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A Review: Gastro Retentive Dosage Form with Special Emphasis on Floating Beads



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ABSTRACT

Any drug delivery system's primary goal is to achieve the desired drug concentration in blood or tissue, which is therapeutically efficacious and nontoxic over an extended length of time. Floating beads have a gastro retentive function without changing gastric emptying rate and are utilized for controlled medication release. Drugs with a limited window of absorption in a specific area of the gastrointestinal tract can help increase absorption and bioavailability by using floating drug delivery systems that can be kept in the stomach this can accomplish by utilizing different polymers. They exhibit significant patient benefits. One innovative medication delivery technique to increase stomach retention duration is floating beads. Several gastro-retentive drug delivery methods, including floating and nonfloating.

INTRODUCTION

Due to its simplicity of administration, patient compliance and adaptability in formulation, oral medication delivery is by far the preferred method of drug delivery. Oral dosage formulations have advanced significantly, going from instant release to site-specific delivery.¹

In the Sustained release dosage form, one or more medications are continually released in a predetermined sequence for a set amount of time, either systemically or locally to a specific target organ. It offers more consistent distribution, decreased dosing frequency, less side effects, improved regulation of plasma drug levels, and reduced side effects.²

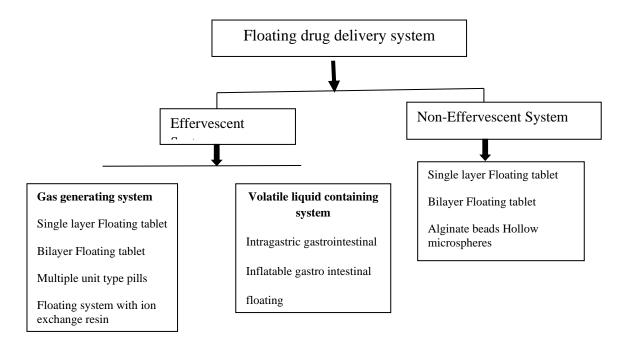
For dosage forms that stay in the stomach longer than conventional dosage forms, the capacity to extend and control the emptying time is a major asset. Gastric emptying of dosage forms is a highly variable process. Currently, a variety of methods are employed to extend stomach retention time. These include delayed gastric emptying devices such as floating drug delivery systems, hydro dynamically balanced systems, swelling and expanding systems, polymeric bioadhesive systems, modified-shape systems, high-density systems, and others.³

To keep pharmaceuticals in the stomach, floating drug delivery systems (FDDS) were developed. These devices are useful for medications that have poor intestinal fluid solubility and stability. Making the dosage form less dense than the stomach fluids allows it to float on them, which is the theory behind it.⁴ Since it is less dense than gastric fluids; it floats in the stomach for a longer duration without slowing down the gastric emptying rate. ⁵

The goal of developing a gastro retentive drug delivery system was to not only sustain drug release but also extend the time that dosage forms remain in the stomach until being released at the right time. The different sphere of beads is a medicine that is coated or enclosed in a microcapsule that serves as the solid substrate for the core of the bead. Beads may have features for controlled release.⁶

Floating drug delivery system is classified as effervescent and non-effervescent system as shown below.^{7,8}

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2. Effervescent system

It is a matrix-type system made with the aid of effervescent compounds and swellable polymers like methylcellulose and chitosan. For example, sodium bicarbonate, citric acid, tartaric acid, and These are designed such that when they come into contact with gastric contents, CO₂ is released and held in swelling hydrocolloid, giving the dosage form buoyancy and causing it to float over time.

They are again classified into:

- a) Gas generating system
- b) Volatile liquid-containing system

a) Gas generating system:

These buoyant systems rely on an effervescent interaction between citric/tartaric acid and carbonate/bicarbonate salts. The system is set up in such a way that when the formulation enters the stomach, carbon dioxide is released, causing it to float there. Other substances, such as a sodium alginate and sodium bicarbonate mixture and floating pills with several units that release carbon dioxide when swallowed, have been described. ⁹

b) Volatile liquid-containing system:

A medication delivery system's GRT can be maintained by including an inflatable chamber, which is filled with a liquid, such as ether or cyclopentane that gasifies at body temperature to expand the stomach chamber.¹⁰

The apparatus may also include a bioerodible plug made of polyvinyl alcohol, polyethylene, or another material that slowly dissolves and causes the inflatable chamber to release gas and collapse after a predetermined period, allowing the inflatable systems to spontaneously eject themselves from the stomach.¹¹

3. Non effervescent system

The formulation of non-effervescent FDDS can be accomplished by combining the drug with hydrocolloids that form a gel when in contact with gastric fluid after oral administration and maintain shape integrity and a bulk density barrier. The air trapped by the swollen polymer gives the dosage forms buoyancy.

They are classified into –

- a) Colloidal gel barrier systems
- b) Microporous Compartment systems
- c) Alginate beads
- d) Hollow microspheres.

a) Colloidal gel barrier system:

This method comprises medication that forms gels with hydrocolloids and floats on the contents of the stomach. This lengthens the GI stay and improves the Drug is delivered to the site of absorption in solution form, ready for absorption. The hydrocolloid hydrates in the system create a colloidal gel barrier surrounding stomach fluid when in contact with it. The barrier of produced colloidal gel regulates the pace of fluid entry into the apparatus, followed by medication release.¹²

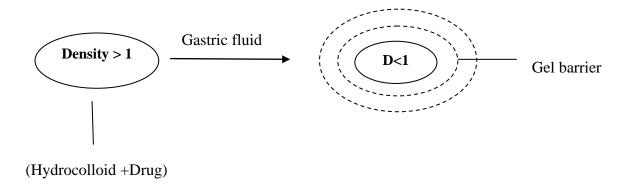


Figure no. 1: Colloidal gel barrier system.

b) Microporous Compartment systems:

A drug reservoir is enclosed in a microporous compartment with pores on the top and bottom of the compartment. To maintain the reservoir compartment's exterior walls were entirely sealed and free of undissolved drugs. New levodopa gastro retentive dosage forms are used that combine extended dimensions with high stiffness. These dosage forms are based on unfolding polymeric membranes.¹³

This method can be used to create huge gelatin capsules. The In vitro unfolded state was obtained within 15 minutes of treatment, according to tests, and it was also confirmed using beagle dogs. The unfolded form was kept in place for at least two hours. It was determined that the treatment of various medications with limited absorption windows might be enhanced by this method.¹⁴

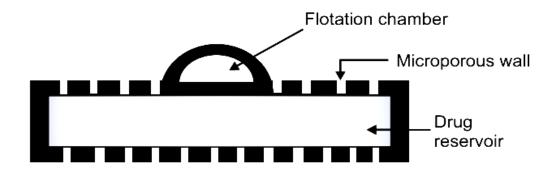


Figure no. 2: Microporous Compartment systems.

c) Hollow microspheres:

Hollow microspheres are non-effervescent gastroretentive drug delivery systems. They are empty spherical particles with no core. They have the distinct advantage of having multiple unit systems, and their center hollow space provides good floating properties, making them promising buoyant systems. These microspheres are free-flowing, low-density powders with a diameter of less than 200 m made of proteins or synthetic polymers. Sustained drug release from buoyant systems improves gastric retention and reduces fluctuations in plasma drug concentration. The number of polymers, plasticizer-polymer ratio, and solvent used in formulation all influence buoyancy and drug release from the dosage form. Polycarbonate, HPMC, cellulose acetate, calcium alginate, Eudragit S, chitosan, and low molecular weight polymers are commonly used to create hollow microspheres. Several studies have shown that hollow microspheres can float continuously over the surface of an acidic dissolution media containing surfactant for more than 12 hours. The contract of the distinct advantage of having multiple unit systems. They are not provides good floating properties, making multiple unit systems. They are not provides good floating properties, making multiple unit systems. They are not provides good floating properties, making multiple unit systems, and their center hollow properties.

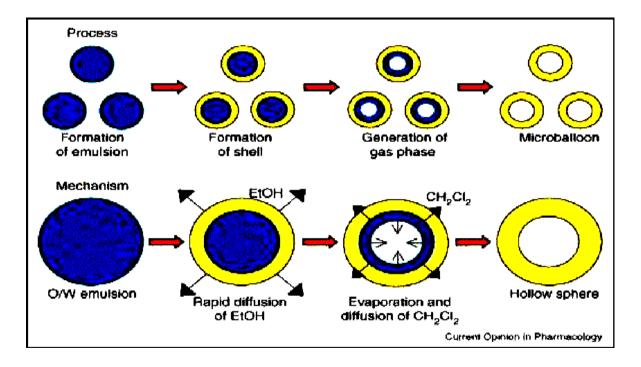


Figure no. 3: Hollow microspheres.

Mechanism of floating drug delivery system

FDDS have a bulk density less than gastric fluids and so remain buoyant in the stomach without affecting the gastric emptying rate for a prolonged period of time. While the system is floating on the gastric contents, the drug is released slowly at the desired rate. After release

of drug, the system is eliminated from the stomach. This results in an increased GRT and a better control of fluctuations in plasma drug concentrations. The floating sustained release dosage forms exhibit most of the characteristics of hydrophilic matrices and are known as 'hydrodynamically balanced systems' (HBS) since they are able to maintain their low apparent density, while the polymer hydrates and builds a gel like barrier at the outer surface. The drug is released progressively from the swollen matrix, as in the case of conventional hydrophilic matrices. These forms are expected to remain buoyant (3–4 h) in the gastric contents without affecting the intrinsic rate of emptying because their bulk density is lower than that of the gastric contents. Many studies have demonstrated the validity of the concept of buoyancy in terms of prolonged GRT of the floating forms, improved bioavailability of drugs and improved effects in clinical situations. The results obtained have also demonstrated that the presence of gastric contents is needed to allow the proper achieve-ment of the buoyancy retention effect. Among the different hydrocolloids recommended for floating form formulations, cellulose ether polymers are hydroxypropylmethylcellulose(HPMC). Fatty material the most popular, especially with a bulk density lower than one may be added to the formulation to decrease the water intake rate and increase buoyancy [18].

In parallel with formulation studies, investigations have been undertaken in animals and humans to evaluate the intragastric retention performance of coating forms. These assessments were carried out either indirectly through pharmacokinetic studies with a drug tracer, or directly by means of X-ray and gamma scintigraphic monitoring of the transit through the GI tract. When a floating capsule is administered to subjects who have consumed a fat and protein meal, it remains buoyant at the surface of the gastric contents in the upper part of the stomach and moves to the lower region progress-ively as the meal empties from the stomach. The reported gastric retention times range from 4 to 10 h. Pharmacokinetic and bioavailability evaluation studies confirm the favourable effect of this prolonged gastric residence time. FDDS have a bulk density less than gastric fluids and so remain buoyant in the stomach without affecting the gastric emptying rate for a prolonged period of time.

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This results in an increased GRT and a better control of fluctuations in plasma drug

concentrations.

Because FDDS have a lower bulk density than gastric fluids, they remain buoyant in the

stomach for an extended period without affecting the gastric emptying rate.

The drug is slowly released at the desired rate while the system is floating on the gastric

contents. The system is eliminated from the stomach after the drug is released.

As a result, GRT is increased and fluctuations in plasma drug concentrations are better

controlled.

The floating sustained-release dosage forms are known as "hydrodynamically balanced

systems" (HBS) because they can maintain their low apparent density while the polymer

hydrates and forms a gel-like barrier at the outer surface. They exhibit the majority of the

properties of hydrophilic matrices. The drug is gradually released from the swollen matrix,

similar to how it is done with traditional hydrophilic matrices.

Because their bulk density is lower than that of the gastric contents, these forms are expected

to remain buoyant in the gastric contents for 3-4 hours without affecting the intrinsic rate of

emptying.

Many studies have demonstrated the validity of the buoyancy concept in terms of prolonged

GRT of floating forms, improved drug bioavailability, and improved clinical effects.¹⁸

The obtained results also showed that the buoyancy retention effect cannot be properly

achieved without the presence of gastric contents. Cellulose ether polymers, especially

hydroxypropylmethylcellulose are the most widely used hydrocolloids among those

suggested for floating form formulations (HPMC).

To reduce the water intake rate and increase buoyancy, a fatty material with a bulk density

less than one may be added to the formulation.

Parallel to formulation studies, animal and human studies have been conducted to evaluate

the intragastric retention performance of floating forms.

These evaluations were conducted either indirectly through pharmacokinetic studies with a

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When a floating capsule is given to subjects after they have eaten a fat and protein meal, it remains buoyant at the surface of the gastric contents in the upper part of the stomach and gradually moves to the lower region as the meal empties from the stomach. Gastric retention times have been reported to range between 4 and 10 hours.

Studies on pharmacokinetics and bioavailability support the beneficial effect of this extended gastric residence time.¹⁹

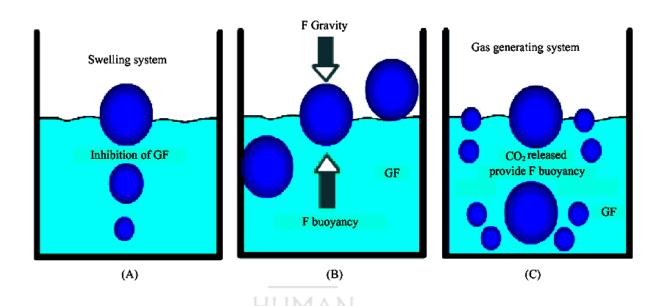


Figure no. 4: Mechanism of floating drug delivery system

Commonly Employed Polymers in Gastro retentive Floating Beads:

Sodium Alginate:

Sodium alginate is a natural polysaccharide as well as an anionic linear polymer with -1, 4-linked L-glucuronic acid and -1, 4-linked D-mannuronic acid residues randomly arranged along the chains. It is a stable gel that contains divalent cations, such as Ca2+, which are used for long-term drug release.²⁰

Alginate has high biocompatibility, mucoadhesion, biodegradability, and mild gelation conditions. Alginate beads are also used for floating drug delivery because they are stable in acidic media, preventing drug degradation in the acidic environment of the stomach.²¹

Chitosan:

Ishak RA and colleagues used the ionotropic gelation method to prepare metronidazole (MZ) in chitosan-treated alginate beads by using a factorially designed in which three viscosity-imparting polymers, MC, carbopol 934P, and -carrageenan Two concentrations of chitosan as an encapsulating polymer (0.2 and 0.4% w/v) and two concentrations of low-density magnesium stearate as a floating aid (2.5 and 5% w/w) were tested.²² Chitosan (HMW) was dissolved in ionic liquid EMIM Ac at B115 1C to make a solution containing 20 mg mL1. At room temperature, the chitosan/EMIM Ac solution was injected into ethanol using a syringe with a 0.33 mm inner-diameter needle. The injection rate was set to 10 ml/h1 and was controlled by a syringe pump (KD Scientific KDS 270, USA). The resulting beads were immersed in ethanol overnight and washed three times to completely remove the ionic liquid. The chitosan beads were then washed three times with water to allow solvent exchange between ethanol and water for 30 minutes. The hydrated chitosan beads were then freezedried to produce anhydrous chitosan beads.²³

Pectin

To form gel particles, an aqueous solution of 6% (w/v) pectin was introduced dropwise by a peristaltic pump through a plastic tubing (0.8 mm inner diameter) into a calcium chloride solution (CaCl 2 6%, w/v). The particles were immersed in a Ca2+ solution for 20 minutes. The beads were then rinsed with distilled water several times until neutral and dried at 37°C.²⁴With agitation, pectin was dissolved in water. The solutions (5% w/w) were extruded into 0.34 M calcium chloride with gentle agitation at room temperature using a nozzle with an inner diameter of 0.80 mm. The gel beads were allowed to stand in the solution for 20 minutes before being separated and washed with distilled water. The beads were either airdried at 37 degrees Celsius for 12 hours or freeze-dried.²⁵

Guar gum

For the experiments, 250 milligrams of guar gum were dissolved in 50 mL of distilled water. Guar gum beads were created by syringing guar gum solution into 100 ml of 0.5 M sodium tetraborate solution (pH adjusted to 7.2).²⁶

Floating beads can be prepared by:

Emulsion gelation method

The emulsion gelation method is used to create oil-entrapped gel beads that can float in

gastric conditions. Gel beads containing an effervescent agent or edible oil are made by

gently mixing or homogenizing an oil phase and a water phase containing pectin or casein,

and then extruding into a calcium chloride solution with gentle agitation at room temperature.

The prepared gel beads are then separated, washed with distilled water, and dried for 12

hours at 370 degrees Celsius.²⁷

Example: oil-entrapped calcium pectinate gel beads.

Ionic Gelation Method

Ionotropic gelation is based on polyelectrolytes' ability to crosslink in the presence of counter

ions (Giri et al., 2013d). The ionic gelation method is most commonly used to prepare

alginate nanoparticles. Alginate-chitosan nanoparticles were created in two steps, beginning

with the ionotropic gelation of polyanion with calcium chloride and ending with polycationic

crosslinking (Sarmento et al., 2007). In a gastric pH environment, the nanoparticles retained

approximately 50% of the protein for up to 24 hours. Under intestinal pH conditions, the

release was close to 75%.

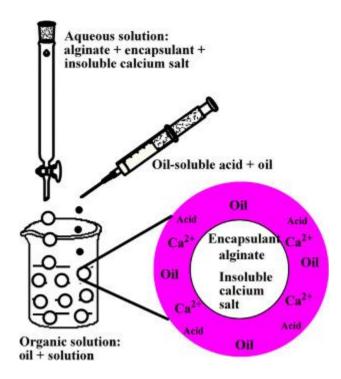
particles were created using a modified emulsification/internal gelation method. This method

of preparing alginate nanoparticles does not require any special equipment and can be done at

room temperature. The main challenge of this method is the nanoparticle-washing step to

remove the residual oil droplets.²⁸

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Evaluation parameters of floating beads:

Study of size and morphology of emulsion gel beads

A screw gauge was used to determine the diameter of the beads. For this purpose, 20 dried beads were chosen at random from each batch, and the mean diameter was determined using a screw gauge. The smallest screw gauge count was 0.005 mm. Each batch's dried beads were noted for their colour and shape.²⁹

DRUG CONTENT:

UV-Spectrophotometry was used to determine the drug content of prepared floating beads.

A precisely weighed quantity of floating beads was taken and dissolved in 100 ml of 0.1N HCl, 1ml of the solution was diluted to 10 ml, and the drug content was estimated using UV at 314nm.³⁰

Swelling studies:

Swelling characteristics of beads were investigated. Only Those batches with high drug concentrations were chosen. More than 50% efficiency in content and entrapment. A sample of drug-loaded beads was taken and weighed and inserted into the wire basket of the USP dissolution apparatus II. The basket containing beads was placed in a beaker containing 100

ml 0.1 N HCl (pH 1.2) kept at 370C. The beads were removed at regular intervals.

predetermined intervals and weighed.³¹

In vitro drug release studies:

The drug was dissolved in vitro for 300 minutes using a USP Type II dissolution apparatus

containing 900 ml of simulated gastric fluid (0,1 N HCl pH 1.2) maintained at 37 0.5°C and

speed At 50 rpm, the agitation is strong. Aliquots (5 ml) were collected at regular intervals

and replaced with fresh dissolution medium. The collected samples were analyzed with a UV

spectrophotometer at a maximum wavelength of 228 nm (UV-1800, Shimadzu). The research

was done in triplicate. The percentage of drugs released at different time intervals was

calculated and plotted versus time.³²

CONCLUSION:

Formulation of FDDS is an efficient and potential approach for gastric retention of dosage

forms to improve bioavailability and also to achieve controlled release of dosage form. The

most important criteria which has to be looked into for the formulation of a FDDS is that the

density of the dosage form should be less than that of gastric fluid by this method we can thus

reduce the dosing frequency and improve patient compliance. In this review, we discussed in

details regarding floating drug delivery system, its classification and its mechanism,

commonly employed polymers in gastro retentive floating beads, methods of preparation and

evaluation of floating beads.

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CONFLICTS OF INTEREST:

The authors declare that there is no conflict of interest.

REFERENCES:

1. Khalifa MY, Saleem MA, HuzaifaPatel SS. Preparation and evaluation of gastroretentive hydrogel beads of Cefdinir by ionotropic gelation method. Asian Journal of Pharmacy and Pharmacology. 2019;5(4):786-792.

2. Ratnaparkhi MP, Gupta Jyoti P. Sustained release oral drug delivery system-an overview. Terminology.

2013;3(4):10-22270.

3. Vallamsetti SD, Nimisha M. Formulation and Evaluation of Floating Beads of Famotidine. International

Journal of Pharmaceutical Sciences Review and Research. 2014;24(1):192-198.

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- 4. Niharika MG, Krishnamoorthy K, Akkala M. Overview on floating drug delivery system. Int J Appl Pharm. 2018;10(6):65-71.
- 5. Kajale AD, Chandewar AV. Formulation and evaluation of oral floating beads of tramodol hydrochloride. Journal of Drug Delivery and Therapeutics. 2016;6(4):7-16.
- 6. Kumar R, Chandra A, Saloni S, Gautam PK. Advanced multiple unit controlled release floating beads: A review. World J Pharm Res. 2017;6(15):238-259.
- 7. Kajale AD, Chandewar AV. Formulation and evaluation of oral floating beads of tramodol 5hydrochloride. Journal of Drug Delivery and Therapeutics. 2016;6(4):7-16.
- 8. Gupta P, Gnanarajan PK, Kothiyal P. Floating drug delivery system: a review. International Journal of Pharma Research & Review. 2015;4(8):37-44.
- 9. Garg RG, Gupta GD. Progress in controlled gastroretentive delivery systems. Tropical journal of pharmaceutical research. 2008;7(3):1055-66.
- 10. Gopalakrishnan S, Chenthilnathan A. Floating drug delivery systems: A Review. Journal of Pharmaceutical Science and Technology. 2011;3(2):548-54.
- 11. Sangekar S, Vadino WA, Chaudry I, Parr A, Beihn R. Evaluation of the effect of food and specific gravity of tablets on gastric retention time. International journal of pharmaceutics. 1987;35(3):187-191.
- 12. Choudhury A, Renthlei L, Dewan M, Ahmed R, Barakoti H, Dey BK. Floating drug delivery system: an outlook. Journal of applied pharmaceutical research. 2019;7(3):01-8.
- 13. Nadigoti J. Floating drug delivery systems. International Journal of Pharmaceutical Sciences and Nanotechnology. 2009;2(3):595-604.
- 14. Kumari B. Recent Development inFloating Drug Delivery System: A Review. Asian Journal of Pharmacy and Pharmacology. 2018;4(2):131-9.
- 15. Kumar K, Rai AK. Development and evaluation of floating microspheres of curcumin. Tropical journal of pharmaceutical research. 2012;11(5):713-9.
- 16. Gholap SB, Banarjee SK, Gaikwad DD, Jadhav SL, Thorat RM. Hollow microspheres: A review. Int J Pharm Sci Rev Res 2010;1:74-9.
- 17. Kurrey A, Suresh P, Singh M. Hollow microspheres as a drug carrier: An overview of fabrication and in vivo characterization techniques. Chronicles of Young Scientists. 2014;5(1):1-10.
- 18. Reddy LH, Murthy RS. Floating dosage systems in drug delivery. Critical Reviews™ in Therapeutic Drug Carrier Systems. 2002;19(6):553-85.
- 19. Shaha SH, Patel JK, Pundarikakshudu K, Patel NV. An overview of a gastro-retentive floating drug delivery system. Asian journal of pharmaceutical sciences. 2009;4(1):65-80.
- 20. Rakesh P, Vipin K, Kanchan K. Alginate Beads Prepared By Ionotropic Gelation Technique: Formulation Design, Research Journal of Chemical Sciences 2015; 5(7):45-47.
- 21. Ranvirsingh TA, Basavaraj BV, Bharath S, Deveswaran R, Madhavan V. Formulation and Evaluation of Floating Alginate Beads of an AntiUlcer Drug, International Journal of Pharmaceutical Sciences Review and Research 2013;21(2):120-24.
- 22. Ishak RA, Awad GA, Mortada ND, Nour SA. Preparation, in vitro and in vivo evaluation of stomach-specific metronidazole-loaded alginate beads as local anti-Helicobacter pylori therapy. Journal of controlled release. 2007;119(2):207-14.
- 23. Li B, Wang J, Gui Q, Yang H. Continuous production of uniform chitosan beads as hemostatic dressings by a facile flow injection method. Journal of Materials Chemistry B. 2020;8(35):7941-6.
- 24. Cabrera JC, Cambier P, Cutsem P. Drug encapsulation in pectin hydrogel beads—A systematic study of simulated digestion media. International Journal of Pharmacy and Pharmaceutical Sciences. 2011;3(5):292-9.
- 25. Sriamornsak P, Sungthongjeen S, Puttipipatkhachorn S. Use of pectin as a carrier for intragastric floating drug delivery: Carbonate salt contained beads. Carbohydrate Polymers. 2007;67(3):436-45.
- 26. Roy I, Sardar M, Gupta MN. Cross-linked alginate—guar gum beads as fluidized bed affinity media for purification of jacalin. Biochemical Engineering Journal. 2005;23(3):193-8.
- 27. Kumar R, Chandra A, Saloni S, Gautam PK. Advanced multiple unit controlled release floating beads: A review. World J Pharm Res. 2017;6:238-59.
- 28. Giri T. Nanoarchitectured Polysaccharide-Based Drug Carrier for Ocular. Amsterdam, The Netherlands: Elsevier Science; 2016 Jul 12.

- 29. Jaiswal D, Bhattacharya A, Yadav IK, Singh HP, Chandra D, Jain DA. Formulation and evaluation of oil entrapped floating alginate beads of ranitidine hydrochloride. International Journal of Pharmacy and Pharmaceutical Sciences. 2009;1(3):128-40.
- 30. Kumar Y, Sheeba FR, B Likitha, Mutta S, Keerthy HS, HM Ashvini. Formulation and Evaluation of Floating Beads of Nizatidine. International Journal of Pharmaceutical Research and Applications. 2022;7(3):449-457.
- 31. Khan AD, Bajpai M. Formulation and Evaluation of Floating beads of Verapamil hydrochloride. International Journal of PharmTech Research. 2011;3(3):1537-46.
- 32. Hendrika Y, Reveny J, Sumaiyah S. Formulation and in vitro evaluation of gastroretentive floating beads of amoxicillin using pectin from banana peel (Musa Balbisiana ABB). Asian J Pharm Clin Res. 2018;11(3):72-7.

