soni *et al* in which there was no relationship between APACHE Score and risk of catheter infection¹⁶⁰. Deliberato RO *et al* observed that the APACHE II mean value ± standard deviation score was 15.43±4.52 and there was no correlation between APACHE score and bloodstream infection¹⁶¹. Both these studies support observations made by our study.

Malnutrition is associated with increased mortality and morbidity, extends hospital length of stay, reduces the quality of life, and increases healthcare costs. Systematic screening at hospital admission by using the Nutritional Risk Screening-2002 score is recommended to detect patients at nutritional risk. In the present study, an attempt was made to correlate the Nutritional Risk Screening-2002 score with catheter colonization. It was found that 28.2% of the colonized cases had NRS (2002) < 4 which was comparable with 30.0% of the cases with NRS (2002) > 4 and the difference was not statistically significant. Thibault R *et al* observed that the proportion of patients at nutritional risk (Nutritional Risk Screening-2002 score >3) was not significantly different between patients with HCAI and non-infected patients and hence even though the Nutritional Risk Screening-2002 score is a validated nutritional screening tool, but its validity to identify patients at risk of HCAI remains to be determined in prospective studies¹⁶². However, this aspect lacks sufficient data and should be further investigated.

The Nutrition Risk in Critically ill (NUTRIC Score) is designed to quantify the risk of critically ill patients developing adverse events that may be modified by aggressive nutrition therapy. In the present study, an attempt was made to correlate the NUTRIC score with catheter colonization. It was found that 28.5% of the colonized cases had a NUTRIC score less than or equal to 5 which was comparable with 31.4% of the cases with a NUTRIC score >5and the difference was not statistically significant. There is a paucity of studies in which

correlation between NUTRIC score and no. of colonizers was established and needs further investigation.

In our study, out of 200 patients, 7 patients were neutropenic, and leukocytosis was present in 110 patients. Out of 7 neutropenic patients, 3 (42.9%) cases developed colonization of catheter and out of 110 leukocytosis patients, 32 (29.1%) patients developed colonization. The rate of colonization in Neutropenic patients was more as compared to 29.1% of the cases with Leukocytosis but the difference was not statistically significant. There is a paucity of studies in which correlation between abnormal WBC count and no. of colonizers was established and needs further investigation.

In the present study, the mean duration of the central catheter was 9.66 days¹⁰⁹. Catheters were placed for \geq 7 days, out of which 37 showed positive organisms comprising 33.9% of the total. Of the 91 catheters placed for less than 7 days, 21 (23.1%) showed biofilm formation. Although this difference was not statistically significant there is a clinical relationship between the duration of the central line *in situ* and the colonization. Thus, this may also indicate the increase in the number of cases of CLABSI with the increased duration of catheterization. Gil *et al* 1983 observed that in 220 central venous catheters in ICU patients, found that the incidence of sepsis rose from 1.5 to 10% when the duration of catheterization exceeded 6 days¹⁶³. Collignon *et al.* (1988) examined 780 catheters to determine the association between different insertion sites, duration of catheter insertion, and catheter-related sepsis¹⁶⁴. Richet H *et al* 1990 observed that central venous catheterization longer than five to seven days was associated with a higher risk of catheter-related infection¹⁶⁵.

In our study, a benchmark of 7 days is selected because according to few studies those catheters that are inserted for a short period (7 days) are usually colonized intraluminally (contamination of the hubs). 7 days catheterization was also used as a reference in a study by Sadoyma *et al*¹⁶⁶.

Out of 200 patients, 197 received antibiotics. 96 patients received antibiotics for <7 days and 101 patients received antibiotics for >7 days. 19.8% of the cases that had colonization used Antibiotics for <7 days which was significantly less as compared to 38.6% of the cases that used Antibiotics >7 days.

Thus, this may indicate that a longer duration of treatment received by the patient indicates a grave prognosis which might explain the increased incidence of catheter colonization in such

critically ill patients. Since there is a paucity of studies regarding the correlation between the duration of antibiotics received by patients and no. catheter colonization, the establishment of correlation needs more research.

There was no temporal relationship between the presence of comorbidities and the colonization of the central line tip. In this study, the occurrence of comorbidities in the colonized samples was Diabetes (53.4%), hypertension (50%), carcinoma(19%), IHD (17.2%), and Chronic Kidney Disease (15.5%) with no clinically and statistically significant difference. Until now there has been no study highlighting the relationship between the colonization rate and the underlying disease.

In our study, out of 200 processed catheters, 58 showed significant growth. Out of which 27 (46.55%) showed growth of gram-positive organisms. which complies with a study done in Manipal⁸¹ where 42.16% of the pathogens causing tip colonization were due to Grampositive. In our study, among gram-positive organisms, the most commonly observed growth was Staphylococcus Epidermis (24.1%) followed by Staphylococcus Aureus (6.9%) followed by Enterococcus Faecalis(5.2%). Wisplinghoff *et al* ¹⁶⁷ showed that the commonest isolates were CONS (31%) followed by S. aureus (20%). Subba Rao *et al* ¹⁶⁸ showed that the commonest isolates in ICU patients were CONS (32.4%). Both these studies support observations found in our study. Patil et al 2011 observed that CONS were the most common organisms responsible for culture-positive catheter colonization and among CONS, Staphylococcus aureus accounted for 15% growth¹⁷⁰. Richet *et al.* in 1990 isolated S. aureus as the second most common microorganism from both central and peripheral venous catheters, accounting for 19.9% of all isolates¹⁶⁵.

In our study, the growth of all three organisms i.e. Enterococcus Cloaace and Enterococcus Faecium, and Staphylococcus Lentus were reported to be 1.7%. Other Enterococcal species showed a total growth of 5.2%.

In our study, out of 200 processed catheters, 58 showed significant growth.

Out of which 24 (41.37%) showed growth of gram-negative organisms. According to a study was done in Manipal 81,57.84% of the pathogens causing tip colonization were due to Gram-negative organisms. Among gram-negative organisms, Pseudomonas and Klebsiella pneumonia showed maximum growth in 13.8% cases each.

According to the done in Manipal⁸¹, P. aeruginosa was the most common gram-negative organism isolated (16.67%). According to a study conducted in AIIMS Delhi, Klebsiella pneumonia was found to be the most common gram-negative organism isolated from catheter tip colonization i.e. 11%¹⁷¹.

Both these studies support observations found in our study. In the study of Almuneef *et al* (2006), the most common organisms isolated were Klebsiella pneumonia 16%, coagulase-negative staphylococci 14%, and Pseudomonas aeruginosa $11\%^{172}$.In the present study, Serratia and Acinetobacter showed growth in 5.2% and 3.4% cases respectively. Proteus species, Enterobacter aerogenes, and Burkholderiacepecia showed growth in 1.7% cases each.

In the present study, out of 200 processed catheters, 58 showed significant growth. Out of which, 9 (15.51%) cases showed fungal growth. The most common fungus grown was Candida Albicans (6.9%) followed by Candida tropicalis (3.4%). Candida Auris and Candida ciferri showed growth of 1.7% cases each. In the study of Ramanathan Parameswaran *et al.*, (2011) 16% of infectious agents were Candida species¹³⁴. According to Pawar *et al.*, (2008) fungal pathogen isolated from CVC was Candida 11.7%^{173.}

These findings were similar to the findings of our study. In one study, Patil et al isolated C. Albicans from a single catheter tip $(5\%)^{158}$. Haslett et al found that 3% of the catheters were infected due to C. tropicalis¹⁷⁴ which is similar to observations of our study.

In the present study colonized cases with staphylococcus, most of which are staphylococcus epidermis (CONS) showed maximum susceptibility to Linezolid (89.5%), followed by Vancomycin and Teicoplanin. Whereas 78.9% of colonized cases with staphylococcus were resistant to antibiotics like Penicillin, 52.6% for Levofloxacin, and 21.5% for Cotrimoxazole, and Tetracyclin. which was similar to that in a study done by Khanna et al.⁸¹ where 100% sensitivity was seen to Linezolid.

The majority of the isolates causing colonization were resistant to Penicillin G, and all were sensitive to Vancomycin in this study. This was similar to studies done in Manipal, study done by Parameswaran *et al*^{134.}

Resistance to penicillin can be attributed to the fact that this is a tertiary care center in which patients are referred from smaller centers causing selection pressure exerted by extensive use of antibiotics and may be due to transmission of resistant clones between the patients.

This study showed that 87.5% of colonized cases with Enterococcus sp. were sensitive to antibiotics like Teicoplanin which is the most common antibiotic used in our hospital. Other antibiotics sensitivity pattern is 75.0% for Linezolid, Vancomycin, and Tigecycline whereas 50.0% of colonized cases with Enterococcus were resistant to antibiotics like Penicillin and Levofloxacin and 25.0% for Tetracyclin. Which was in concordance with a study done by Deliberato *et al* ^{161.}

Multiple drug resistance was found in gram-negative organisms. However, it should be noted that 80.0% of colonized cases with Gram-negative were sensitive for antibiotics Ceftriaxone/Sulbactam/EDTA, whereas no resistance was found for Ceftriaxone/Sulbactam/EDTA.

Among the other antibiotics gram-negative bacilli having a sensitivity of 64.0% for Colistin and Minocycline, 56.0% for Amikacin. 44.0% of colonized cases with Gram-negative were resistant to antibiotics like Ciprofloxacin, 40.0% for Meropenem and Piperacillin/Tazobactam.

This was similar to a study by Amin *et al.*, which showed 50% gram-negative isolates being resistant to Piperacillin-Tazobactam and meropenem, and 33% being resistant to imipenem¹⁷⁵.

Fungal colonization which predominantly grown Candida albicans shown that 87.5% of colonized cases with Fungal were sensitive for antibiotics like Caspofungin and Micafungin, 37.5% for Flucytosine whereas 75.0% of colonized cases with Fungal were resistant to antibiotics like Voriconazole, 62.5% for Amphotericin – B and Fluconazole.

CONCLUSION

There have been very few studies regarding the colonization rate of central lines. The data of true colonization rate from our single-center reveal that central lines almost equally get colonized with gram-positive and gram-negative organisms. This data also seems to suggest that comorbidities, severity of illness, or length of time that the catheter is in situ do not

change the colonization rate. This data may thus help us to make protocols for clinical management and in devising materials for making catheters.

Limitations:

1. Single-center study 2. Central lines were removed when indicated.

Clinical Significance:

This is the first study looking at the actual colonization rate as the authors have sampled every single line that was inserted during the study period. Data derived from this study will help to set protocols and device new catheter materials in the future.

REFERENCES

1. Wenzel RP, Edmond MB. The impact of hospital-acquired bloodstream infections. Emerg Infect Dis. 2001; 7:174-7.

2. Hadaway L C. Flushing vascular access catheters: Risks for infection transmission. Infection Control Resource.2006; 4(3): 1-7.

3. O'Grady NP, Alexander M, Dellinger EP, Gerberding JL, Heard SO, Maki DG, Masur H, McCormick RD, Mermel LA, Pearson ML, Raad II, Randolph A, Weinstein RA (2002) Guidelines for the prevention of intravascular catheter-related infections. MMWR Recomm Rep 51: 1-29.

4. Pascual A, Cercenado E, Salavert M, Sanchez-G J E, Eiros J M, Linares J et al. Update on pathogenesis and diagnosis of intravascular catheter-related infections. EnfermInfeccMicrobiol Clin.2011; 29(supl4): 16-21.

5. Eggiman P. Diagnosis of intravascular catheter infection. CurrOpin Infect Dis.2007;20:353-9.

6. M. Gominet, F. Compain, C. Beloin, D. Lebeaux. Central venous catheters and biofilms: where do we stand in 2017? APMIS 125 (4) (2017): 365-75.

7. Lebeaux, D, Ghigo, JM, Beloin, C. Biofilm related infections: bridging the gap between clinical management and fundamental aspects of recalcitrance toward antibiotics. MicrobiolMolBiol Rev2014; 78: 510–43.

8. Band, J. (2010). Pathogenesis of and risk factors for central venous catheter-related infections. Retrieved from http://www.uptodate.com/online/content/topic.do?topicKey=hosp_inf/6194&view=print

9. Barsanti MC, Woetje KF. Infection prevention in the intensive care unit. Infect Dis Clin N Am. 2009;23:703–25

10. Maiefski, M., Rupp, M., &Hermsen, E. Ethanol lock technique: Review of the literature. Infection Control and Hospital Epidemiology 2009; 30(11):1096-108.

11. Opilla, M., Kirby, D., & Edmond M. Use of ethanol lock therapy to reduce the incidence of catheter related infections in home parenteral nutrition patients. Journal of Parenteral and Enteral Nutrition; 31(4): 302-305.

12.Peters J. The history of central venous access. In: Hamilton H, Bodenham AR, editors. Central Venous Catheters. Chichester, United Kingdom. Wiley-Blackwell; 2009:1-13.

13. Seldinger SI. Catheter replacement of the needle in percutaneous arteriography (a new technique). ActaRadiol1953;39:368-76.

14. Niederhuber JE, Ensminger W, Gyves JW, Liepman M, Doan K, Cozzi E. Totally implanted venous and arterial access system to replace external catheters in cancer treatment. Surgery 1982;92(4):706-12.

15. CDC& P. Guidelines for prevention of intravascular catheter related infections. MMWR 2002:51 (RR10); 1-26.

16. Parker L. Management of intravascular devices to prevent infection. British J Nursing. 2002; 11:240-5.

17. Farkas J.C., Liu N., Bleriot J.P., Chevret S., Goldstein F.W. & Carlet J. Single versus triple-lumen central catheter-related sepsis: a prospective randomised study in a critically ill population. American Journal of Medicine 1992;93:277–82.

18. Dezfulian C., Lavelle J., Nallamothu B.K., Kaufman S.R. & Saint S. Rates of infection for single lumen versus multilumen central venous catheters: a meta-analysis. Critical Care Medicine 2003;31:2385–2390.

19. Pratt R.J., Pellowe C., Loveday H.P., Robinson N., Smith G.W., Barrett S., Davey P et al. The epic project: developing national evidence-based guidelines for preventing healthcare associated infections. Phase I: guidelines for preventing hospital-acquired infections. Department of Health (England). Journal of Hospital Infection 2001;47 (Suppl.), S1–S82.

20. Pellowe C.M., Pratt R.J., Harper P., Loveday H.P., Robinson N., Jones S.R. et al. Guideline Development Group. Evidence based guidelines for preventing healthcare-associated infections in primary and community care in England. Journal of Hospital Infection 2003;55 (Suppl. 2), S2–S127.

21. Cicalini S., Palmieri F. & Petrosillo N. Clinical review: new technologies for prevention of intravascular catheter-related infections. Critical Care 2004;8:157–162.

22. Randolph A., Cook D.J., Gonzales C.A. &Brun-Buisson C. Tunnelling short-term central venous catheters to prevent catheter-related infection: a meta-analysis of randomized controlled trials. Critical Care Medicine 1998;26:1452–57.

23. Pegues D., Axelrod P., McClarren C., Eisenberg B.L., Hoffman J.P. Ottery F.et al Comparison of infections with Hickman and implanted port catheters in adult solid tumor patients. Journal of Surgical Oncology 1992;49:156–62.

24. Camp-Sorrell D. Implantable ports: everything you always wanted to know. Journal of Intravenous Nursing 1992;15, 262–73.

25. Whitman E.D. Complications associated with the use of central venous access devices. Current Problems in Surgery 1996; 23:309–88.

26. Cortelezzi A.N., Moia M., Falanga A., Pogliani E.M., Agnelli G., Bonizzoni E., CATHEM Study Group. Incidence of thrombotic complications in patients with haematological malignancies with central venous catheters: a prospective multicentre study. British Journal of Haematology 2005;129:811–17.

27. Galloway S. & Bodenham A.R. Long-term central venous access. British Journal of Anaesthesia 2004:92, 722–34.

28. Hadaway L.C. Comparison of vascular access devices. Seminars in Oncology Nursing 1995; 11: 154-66.

29. BCSH Guidelines for the use of platelet transfusions. British Journal of Haematology 2003; 122:10–23.

30. Ansell J., Hirsh J., Poller L., Bussey H., Jacobson A. &Hylek E. The pharmacology and management of the vitamin K antagonists: the seventh ACCP conference on antithrombotic and thrombolytic therapy. Chest 2004;126:204–33.

31. Hatfield A. & Bodenham A.R. Portable ultrasound for difficult venous access. British Journal of Anaesthesia 1999;82:822–26.

32. O'Grady N.P., Alexander M., Dellinger E.P., Gerberdin J.L., Heard S.O., Maki D.G., et al Guidelines for the prevention of intravascular catheter-related infections. American Journal of Infection Control 2002;30:476–89.

33. Mermel L.A., McCormack R.D., Springman S.R. & Maki D.G. The pathogenesis and epidemiology of catheter-related infection with pulmonary artery Swan-Ganz catheters: a prospective study using molecular subtyping. American Journal of Medicine 1991;91 (Suppl. 3B):197S–205S.

34. Raad I.I., Hohn D.C., Gilbreath B.J., Suleiman N., Hill L.A., Bruso P.A., Marts K.et al. Prevention of central venous catheter-related infections by using maximal sterile barrier precautions during insertion. Infection Control and Hospital Epidemiology 1994; 15: 231–38.

35. Fletcher S.J. Catheter related bloodstream infection. Continuing Education in Anaesthesia. Critical Care & Pain 2005;5: 49–451.

36. Maki D.G., Ringer M. & Alvarado C.J. Prospective randomized trial of povidone iodine, alcohol and chlorhexidine for prevention of infection associated with central venous and arterial catheters. Lancet 1991:338:339–343.

37. Hind D., Calvert N., McWilliams R., Davidson A., Paisley S., Beverley C. et al Ultrasonic locating devices for central venous cannulation: a meta-analysis. British Medical Journal 2003;327: 361–68.

38. Pellowe C.M., Pratt R.J., Harper P., Loveday H.P., Robinson N., Jones S.R et al. Guideline Development Group. Evidence based guidelines for preventing healthcare-associated infections in primary and community care in England. Journal of Hospital Infection 2003;55 (Suppl. 2):S2–S127.

39. Goetz A., Wagener M., Miller J.M. & Muder R.R. .Risk of infection due to central venous catheters: effect of site of placement and catheter type. Infection Control and Hospital Epidemiology 1998;19, 842–45

40. Mermel L.A., McCormack R.D., SpringmanS.R. & Maki D.G. The pathogenesis and epidemiology of catheter-related infection with pulmonary artery Swan-Ganz catheters: a prospective study using molecular subtyping. American Journal of Medicine 1991; 91 (Suppl. 3B), 1978–205S.

41. Knutstad K., Hager B. & Hauser M. Radiological diagnosis and management of complications related to central venous access. Acta Radiologica 2003 ;44;508–16.

42. Maki D. G. Skin as a source of nosocomial infection: Directions for future research. Infection Control, 1986;7(2): 113-16.

43. Bjornson H. S., R. Colley, R. H. Bower er al. Association between microorganism growth at the catheter site and colonization of the catheter in patients receiving total parenteral nutrition.Surgery, 1982;92: 720-27.

44. Maki D. G. and M. Ringer .Evaluation of dressing regimens for prevention of infection with peripheral intravenous catheters. Journal of the American Medical Association1987;258: 2396-403.

45. Jarrard M. M., C. M. Olsen and J. B. Freeman. Daily dressing change effects on skin flora beneath subclavian catheter dressings during total parenteral nutrition. Journal of Parenteral and Enteral Nutrition, 1980;4(4): 391-92.

46. Olson, C., &Heilman, J. M. Clinical performance of a new transparent chlorhexidine gluconate central venous catheter dressing. Journal for the Association of Vascular Access 2008;13(1):13-19.

47. Zitella, L.Central venous catheter site care for blood and marrow transplant recipients. Clinical Journal of Oncology Nursing 2003;7(3):289-98.

48. Casey, A. L., & Elliott, T., S. Prevention of central venous catheter-related infection: update. British Journal of Nursing, 2010;19(2): 78-87.

49. Centers for Disease Control. Guidelines for the prevention of intravascular catheter related infections. (2002) http://www.cdc.gov/mmwr/peview/mmwrhtml/rr5110a1.htm .

50. Bowdle A. Vascular complications of central venous catheter placement: Evidence-based methods for prevention and treatment. J CardiothoracVascAnesth. 2014;28:358–68.

51. McGee DC, Gould MK. Preventing complications of central venous catheterization. N Engl J Med. 2003;348:1123–33.

52. Troianos CA, Jobes DR, Ellison N. Ultrasound-guided cannulation of the internal jugular vein. A prospective, randomized study. AnesthAnalg. 1991;72:823–6.

53. Randolph AG, Cook DJ, Gonzales CA, Pribble CG. Ultrasound guidance for placement of central venous catheters: A meta-analysis of the literature. Crit Care Med. 1996;24:2053–8.

54. Bowdle A. Vascular complications of central venous catheter placement: Evidence-based methods for prevention and treatment. J Cardiothorac Vasc Anesth. 2014;28:358–68

55. Vats HS. Complications of catheters: Tunneled and nontunneled. Adv Chronic Kidney Dis. 2012;19:188–9456. McGee DC, Gould MK. Preventing complications of central venous catheterization. N Engl J Med. 2003;348:1123–33.

57. Bhutta ST, Culp WC. Evaluation and management of central venous access complications. Tech VascIntervRadiol. 2011;14:217–24.

58. Khouzam RN, Soufi MK, Weatherly M. Heparin Infusion through through a central line misplaced in the carotid artery leading to hemorrhagic stroke. J Emerg Med. 2013;45:e87–9.

59. Konichezky S, Saguib S, Soroker D. Tracheal puncture. A complication of percutaneous internal jugular vein cannulation. Anaesthesia. 1983;38:572–4.

60. Malik IA, Adams RG. Tracheal cuff puncture: A complication of percutaneous internal jugular vein cannulation. Am J Med. 2003;115:590–1.

61. Huang YC, Huang JC, Chen SC, Chang JM, Chen HC. Lethal cardiac arrhythmia during central venous catheterization in a uremic patient: A case report and review of the literature. Hemodial Int. 2013;17:644–8.

62. Kusminsky RE. Complications of central venous catheterization. J Am Coll Surg. 2007;204:681–96.

63. Khouzam RN, Soufi MK, Weatherly M. Heparin Infusion through a central line misplaced in the carotid artery leading to hemorrhagic stroke. J Emerg Med. 2013;45:e87–9.

64. Andrews RT, Geschwind JF, Savader SJ, Venbrux AC. Entrapment of J-tip guidewires by Venatech and stainless-steel Greenfield vena cava filters during central venous catheter placement: Percutaneous management in four patients. CardiovascInterventRadiol. 1998;21:424–8.

65. Early TF, Gregory RT, Wheeler JR, Snyder SO, Jr, Gayle RG. Increased infection rate in double-lumen versus single-lumen Hickman catheters in cancer patients. South Med J. 1990;83:34–6.

66. Hilton E, Haslett TM, Borenstein MT, Tucci V, Isenberg HD, Singer C. Central catheter infections: Singleversus triple-lumen catheters. Influence of guide wires on infection rates when used for replacement of catheters. Am J Med. 1988;84:667–72.

67. Yeung C, May J, Hughes R. Infection rate for single lumen v triple lumen subclavian catheters. Infect Control HospEpidemiol. 1988;9:154–8.

68. Watson CM, Al-Hasan MN. Bloodstream infections and central line-associated bloodstream infections. SurgClin North Am. 2014;94:1233–44.

69. O'Grady NP, Alexander M, Burns LA, Dellinger EP, Garland J, Heard SO, et al. Healthcare Infection Control Practices Advisory Committee (HICPAC) (Appendix 1). Summary of recommendations: Guidelines for the prevention of intravascular catheter-related infections. Clin Infect Dis. 2011;52:1087–99.

70. Marsteller JA, Sexton JB, Hsu YJ, Hsiao CJ, Holzmueller CG, Pronovost PJ, et al. A multicenter, phased, cluster-randomized controlled trial to reduce central line-associated bloodstream infections in intensive care units.Crit Care Med. 2012;40:2933–9.

71. Hoshal VL., Jr Total intravenous nutrition with peripherally inserted silicone elastomer central venous catheters. Arch Surg. 1975;110:644–6.

72. Klevens RM, Edwards JR, Richards C L et al. Estimating health-care associated infections and deaths in U.S hospitals, 2002. Public Health Rep.2007; 122(2):160-166.

73. Maki DG, Kluger DM, Crnich C J. The risk of bloodstream infection in adults with different intravascular devices: a systematic review of 200 published prospective studies. Mayo ClinPrac 2006; 81(9):1159-1171.

74. Fauci, Braunwald, Kasper, Hauser, Longo, Jameson et al., chapter 125: Health care-associated infections. Harrison's Principles of Internal Medicine, 17th edition, p 170-182.

75. Raad I, Hanna H, Maki D. Intravascular catheter-related infections: advances in diagnosis, prevention, and management. Lancet Infect Dis.2007;7: 645-57.

76. Horan T C, Andrus M, Dudeck M A. CDC/NHSN surveillance definition of health care-associated infection and criteria for specific types of infections in the acute care setting. Am J Infect Control.2008; 36(5):309-332.

77. Mandell G.L, Bennett J.E, Dolin R. Mandell, Douglas And Bennett's Principles And Practice Of Infectious Diseases; 7th edition; vol 2: 3696-715.

78. Maki D. G. Marked differences in skin colonisation of insertion sites for central venous catheters, arterial and peripheral IV catheters: The major reason for differing risks of catheter related infection. Abstracts of the Inter-science Conference on Antimicrobial Agents and Chemotherapy, No. 1990; 712: 205.

79. Chopdekar K, C Chande, P Veer, S Chavan, V Wabale, K Vishwakarma et al. Central venous catheter– related bloodstream infection rate in critical care units in a tertiary care, teaching hospital in Mumbai. Indian Journal of Medical Microbiology 2011; 29(2):169-71.

80. Rao S S D, Joseph M P, Lavi R and Macaden R. Infection related to vascular catheters in paediatric intensive care unit. Indian Paediatrics 2005; 42:667-672

81. Khanna V, Mukhopadhayay C, K. E Vandana, Verma M, and Dabke P. Evaluation of Central Venous Catheter Associated Blood Stream Infections: A Microbiological Observational Study. Journal of Pathogens 2013;2013:936864.

82. Lane, R. & Matthay, M. Central line infections. Current Opinion in Critical Care 2002;441-48.

83. Soufir L, Timsit JF, Mahe C, Carlet J, Regnier B, Chevret S. Attributable morbidity and mortality of catheter-related septicemia in critically ill patients: a matched, risk adjusted, cohort study. Infect Control HospEpidemiol 1999;20(6):396-401.

84. Crnich CJ, Maki DG. The promise of novel technology for the prevention of intravascular device-related bloodstream infection. II. Long-term devices. Clin Infect Dis 2002;34(10):1362-8.

85. Farr BM. Preventing vascular catheter-related infections: current controversies. Clin Infect Dis 2001;33(10):1733-8.

86. Bagnall-Reeb, H. (2004). Evidence for the use of antibiotic lock technique. Journal of InfusionNursing; 27(2), 118-122.

87. Galloway, M. (2010). Insertion and placement of central catheters in the oncology patient. Seminars in Oncology Nursing; 26(2), 102-112.

88. Cirioni O., Giacometti A., Ghiselli R., Dell' Acqua G., Orlando F., Mocchegiani F., et al. (2006). RNAIIIinhibiting peptide significantly reduces bacterial load and enhances the effect of antibiotics in the treatment of central venous catheter-associated Staphylococcus aureus infections. J. Infect. Dis. 193 180–186. 10.1086/498914.

89. Maki D. G., C. E. Weise and H. W. Sarafin A semi quantitative method for identifying intravenous catheterrelated infection. New England Journal of Medicine 1977;296(23): 1305-09.

90. Snydman D. R. and B. R. Murphy. Parenteral nutrition-related infections. American Journal of Medicine 1982;73: 695-99.

91. Armstrong C. W., C. G. Mayhall, K. B. Miller, et al .Clinical predictors of infection of central venous catheters used for total parenteral nutrition. Infection Control and Hospital Epidemiology, 1990;11: 71-78.

92. Cheesbrough J. S., R. G. Finch and R. P. Burden. A prospective study of the mechanisms associated with haemodialysis catheters. Journal of Infectious Diseases 1986;154(4): 579-89.

93. Kristinsson K. G., I. A Burnett and R. C. Spencer. Evaluation of three methods for culturing long intravascular catheters. Journal of Hospital Infection 1989 14: 183-91.

94. Bozzetti F., G. Bonfanti, E. Regalia et al. A new approach to the diagnosis of central venous catheter sepsis. Journal of Parenteral and Enteral Nutrition 1991; 15(4): 42-416.

95. Raad I, Luna M, Khalil SA, et al. The relationship between the thrombotic and infectious complications of central venous catheters. JAMA. 1994; 271:1014-1016.

96. Templeton A, Schlegel M, Fleisch F, et al. Multilumen central venous catheters increase risk for catheterrelated bloodstream infection: prospective surveillance study. Infection. 2008; 36:322-327.

97. Merrer J, De Jonghe B, Golliot F, et al. Complications of femoral and subclavian venous catheterization in critically ill patients: a randomized controlled trial. JAMA. 2001; 286:700-707.

98. Richet H, Hubert B, Nitemberg G, et al. Prospective multicentre study of vascular-catheter-related complications and risk factors for positive central-catheter cultures in intensive care unit patients. JClin Microbiol. 1990; 28:2520-25.

99. Rhame F, Maki D, Bennett J. Intravenous Cannula-Related Infections. Boston, Mass: Little, Brown; 1979.

100. Maki D, Goldmann D, Rhame F. Infection control in intravenous therapy. Ann Intern Med. 1973;79:867-87.

101. Soifer NE, Borzak S, Edlin BR, et al. Prevention of peripheral venous catheter complications with an intravenous therapy team: a randomized controlled trial. Arch Intern Med.1998; 158:473-77.

102. Fridkin SK, Pear SM, Williamson TH, et al. The role of understaffing in central venous catheter-associated bloodstream infections. Infect Control HospEpidemiol. 1996; 17:150-58.

103. Raad I, Hachem R, Hanna H, et al. Sources and outcomes of bloodstream infections in cancer patients: the role of central venous catheters. Eur J Clin Microbiol Infect Dis. 2007; 26:549-56.

104. Robert J, Fridkin SK, Blumberg HM, et al. The influence of the composition of the nursing staff on primary bloodstream infection rates in a surgical intensive care unit. Infect Control Hosp Epidemiol. 2000; 21:12-17

105. Maki D. Growth properties of microorganisms in infusion fluid and method of detection. In: Phillips I, ed. Microbiologic Hazards of Intravenous Therapy. Lancaster, UK: MTP Press; 1977:13-47.

106. Jarvis W, Highsmith A. Bacterial growth and endotoxin production in lipid emulsion. J ClinMicrobiol. 1984; 19:17-20.

107. Douce RW, Zurita J, Sanchez O, et al. Investigation of an outbreak of central venous catheter-associated bloodstream infection due to contaminated water. Infect Control Hosp Epidemiol.2008; 29:364-66.

108. Sitges-Serra A, Puig P, Jaurrieta E, et al. Hub colonization as the initial step in an outbreak of catheterrelated sepsis due to coagulase negative staphylococci during parenteral nutrition. J Parenter Enteral Nutr. 1984; 8:668-72.

109. Maki D. G. and I. M. McCormack. Acetone defatting catheter insertion sites in total parenteral nutrition: Its value as an infection control measure. American Journal of Medicine 1987; 83: 833-41.

110. Frasca D et al. Prevention of central venous catheter related infection in the intensive care unit. Critical Care 2010;14:212.

111. Wickham R., S. Purl and D. Welker. Long-term central venous catheters: Issues for care. Seminars in Oncology Nursing. 1992; 8(2): 133-147.

112. Maki D. G., L. Cobb J. K. Garman et al. An attachable silver-impregnated cuff for prevention of infection with central venous catheters: a prospective randomized multicenter trial. American Journal of Medicine 1988; 85(Sept): 307-14.

113. Toole G.O, Kaplan H.B, Kolter R. Biofilm formation as microbial development. Annual Review of Microbiology 2000; 54:49-79.

114. Kokare C R, S Chakraborthy, Khopade A N and Mahadik K R. Biofilm: Importance and applications. Indian Journal of Biotechnology. 2009; 8:159-168.

115. Oliveira A and M L R S Cunha. Bacterial biofilms with emphasis on coagulase-negative Staphylococci. J.Venom. Anim. Toxins incl. Trop. Dis. 2008;14;4:572-96.

116. O'Toole G. To build a biofilm. J Bacteriol. 2003;185:2687-89.

117. Nickel J. C., A. J. Gristina and J. W. Costerton. Electron microscopic study of an infected Foley catheter. The Canadian Journal of Surgery. 1985;28(1): 50-52.

118. El Helou, G, Viola, GM, Hachem, R, Han, XY, Raad, II. Rapidly growing mycobacterial bloodstream infections. Lancet Infect Dis 2013;13: 166–74.

119. Safdar, N, Maki, DG. The pathogenesis of catheter-related bloodstream infection with noncuffed short term central venous catheters. Intensive Care Med 2004;30:62–7.

120. Christensen GD, Simpson WA, Bisno AL, BeacheyEH. Adherence of slime-producing strains of Staphylococcus epidermidis to smooth surfaces. Infect Immun1982;37:318–26.

121. Costerton JW, Cheng KJ, Geesey GG, Ladd TI, Nickel JC, Dasgupta M, et al. Bacterial biofilms in nature and disease. Annu Rev Microbiol1987;41:435–64.

122. Donlan RM. Biofilms: microbial life on surfaces.Emerg Infect Dis 2002;8:881-90.

123. Flemming H-C, Wingender J. The biofilm matrix. Nat Rev Microbiol2010;8:623-33.

124. Gallis H. A. Normal flora and opportunistic infection.InJoklik W. K., H. P. Willett and D. B. Amos Eds. Zinnser Microbiology Connecticut: Appleton-Century –Crofts 1984;435-42.

125. Peters G., R. Locci and G Pulverer. Adherence and growth of staphylococci on surfaces on intravenous catheters. Journal of Infectious Diseases. 1982;146: 479-82.

126. Bose S, Khodke M, Basak S, Mallick S K. Detection of biofilm producing Staphylococci: Need of the hour. Journal of Clinical And Diagnostic Research. Dec 2009:3(6):1915-20.

127. Lorente L, Henry C, Martín MM, Jiménez A and Mora M L. Central venous catheter-related infection in a prospective and observational study of 2,595 catheters. Critical Care 2005; 9(6):R631-R635.

128. Akan O A. Microorganisms isolated from catheter tip cultures: Ibn-ina Hospital 2002. Journal Of Ankara Medical School 2003; 25(3):113-18.

129. Fluit A.C, Schmitz F.J and Verhoef J. Frequency of isolation of pathogens from bloodstream, nosocomial pneumonia, skin and soft tissue, and urinary tract infections occurring in European patients. Eur J ClinMicobiol Infect Dis.2001; 20:188-91.

130. Sandoe J.A.T, Witherden I.R, Cove J.H, Heritage J and Wilcox M.H. Correlation between Enterococcal biofilm formation in vitro and medical-device-related infection potential in vivo. J Med Microbiol, July 2003; 53(7):547-50.

131. Sarkar B,Biswas D, Prasad R, Sharma JP. A clinical microbiological study on the Importance of Pseudomonas in nosocomially infected ICU patients, with special reference to metallo beta lactamase production. Ind J PatholMicrobiol2006; 49:44-48.

132. Lorente L, Henry C, Martín MM, Jiménez A and Mora M L. Central venous catheter-related infection in a prospective and observational study of 2,595 catheters. Critical Care 2005; 9(6):R631-R635.

133. Allen SD, Janda WM, Koneman EW, Schreckenberger PC, Winn WC. The Enterobacteriaceae. In Koneman's Color Atlas and textbook of diagnostic microbiology 6th edition, Philadelphia. J.B. Lippincott, 2006; 212-302.

134. Parameswaran R, Sherchan J B., Varma M, Mukhopadhyay C, Vidyasagar S. Intravascular catheter-related infections in an Indian tertiary care hospital. J Infect Dev Ctries 2011; 5(6):452-58.

135. Pfaller MA, Diekema DJ, Jones RN, Sader HS, Fluit AC, Hollis RJ, Messer. SASENTRY Participant Group. International surveillance of bloodstream infections due to Candida species: frequency of occurrence and invitro susceptibilities to fluconazole, ravuconazole, and voriconazole of isolates collected from1997 through 1999 in the SENTRY antimicrobial surveillance program. JClinMicrobiol2001;39:3254–9.

136. Wisplinghoff H, Bischoff T, Tallent SM, Seifert H, Wenzel RP, Edmond MB. Nosocomial bloodstream infections in US hospitals: analysis of 24,179 cases from aprospective nationwide surveillance study. ClinInfectDis. 2004;39:309–17.

137. Mean M, Marchetti O, Calandra T. Bench -to -bedside review: Candida infections in the intensive care unit.CritCare2008;12:204.4.

138. Diekema DJ, Messer SA, Brueggemann AB, Coffman SL, Doern GV, Herwaldt LA, Pfaller MA. Epidemiology of candidemia:3-year results from the emerging infections and the epidemiology of Iowor ganisms study. JClin Microbiol.2002;40:1298–302.

139. Kao AS, Brandt ME, Pruitt WR, Conn LA, Perkins BA, Stephens DS, et al. The epidemiology of candidemiain two United States cities: results of apopulation-based active surveillance.ClinInfectDis1999;29:116470.

140. Tortorano AM, Kibbler C, Peman J, Bernhardt H, Klingspor L, GrillotR. Candidaemiain Europe:epidemiology and resistance. IntJ Antimicrob Agents. 2006;27:359–66.

141. Richet H, Roux P, Des Champs C, Esnault Y, Andremont A. Candidemiain French hospitals: incidence rates and characteristics. Clin MicrobiolInfect2002;8:405–12.

142. Yusuke Yoshino, Yoshitaka Wakabayashi, Satoshi Suzuki, Kazunori Seo, Ichiro Koga, Takatoshi Kitazawa, Shu Okugawa, Yasuo Ota Singapore Med J. 2014 Nov; 55(11): 579–82.

143. Norwood S., A. Ruby, J. Civetta and V. Cortes Catheter-related infections and associated septicemia. Chest. 1991;99(4): 968-75.

144. Kruse J. A. and N. J. Shah .Detection and prevention of central venous catheter-related infections. Nutrition in Clinical Practice 1993;8(4): 163-70.

145. Sitges-Serra A, and J. Linares .Limitations of semi-quantitative method for catheter culture. Journal of Clinical Microbiology. 1988;26: 1074-1076.

146. Hampton A. A. and R. J. Sheretz.Vascular-access infections in hospitalized patients. Surgical Clinics of North America 1988;68(1): 57-71.

147. Maki D. G., C. E. Weise and H. W. Sarafin (1977) A semi quantitative method for identifying intravenous catheter-related infection. New England Journal of Medicine, 296(23): 1305-09.

148. Cleri D. J., M. L. Corrado, S. J. Seligman et al. Quantitative culture of intravenous catheters and other intravascular inserts. Journal of Infectious Diseases 1980;141(6): 781-86.

149. Pettigrew R. A., S. D. R. Lang, D. A. Haydock et al. Catheter-related sepsis in patients on intravenous nutrition: A prospective study of quantitative catheter cultures and guidewire changes for suspected sepsis.British Journal of Surgery 1985;72: 52-55.

150. Meadows C, Creagh-Brown B, Nia T, Bonnici K, Finney S. Definition of catheter-related bloodstream infection as a quality improvement measure in intensive care. Critical Care. 2009;13:191. Crossref

151.Tullu MS, Deshmukh CT, Baveja SM. Bacterial profile and antimicrobial susceptibility pattern in catheter related nosocomial infections. Postgrad. Med. J 1998;44:7-13.

152. Karpel E, Kunsdorf-Wnuk A, Musiol E, Skorupa A, Arct-DanielakD, Jarosz U. Cathether related blood stream infection in ICU patients with prolonged central venous catheterisation cause and prevention]. Polskimerkuriuszlekarski : organ Polskiego Towarzystwa Lekarskiego. 2006;21:211-7.

153. Pérez-Granda MJ, Guembe M, Cruces R, Barrio JM, Bouza E. Assessment of central venous catheter colonization using surveillance culture of withdrawn connectors and insertion site skin. Critical Care. 2016;20:32. Crossref.

154. Hodzic S, Tihic N, Smajic J, Omerbegovic M, Sljivic M. Frequency of the central venous catheter colonization in surgical intensive care unit. Medicinskiarhiv. 2010;64:245-7. Crossref.

155. MaríaJesús Pérez-Granda, MaríaGuembe, Raquel Cruces, José María Barrio, Emilio Bouza Crit Care. 2016; 20:32. Published online 2016 Feb 2. doi: 10.1186/s13054-016-1201-0.

156. Hagau N, Flonta, M.; Slavcovici, A.2; Studnicska, D.1; Cocu, S.1; Mlesnite, M.1; Gavrus, R.1; Laslo, C.1 European Journal of Anaesthesiology (EJA). 2006;23: 200.

157. Gahlot R, Nigam C, Kumar V, Gupta M. Catheter related bloodstream infections in ICU: A study from North India. International Journal of Infection Control. 2013;20:9(2).

158. Patil HV, Patil VC, Ramteerthkar MN, Kulkarni R. Central venous catheter related bloodstream infections in the intensive care unit. Indian J Crit Care Med 2011;4:213–23.

159. Harshal Shah, Wendelyn Bosch, Kristine M. Thompson, Walter C. Hellinger Neurohospitalist. 2013; 3(3): 144–51.

160. Soni, Ranju A. et al.Catheter-related bloodstream infection (crbsi) rates in a mixed medical-surgical icu population before and after the implementation of a central line bundle (clb).Chest, volume 134, Issue 4, 3S.

161. Deliberato RO, Marra AR, Corrêa TD, et al. Catheter related bloodstream infection (CR-BSI) in ICU patients: making the decision to remove or not to remove the central venous catheter. PLoS One. 2012;7(3):e32687.

162. Thibault R, Makhlouf A-M, Kossovsky MP, Iavindrasana J, Chikhi M, Meyer R, et al. Healthcare-Associated Infections Are Associated with Insufficient Dietary Intake: An Observational CrossSectional Study. PLoS ONE 2015;10(4): e0123695.

163. Gill V. J., S. T. Selepak and E. C. Williams. Species identification and antibiotic susceptibilities of coagulase-negative staphylococci isolated from clinical specimens. Journal of Clinical Microbiology. 1983;18:1314-19.

164. Collignon P., N.Soni, I. Pearson et al. Sepsis associated with central vein catheters in critically ill patients. Intensive Care Medicine 1988;14: 227-31.

165. Richet H, Hubert B, Nitemberg G, Andremont A, Buu-Hoi A, Ourbak P, Galicier C, Veron M, Boisivon A, Bouvier AM .Prospective multicenter study of vascular-catheter-related complications and risk factors for positive central-catheter cultures in intensive care unit patients. J ClinMicrobiol1990;28: 2520-25.

166. Sadoyma, Geraldo, DiogoFilho, Augusto, &GontijoFilho, Paulo Pinto. Central venous catheter-related bloodstream infection caused Staphylococcus aureus: microbiology and risk factors. Brazilian Journal of Infectious Diseases 2006;10(2):100-06

167. Wisplinghoff, H, Bischoff, T, Tallent SM, Seifert H, Wenzel RP, Edmond MB .Nosocomial bloodstream infections in US hospitals: Analysis of 24,179 cases from a prospective nationwide surveillance study. Clin Infect Dis. 2004;39: 309-17.

168. Subba Rao, Joseph MP, Lavi R, Macaden R. Infections related to vascular catheters in a pediatric intensive care unit. Indian Pediatrics 2005;42: 667-72.

169. J Chander, S Gombar, V Gupta, M Kaur, T Sahoo. Incidence, risk factors, microbiology of venous catheter associated bloodstream infections - A prospective study from a tertiary care hospital Indian Journal of Medical Microbiology. 2015;33(2): 248-54.

170. Harsha V. Patil, Virendra C. Patil, M. N. Ramteerthkar, R. D. Kulkarni .Central venous catheter-related bloodstream infections in the intensive care unit, Indian J Crit Care Med. 2011;15(4): 213–23.

171.Deepti, Sanjeev Sinha, Surendra K. Sharma .Central Venous Catheter Related Bloodstream Infections in Medical Intensive Care Unit Patients in a Tertiary Referral Centre, Indian J Chest Dis Allied Sci. 2014;56:85-91.

172. Almuneef MA, Memish ZA, Balkhy HH, Hijazi O, Cunningham G, Francis C. Rate, risk factors and outcomes of catheterrelated bloodstream infection in a paediatric intensive care unit in Saudi Arabia. J Hosp Infect. 2006;62:207–13.

173. Pawar M, Mehta Y, Kapoor P, Sharma J, Gupta A, Trehan N. Central venous catheter-related bloodstream infections:incidence, risk factors, outcome, and associated pathogens. J CardiothoracVascAnesth. 2004;18:304–8.

174. Haslett TM, Isenberg HD, Hilton E, Tucci V, Kay BG, Vellozzi EM. Microbiology of indwelling central intravascular catheters. J ClinMicrobiol1988;26:696-701.

175. Amin N.S, Sh.Devi N and Bhat S. Evaluation of semiquantitative culture method in the diagnosis of central venous intravascular catheter related infections. J Pharm Biomed Sci. 2013;29 (29):724-28.

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