



IJPPR

INTERNATIONAL JOURNAL OF PHARMACY & PHARMACEUTICAL RESEARCH  
An official Publication of Human Journals

ISSN 2349-7203




Human Journals

Review Article


April 2024 Vol.:30, Issue:4

© All rights are reserved by Manoj A et al.

## A Review On phytochemicals and Pharmacological Activity of Medicinal Plant *Cyperus rotundus*



IJPPR  
INTERNATIONAL JOURNAL OF PHARMACY & PHARMACEUTICAL RESEARCH  
An official Publication of Human Journals



ISSN 2349-7203

**Manoj A\*, Kumudhaveni B, Abirami R, Vanitha N, Pooja S, Radha R**

*Department of Pharmacognosy, College of Pharmacy  
Madras Medical College, Chennai, India.*

**Submitted:** 25 March 2024  
**Accepted:** 31 March 2024  
**Published:** 30 April 2024

**Keywords:** *Cyperus rotundus*, Motha, Traditional Uses, Phytochemical study, Pharmacological action.

### ABSTRACT

This innovative endeavor is to offer comprehensive details about *Cyperus rotundus*, a member of the Cyperaceous<sup>1</sup> family and referred to as "Motha" or "Mutha." In the herbal medicine system, this plant is widely renowned for its remarkable ability to treat a wide range of illnesses. The herb has been shown to have both pharmacological and biological properties, such as anti-inflammatory, antipyretic, anti-amoebic, anti-emetic, hypolipidemic, wound-healing, anticancer, coronary vasodilator, anti-alcoholic, anti-ulcer, ovicidal and larvicidal effects, antipyretic, antiviral, anti-malarial, anti-convulsant, central nervous system influence, influence on platelet function, antioxidant activity, and anti-diarrheal properties. The current review paper emphasizes the importance of the following: species, botanical name, phytocomponents, chemical structures, microscopic traits, traditional applications, life cycle, and pharmaceutical efficacy. Future scholars will find significant value in this vast material.



HUMAN JOURNALS

[ijppr.humanjournals.com](http://ijppr.humanjournals.com)

## INTRODUCTION

Herbal medicine is the oldest kind of medicine that is currently recognized. It formed the foundation of numerous ancient cultures and is still the most widely utilized kind of medicine worldwide<sup>[1]</sup>.

First of all by World Health Organization's estimates, around 80% of the global population mostly obtains medical care from conventional medicines. For instance, in the developed world, between 1959 and 1980, plant-based extracts or active compounds derived from higher plants were included in 25% of all medication filled at neighborhood pharmacies in the USA<sup>[2]</sup>.

### Taxonomic classification

Kingdom : Plantae  
Subkingdom : Tracheobionta  
Superdivision : Spermatophyta  
Division : Magnoliophyta  
Class : Liliopsida  
Subclass : Commelinidae  
Order : Cyperales  
Family : Cyperaceae  
Genus : *Cyperus* L  
Species : *Cyperus rotundus*<sup>[1&3]</sup>.

### Common names

In India, the herb *C. rotundus*, commonly referred to as “Motha” or “Mutha,”<sup>[4]</sup>

English: Coco-grass, red nut sedge Arabic: Sa'ed; Chinese: Suo cao, Xiang fu zi; French: Souchetron; German: Knolliges Zypergras; Italian: Zigoloinfestante; Japanese: Hamasuge; Korean: Hyangbuja; Portuguese: Alho-bravo, Capim-alho, Capim-dandá, Tiririca, Tiririca-vermelha; Spanish: Castañuela, Cipero, Coquito, Juncia real; Swedish: Nötåg<sup>[5]</sup>.

### Synonyms

Ground-almond, Java-grass, Nut sedge, Nut-grass, purple nut, Sedge, and Purple nut-grass<sup>[1]</sup>.

## Distribution

Asia: China, Japan, Korea, Taiwan, India, Nepal; Pakistan, Sri Lanka, Myanmar; Thailand, Vietnam, Indonesia, Malaysia, Philippines; Europe; Pacific; the United States of America; and Southern America were among the regions it was dispersed in<sup>[12]</sup>.

## Life cycle

Despite some small differences, there are significant similarities in the life cycle cycles of purple and yellow nutsedge species. Plants of both species are vegetative, meaning they produce a complex underground network of basal bulbs, rhizomes, and tubers. The basal bulbs are the building bricks of vegetative growth because they contain meristems that are responsible for leaves, rhizomes, roots, and flower stalks. Similar to yearly seeds, the tubers are the primary mechanism of dispersal and feature latent buds<sup>[6]</sup>.

## Characteristics of *C. rotundus*

Growing from a rhizome, the common perennial *C. rotundus* can reach heights of between 15 and 30 centimeters as well as 50 cm in tropical and subtropical areas. It produces tiny, nut-shaped tubers at the center of its stem (Figures 1 and 2). The leaves among the plant are upright and oriented in three distinct directions<sup>[11]</sup>.



Figure: 1



Figure: 2

Areal part of *C. rotundus*<sup>[2&5]</sup>.

## Macroscopy

**Stem:** These leaves are usually taller (20–100 cm) and have a smooth, triangular cross form. Usually, they are longer than the base leaves. These culms align with the blossoming axis. At the increased base of the plant, a thickening layer called a basal bulb grows.

**Leaf:** The basal leaves are linear, pointed, and directed in three directions. They vary in length from 10 to 50 cm. Their source is the upright fertile stem that arises from very compact nodes in three-row basal clusters.

**Glabrous blade with border and section that are shiny and dark green, folded twice.**

**Fruit-Nuts:** elliptical or oblong-ovate, 1.5 mm long, 0.8 mm broad, three-angled, and stigma-topped; olive-gray to brown or black in hue.



Figure 3: Nut-shaped tubers of *C. rotundus*<sup>[2]</sup>.

## Microscopic features of *C. rotundus*

The microscopic examination of *C. rotundus* root structures, as seen in Figure 4, reveals an epidermis composed of parenchymatous cells with a distinctive brownish tint. The cortex is made up of parenchymatous cells, while the hypodermis is made up of 1-2 distinct layers of thickly walled cells. The internal portion is parenchymatous and features wide intercellular gaps, whereas the exterior portion is compacted. Sharma and Singh (2011) report that brownish oleo resinous material and additional starch grains are present in particular cells in the frontal area<sup>[13-18]</sup>.

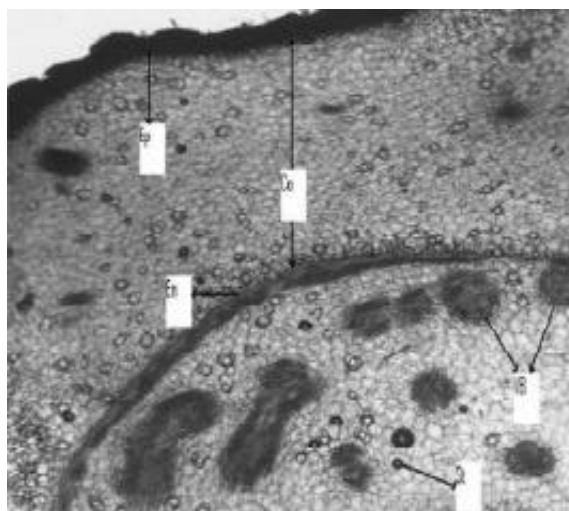


Figure 4: T.S of *C. rotundus*<sup>[2]</sup>.

A great deal of dyed cells wrapped into a reddish-brown oleo-resin material, basic round or expanded starch grains, current passing through the cortex and stele, vascular bundles encircled by fiber bundle sheaths, vessels spiraling to simple pithed, scattered in this region, vessels that are the xylem vessels' lignified secondary wall thickenings could be extended, oval, or simply round <sup>[13-18]</sup>.

### Traditional uses

The herb *Cyperus rotundus* was used for intestinal cramps, problems with the stomach, nausea, vomiting, parasitic infections of the intestines, food poisoning, indigestion, and colon irritability. It was also used to treat respiratory conditions like amenorrhea and dysmenorrhea, fevers, bruising, and carbuncles; malaria; cough, bronchitis, renal and vesical calculi; urinary tenesmus; insect bites; loss of memory; infertility; cervical cancer; and menstrual disorders. The astringent, diaphoretic, diuretic, analgesic, antispasmodic, aromatic, carminative, antitussive, emmenagogue, litholytic, sedative, stimulant, stomachic, vermifuge, tonic, and antibacterial qualities of *Cyperus rotundus* rhizomes were recorded throughout the literature on Ayurveda <sup>[13, 18-22]</sup>.

### Chemical constituents

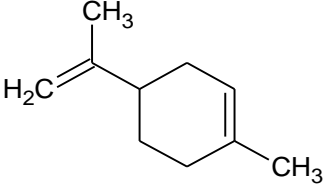
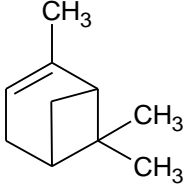
Phytochemical examinations revealed that *Cyperus rotundus* contained a wide range of secondary metabolites, such as flavonoids, tannins, glycosides, furochromones, monoterpenes, sesquiterpenes, sitosterol, alkaloids, saponins, terpenoids, essential oils, starch, carbohydrates, proteins, and amino acids as well <sup>[23-27]</sup>.

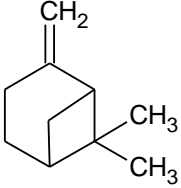
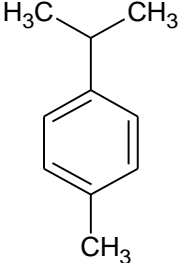
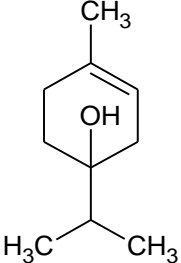
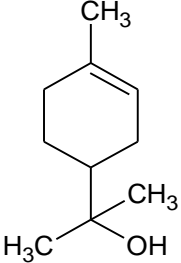
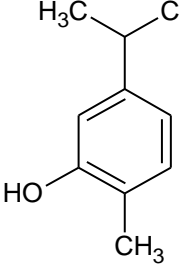
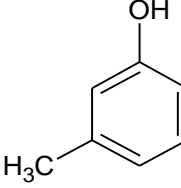
Quinones, phenolic acids: salicylic acid, protocatechuic acid, caffeic acid, and p coumaric acid; flavonoids: visnagin, khellin, ammiol, isorhamnetin, and triclin; saponins; alkaloids; coumarins; and steroids: steroidal glycoside, sitosterol-(6'-hentriacontanoyl)- $\beta$ -D-galactopyranoside<sup>[28]</sup>.

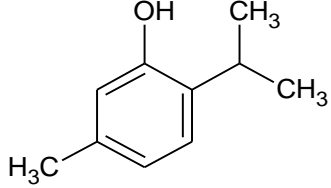
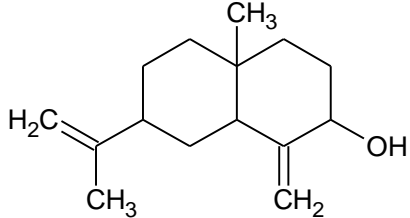
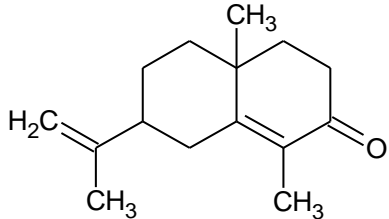
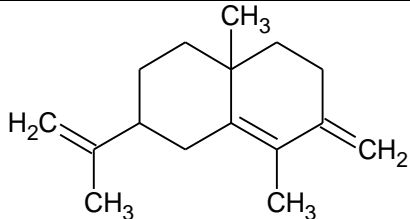
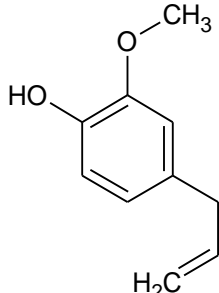
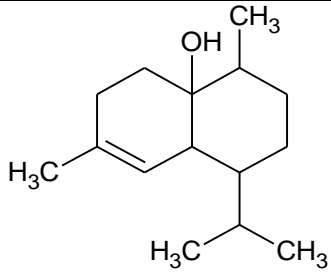
The concentration of essential oils in *Cyperus rotundus* tubers is 0.19%, and the tubers have a specific gravity of 0.9689 and a refractive index of 1.54511. Fifty two compounds were acquired from the Egyptian plant *Cyperus rotundus*. The main components of the oil were trans-pinocarveol (7.92%), cyperene (7.83%), (+) oxo- $\alpha$  ylangene (9.35%), and (+)  $\alpha$ -cyperone (9.07%)<sup>[29,30]</sup>.

However, 0.2% (w/w) of the tuber of *Cyperus rotundus*, a plant that grows naturally in the Isfahan area of Iran, was discovered to hold onto essential oils. The essential oil contained 60 compounds that were found to exist naturally. Sesquiterpenes accounted for most of the compounds in the oil. The primary components of the oil were Cyrene (16.9%), caryophyllene oxide (8.9%),  $\alpha$ -longipinane (8.4%), and  $\beta$ -selinene (6.6%)<sup>[31]</sup>.

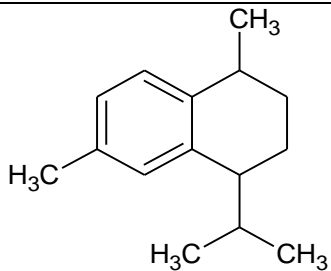
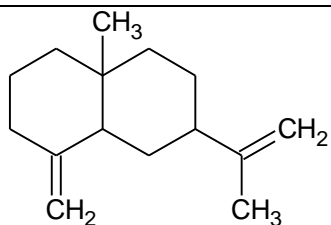
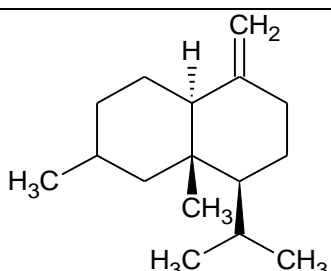
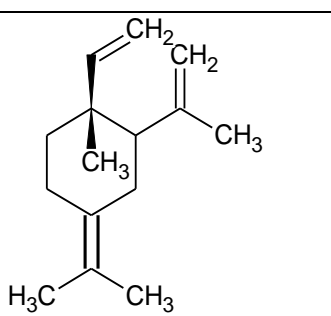
Comparing ethanol extracts (6.44–13.77 mg CE – (Catechin equivalents)/g of dry matter) with the methanol extract from *Cyperus rotundus* revealed higher total flavonoid contents (8.15–18.25 mg CE/g of dry matter). In addition, *Cyperus rotundus* extracted with methanol had higher total phenolic contents (27.40–37.85 mg GAE/g of dry matter) than extracts made with ethanol (25.21–30.23 mg GAE/g of dry matter)<sup>[32, 33]</sup>.

1.		Limonene
2.		$\alpha$ -Pinene

3.		$\beta$ -Pinene
4.		p-Cynene
5.		4-Terpinol
6.		$\alpha$ -Terpinol
7.		Carvacrol
8.		Thymol

9		o-Cadinol
10.		Isocyperol
11.		o-Cyperone
12.		Cyperol
13.		Eugenol
14.		Cubenol



15.		Trans-Calamenene
16.		$\beta$ Selinone
17.		$\gamma$ Cadinene
18.		$\gamma$ Elenone

### Pharmacological activity of *Cyperus rotundas*

#### Anti-inflammatory activity

Gupta M et al. found that 400–600 mg/kg methanol and chloroform extract of *Cyperus rotundus* rhizomes was effective in treating Wistar rats' carrageenan-induced paw edema. Twelve when taken orally at levels between 300 and 500 mg/kg, the herbal compound showed anti-inflammatory characteristics, accordance to a study by Ahamad M et al<sup>[34-37]</sup>.

### **Antipyretic activity**

Pyrexia was seen in albino rats that were subcutaneously administered with a solution of dried Brewer's yeast in gum acacia in regular saline. Versus these rats, the alcoholic extract exhibited a very significant ( $P < 0.001$ ) antipyretic effectiveness. When an extract of petroleum ether was administered to the same animal model, a particular portion that was made by chromatography had a significant antipyretic activity that was similar to acetylsalicylic acid<sup>[38-40]</sup>.

### **Anti-amoebic activity**

Additionally, In vitro, investigations revealed that the whole *Cyperus rotundus* plant had greater anti-amoebic efficacy towards *Entamoeba histolytica* trophozoites. After 96 hours of additional research, the entire plant *Cyperus rotundus* extract of ethanol showed 100% inhibition at 500 µg/ml<sup>[41, 42]</sup>.

### **Anti-emetic activity**

It has been demonstrated that the ethanol extract of *C. rotundus* at a concentration of 128.1± 11.6 mg/kg prevented 50% of dogs from vomiting produced by apomorphine<sup>[38-44]</sup>.

### **Anti-hyperlipidemic activity**

In this investigation, a high-fat diet was used to produce hyperlipidemia since it is a helpful tool for evaluating drugs that obstruct the absorption, breakdown, and excretion of cholesterol. Serum levels of low-density lipoprotein (LDL), triglycerides (TG), and total cholesterol (TC) increased significantly ( $P < 0.05$ ) after feeding a high-fat meal compared to the baseline. High-density lipoprotein (HDL) levels increased while on a high-fat diet, although this effect was not shown to be statistically significant. After 15 days of intervention, treatment with the standards and various extract dosages resulted in a statistically significant ( $P < 0.05$ ) decrease in blood levels of TC, LDL, TG, and HDL in the current study<sup>[11, 60]</sup>.

Based on a study by Okwu1 et al., bioactive substances in *Cyperus rotundus* rhizome are having hypolipidemic potentials. An alcoholic extract of *Cyperus rotundus* at dose levels of 70 mg/kg/day, 140 mg/kg/day, and 280 mg/kg/day showed higher improvement in cholesterol levels after taking medication for 15 days, according to Chandra et al<sup>[41, 45]</sup>. In an alloxan-induced diabetes model, Sprague Dawley rats demonstrated antihyperglycemic

effects from the various hydro ethanol extract fractions (methanol, acetone, chloroform, and ethyl acetate). Because of the high phenolic content, it has an antioxidant characteristic that contributes to its anti-hyperglycemic activity<sup>[3, 45, 52]</sup>.

### **Wound healing property**

*Cyperus rotundus*'s active ingredients appear to be what give the herb its curative properties; they both quicken the healing process and give the healed wound its tearing strength. Based on the findings of the research, it is possible to conclude that the ointment containing *Cyperus rotundus* extract has potent wound-healing qualities at all tested doses<sup>[41, 47-49]</sup>.

### **Anticancer activity**

Anti-tumor the ethanolic extract of *C. rotundus* has been demonstrated to have only weak to moderate anticancer activity (LC50=2.528-4.939 mg/ml estimated from dose-dependent cell death) in a study that used neuro-2a cells for testing plants with tumoricidal capabilities. Subsequent research proved that *C. rotundus* essential oil was effective in combating the leukemia cell line L1210. This result was linked to a significant increase in apoptotic fragmentation of DNA<sup>[38, 50]</sup>.

### **Coronary vasodilator activity**

A water-based extract of a *C. rotundus* rhizome given IV has positive results in cats, rabbits, and frogs<sup>[3, 51]</sup>.

### **Anti-alcoholic activity**

The fermenting tuber of *C. rotundus* showed effectiveness in the rats' diet when given at a level of 5.0 ml/animal. The effects of alcohol on the electroencephalogram (ECG), liver fat accumulation, symptoms of bleeding, demyelination, and spongiosis were reversed by taking a daily dose for ninety days<sup>[3, 53]</sup>.

### **Anti-ulcer activity**

Utilizing a 1.25 g/kg dose of *Cyperus rotundus* tuber powder, the ulcer index was considerably decreased in the histamine-induced stomach ulcer approach. This research suggests that this substance may also have anti-ulcer qualities<sup>[41, 42]</sup>.

The powdered rhizome of *C. rotundus* has ulcer-prevention properties. Two distinct models of animals were used in the investigation. Histamine (50 mg base i.p) accustomed to causing stomach ulcers in guinea pigs, whereas aspirin (500 mg/kg orally) accustomed to causing ulcers in albino rats. Oral administration of *C. rotundus* powdered rhizome was administered 45 minutes before the histamine and one hour before the aspirin. In every instance, *C. rotundus* significantly reduced the ulceration index and yielded results equivalent to the individuals of the reference drug ranitidine. Because *C. rotundus* possesses a powerful antioxidant profile, it offers antiulcer properties<sup>[3]</sup>.

### **Ovicidal and larvicidal effects**

Investigations were conducted into the ovicidal and larvicidal effects of *C. rotundus* essential oils. Research has been done on the eggs and larvae of *Aedes albopictus* (Skuse). The eggs and fourth instar larvae were treated with the essential oil for an entire day, at concentrations ranging from 5 to 150 ppm. The essential oil of *C. rotundus* had a potent larvicidal and ovicidal action, as demonstrated by its half-maximal effective concentration (EC<sub>50</sub>) of 5 ppm and fatal dosage (LD<sub>50</sub>) of 20 parts per million (ppm)<sup>[3, 54]</sup>.

### **Anti-pyretic activity**

The alcohol extract of *C. rotundus* efficiently avoids pyrexia in albino rats who have received a subcutaneous injection of a dehydrated yeast solution acacia in a saline solution. The *C. rotundus* rhizome's methanol extract was regularly isolated using column chromatography into its soluble fractions so that the structure of the separated components could be examined using UV, IR, H-NMR, and MS spectra (2016). After injecting a solution of dried Brewer's yeast in gum acacia in normal saline subcutaneously, albino rats were effectively treated with the alcohol extract of *C. rotundus*. In the same animal model, the extract was found to have an antipyretic effect comparable to acetylsalicylic acid<sup>[3, 39, 40, 55]</sup>.

### **Anti-fungal activity**

Rhizome was found to be effective towards *Phytophthora capsid*, *Colletotrichum chardonianum*, and *Sclerotinia scleroliorum* in an agar plate test. On an agar plate, *Helminthosporium turcicum* did not react to an undiluted fresh shoot water extract<sup>[3, 56]</sup>.

### **Anti-viral activity**

The hydroalcoholic extract of *C. rotundus* and the antiviral properties of forty-one Egyptian medicinal plants were tested. Three viruses were tested against the extract (vesicular stomatitis virus): HSV1, POLIO (poliomyelitis-1 virus), and VSV (herpes simplex-1 virus). The end-point titration approach was employed to determine the antiviral activity. *C. rotundus* showed virucidal effects from HSV<sup>[3, 57]</sup>.

### **Anti-malarial activity**

Through activity-guided research, compounds such as 4,7-dimethyl-1-tetralone, 10,12-peroxycalamenene, caryophyllene oxide, and patchoulene were isolated from *C. rotundus* tubers. The new endoperoxide sesquiterpene, 10, 12-peroxycalamenene, has the highest impact at EC<sub>50</sub> 2.33 × 10<sup>6</sup> M. These compounds have anti-malarial activity within the range of EC<sub>50</sub> 10<sup>-4</sup>-10<sup>-6</sup> M<sup>[11, 58, 59]</sup>.

### **Anti-convulsant activity**

Given the significant prevalence of epilepsy in human civilization and the lack of effective treatments, the anticonvulsant activity of *C. rotundus* extract was investigated experimentally in the current study. Six groups of sixty male mice each were created by random selection: (1) control group, (2) group mice that had been given pentylentetrazole (PTZ), (3) group is a positive control group that was given valproate (100 mg/kg) as an anticonvulsant medication, and (4-6) a group that was given *C. rotundus* rhizome extract at three different doses of 100, 200, and 400 mg/kg intra-peritoneally<sup>[11]</sup>.

Eleven intraperitoneal (i.p) administrations of PTZ (35 mg/kg; 48 h apart) were administered to all animals, excluding the control group. Every group—aside from the control group was evaluated for the PTZ challenge dosage (75 mg/kg) after the twelfth injection. After the PTZ injection, the seizure phases (0–6) that were displayed were watched and recorded for 30 minutes. After the mice's brains were completely removed, the amounts of nitric oxide (NO), superoxide dismutase (SOD), and malondialdehyde (MDA) in the brain tissues were measured. Data research revealed that *C. rotundus* hydroalcoholic extract might lessen seizure duration and intensity. Moreover, the extract may raise SOD and NO levels while lowering MDA levels in mice's brains. The conclusion is that the *C. rotundus* rhizome extract may have had a strong antiepileptic impact due to its antioxidant qualities<sup>[11]</sup>.

### Central nervous effect

In several experiments, the ethanolic extract of *Cyperus rotundus* showed strong tranquilizing effects. In rats, it eliminated the conditioned avoidance response, decreased spontaneous motor activity, increased pentobarbital narcosis, and disrupted motor coordination.<sup>[61]</sup> The neuropharmacological effects of 300 and 500 mg/kg of *Cyperus rotundus* extract were studied using open field, head dip, rearing traction, and forced swimming tests. All tests on the crude extract revealed a small reduction, and it also demonstrated a minor muscle relaxant action<sup>[62]</sup>. The ethanol extract of *Cyperus rotundus* showed CNS depressing action, according to behavioral investigations conducted on mice. In a dose-dependent way, the ethanol extract of *Cyperus rotundus* markedly increased the amount of time that mice slept while hypnotized with conventional drugs (pentobarbitone sodium, diazepam, and meprobamate)<sup>[18]</sup>.

### Effect on platelet function

The antiplatelet properties of *Cyperus rotundus* ethanolic extract (CRE) and eight of its component compounds were assessed by measuring the impact on mice tail bleeding times and rat platelet aggregations both In vitro and In vivo. In the In vitro platelet aggregation study, CRE shown notable and concentration-dependent inhibitory effects on platelet aggregation produced by thrombin, collagen, and arachidonic acid (AA). It was discovered that among its eight constituents, (+) nootkatone has the most inhibitory action against platelet aggregation produced by thrombin, collagen, and AA. Furthermore, mice treated with CRE and (+) nootkatone showed considerably longer bleeding times. Moreover, the ex vivo rat platelet aggregation was significantly inhibited by (+)-nootkatone. *Cyperus rotundus* can enhance all haemorrhoeological indices, including erythrocyte electrophoresis, plasma-specific viscosity, whole blood-specific viscosity, and others, according to research on the plant's impact on hemorrhage alterations in normal rats<sup>[18, 63]</sup>.

### Anti-oxidant activity

In vitro tests using reactive oxygen species and free radicals were used to assess the antioxidant activity of *Cyperus rotundus* rhizomes extract (CRRE), and IC<sub>50</sub> (Inhibitory concentration) values were found. With respect to superoxide anion radicals, hydroxyl radicals, nitric oxide radicals, hydrogen peroxide, and the ability of metals to chelate and reduce, CRRE demonstrated its scavenging function in a concentration-dependent way. Using both young and old rat brain mitochondria, the extract was examined in more detailed for the

lipid peroxidation assay by thiobarbituric acid-reactive compounds (TBARS). Additionally, in a concentration-dependent manner, the extract was successful in reducing mitochondrial lipid peroxidation caused by FeSO<sub>4</sub> ascorbate. The current study's findings suggest that the rhizomes of *C. rotundus* extract might be a useful natural antioxidant source [64, 65].

### Anti-diarrhoeal

In mice with castor oil-induced diarrhea, the methanol extract of *Cyperus rotundus* rhizome, administered orally at dosages of 250 and 500 mg/kg bowl, had strong antidiarrheal action. Following testing at 250 mg/kg, it was discovered that the residual methanol fraction (RMF) and petroleum ether fraction both retained their activity, with the RMF being significantly more active than the control group. There was no evidence of antidiarrheal activity in the ethyl acetate fraction [65, 66].

### CONCLUSION

Due to its and potency, *Cyperus rotundus* is a potential medicinal plant. This study covers its life cycle, characteristics, microscopic features, chemical composition, pharmacological properties, and medicinal benefits.

### REFERENCES

1. Al-Snafi AE. A review on *Cyperus rotundus* A potential medicinal plant. IOSR Journal of Pharmacy. 2016;6 (7):32-48.
2. Jeurkar MM, Kosalge SB, Sheikh NW, Telrandhe UB. *Cyperus rotundus* L.: Phytochemistry and pharmacological activities. Annals of Phytomedicine. 2022;11(2):186-96.
3. Barai M. Motha (*Cyperus rotundus* Linn.): An Ayurvedic Perspective. Rasamruta, 9:18 September 2017:01-18
4. David WH, Vernon VV, Jason AF. Purple nutsedge, *Cyperus rotundus* L. Florida (U.S.A): Institute of Food and Agricultural Sciences, University of Florida; 2012. p. 02-15.
5. Smith J. USDA, ARS, National Genetic Resources Program. Germplasm Resources Information Network–(GRIN). National Germplasm Resources Laboratory, Beltsville. 2010
6. Stoller, E. W. (1973). "Effect of minimum soil temperature on differential distribution of *Cyperus rotundus* and *C. esculentus* in the United States.". Weed Research 13(2): 209-217.
7. Global Invasive Species Database, *Cyperus rotundus*, <http://www.issg.org/database/species/ecology.asp?si=1448> [27 April 2009]
8. Wills GD. Description of purple and yellow nutsedge (*Cyperus rotundus* and *C. esculentus*). Weed Technology 1987; 1: 2-9.
9. Riemens MM, Weide RY, Runia WT. Nutsedge: biology and control of *Cyperus rotundus* and *Cyperus esculentus*-review of a literature survey. Wageningen, Plant Research International, 35p. 2008.
10. Scheepens, P. C. and A. Hoogerbrugge (1991). "Host specificity of *Puccinia canaliculata*, a potential biocontrol agent for *Cyperus esculentus*." Netherlands Journal of Plant Pathology 97: 245-250.
11. Imam, H.; Zarnigar.; Sofi, G.; Sheikh, A. and Lone, A. The incredible benefits of Nagarmotha (*Cyperus rotundus*). Int. J. Nutr. Pharmacol. Neurol. Dis., 4(1) 2014:23-27.



12. USDA, ARS, National Genetic Resources Program. Germplasm Resources Information Network- (GRIN). National Germplasm Resources Laboratory, Beltsville, Maryland. URL: <http://www.arsgrin.gov/4/cgi-bin/npgs/html/taxon.pl?316644> (18 October 2015)
13. Sharma, S. K. and Singh, A. P. (2011). Morphological, microscopical and physico-chemical investigations on the rhizomes of *Cyperus rotundus* Linn. Res. J. Pharm. Biol. Chem. Sci., 2(3):798-806.
14. Ali, E. A. (2016). A review on *Cyperus rotundus*: A potential medicinal plant, IOSR J. Pharm., 6(7):32-48.
15. Radanachalee, T.; Maxwell, J. F. and Meechai, M. (1994). Weeds of soybean fields in Thailand. Edn 1, Multiple Cropping Center, Faculty of Agriculture, Chaing Mai University, Thailand.
16. Galinato, M. I.; Moody, K. and Piggan, C. M. (1999). Upland rice weeds of South and Southeast Asia. IRRI, Philippines, pp:108-122.
17. Nidugala, H.; Avadhani, R.; Naraya, S. K.; Bhaskar, B. and Noojibail, A. (2013). Atlas of macro-microscopy of raw drug sold as musta – *Cyperus rotundus*. Int. J. Pharm. Sci. Res., 4(6):2308-2311.
18. Al-Snafi AE. A review on *Cyperus rotundus* A potential medicinal plant. IOSR Journal of Pharmacy. 2016;6(7):32-48.
19. Talukdar AD, Tarafdar RG, Choudhury MD, Nath D and Choudhury S. A review on pteridophyte antioxidants and their potential role in discovery of new drugs. Sci and Tech 2011; 7(1): 151-155.
20. Yeung HC. Handbook of Chinese herbs and formulas. Institute of Chinese Medicine; 1995.
21. Duke JA and Ayensu ES. Medicinal plants of China. Reference Publications, Inc. 1985.
22. Bown D. Encyclopaedia of herbs and their uses. Dorling Kindersley, London. 1995.
23. Chopra RN, Nayar SL and Chopra IC. Glossary of Indian medicinal plants. Council of Scientific and Industrial Research, New Delhi 1986.
24. Sivapalan SR and Jeyadevan P. Physico-chemical and phyto-chemical study of rhizome of *Cyperus rotundus* Linn. International Journal of Pharmacology and Pharmaceutical Technology (IJPPT) 2012; 1(2): 42-46.
25. Sharma SK and Singh AP. Morphological, microscopical and physico-chemical investigations on the rhizomes of *Cyperus rotundus* Linn. Research Journal of Pharmaceutical, Biological and Chemical Sciences 2011; 2(3): 798-806.
26. Emelugo BN, Umerie SC, Okonkwo IF and Achufusi JN. Evaluation of the tubers and oil of *Cyperus rotundus* Linn (Cyperaceae). Pakistan Journal of Nutrition 2011; 10(2): 147-150.
27. Ghannadi A, Rabbani M, Ghaemmaghami L and Malekian N. Phytochemical screening and essential oil analysis of one of the Persian sedges; *Cyperus rotundus* L. IJPSR 2012; 3(2): 424-427.
28. Chandratre RS, Chandarana S and Mengi SA. Lipid lowering activity of alcoholic extract of *Cyperus rotundus*. IJRPC 2011; 1(4): 1042- 1045.
29. Huang KC. The dried tuber of *Cyperus rotundus* L. The pharmacology of Chinese herbs, 2nd ed. 1999: 320-321.
30. El-Gohary HMA. Study of essential oils of the tubers of *Cyperus rotundus* L and *Cyperus alopecuroides* ROTTB. Bull Fac Pharm Cairo Univ 2004; 42(1) :157-164.
31. Bisht A, Bisht GRS, Singh M, Gupta R and Singh V. Chemical composition and antimicrobial activity of essential oil of tubers of *Cyperus rotundus* Linn. collected from Dehradun (Uttarakhand). International Journal of Research in Pharmaceutical and Biomedical Sciences 2011; 2(2); 661-665.
32. Bashir A, Sultana B, Akhtar FH, Munir A, Amjad M, Hassan Q. Investigation on the antioxidant activity of Dheela Grass (*Cyperus rotundus*). Afr J Basic Appl Sci. 2012;4(1):1-6.
33. Peerzada, A. M.; Ali, H. H.; Naem, M.; Latif, M.; Bukhari, A. H. and Tanveer, A. (2015). *Cyperus rotundus* L.: Traditional uses, phytochemistry, and pharmacological activities. J. Ethnopharmacol., 174(4):540-560.
34. Puratchikody A, Devi CN, Nagalakshmi G. Wound Healing Activity of Wound Healing Activity of *Cyperus rotundus* *Cyperus rotundus* Linn. Indian journal of pharmaceutical sciences. 2006 Jan:97-100.
35. Gupta, A.K., Eds., In; Quality Standards of Indian Medicinal Plants, Vol. I, Indian Council of Medical Research., New Delhi, 2003, 89-94.
36. Puratchikody, A., Jaswanth. A., Nagalakshmi, A., Kanagumeenal, P. and Ruckmani, K., Indian J. Pharm. Sci., 2001, 63, 326.



37. Puratchikody A, Devi Nithya C, Nagalakshmi G. Wound healing activity of *Cyperus rotundus* linn. Indian journal of pharmaceutical sciences 2006; 68: 97-101.
38. Sivapalan SR. Medicinal uses and pharmacological activities of *Cyperus rotundus* Linn-A Review. International Journal of Scientific and Research Publications. 2013 May;3(5):1-8.
39. Rajamanickam, M. and Rajamanickam, A. (2016). Analgesic and anti inflammatory activity of the extracts from *Cyperus rotundus* Linn rhizomes. J. Appl. Pharm. Sci., 6(9):197-203.
40. Gupta MB, Palit TK, Singh N, Bhargava KP. Pharmacological studies to isolate the active constituents from *Cyperus rotundus* possessing anti inflammatory, anti-pyretic and analgesic activities. Indian Journal of Medical Research 1971; 59: 76–82.
41. Kumar M, Rani M, Meher B. Review on Pharmacology and Phytochemistry of *Cyperus rotundus* L. Current Research in Pharmaceutical Sciences. 2017 May 8:11-5.
42. Kabbashi AS, Osman EE, Amel M, Abuzeid AN, Garbi MI, Waleed S, Mahmoud M K. Antiamoebic activity and cytotoxicity of ethanolic extract of *Cyperus rotundus*L. Advancement in Medicinal Plant Research, 2015;3(4):155-161.
43. Mohammad A, Nagarajaiah BH, Kudagi BL. Experimental Evaluation of Antiulcer Activity of *Cyperus rotundus*. AJBP. 2012; 2(2):261-268.
44. Singh N, Kulshrestha VK, Gupta MB and Bhargava K P., A pharmacological study of *Cyperus rotundus*, Indian J Med, Res, 1970,58, 103-109.
45. Chandratre RS, Chandarana S, Mengi SA. Lipid lowering activity of alcoholic extract of *Cyperus rotundus*. IJRPC 2011; 1(4): 1042-1045
46. Okwu GN, Abanobi SE, Nnadi UV, Ujowundu CO, Ene AC. OJBRR.2015;7(3):132-138
47. Ahmad M, Rehman AB, Mahummad R, and Wazir A. Assessment of Anti-inflammatory, Anti ulcer and Neuro Pharmacological activities of *Cyperus rotundus*L. PJPSC. 2014; 27(6):2241-2246.
48. Seo, W. G.; Pea, H. O.; Oh, G. H.; Chai, K. Y.; Kwon, T. O.; Yun, Y.G.; Kim, N. Y and Chung, H. T. (2001). Inhibitory effects of methanol extract of *Cyperus rotundus* Linn. Linn. rhizomes on nitric oxide and superoxide productions by murine acrophage cell line, RAW 264.7 cells. J. Ethnopharmacol., 76(1):59-64.
49. Sundaram, M. S.; Shivakumar, T. and Balamurugan, G. (2008).Anti inflammatory effect of *Cyperus rotundus* Linn. Leaves on acute and subacute inflammation in experimental rat models. Biomed., 28(2):302-304
50. Mazzio E A and Soliman K F A, In vitro screening for the tumoricidal properties of international medicinal herbs, Phytoter Res,2009, 23(3), 385- 398.
51. Kapadia, V. H.; Naik, V. G.; Wadia, M. S. and Sukh, D. (1967). Sesquiterpenoids from the essential oil of *Cyperus rotundus*. Tetrahedron Lett., 8(47):4661-4667.
52. Raut, N. A. and Gaikwad, N. J. (2012). Antidiabetic potential of fractions of hydro-ethanol extract of *Cyperus rotundus* L. (Cyperaceae). Res. J. Pharm. Biol. Chem. Sci., 3(4):1014-1019.
53. Shanmugasundaram, E. R.; Subramaniam, U.; Santhini, R. and Shanmugasundaram, K. R. (1986). Studies on brain structure and neurological function in alcoholic rats controlled by an Indian medicinal formula (SKV). J. Ethnopharmacol., 17(3):225-45.
54. Mohammad, A.; Nagarajaiah, B. H. and Kudagi, B. L. (2012).Experimental evaluation of antiulcer activity of *Cyperus rotundus*. Asian J. Biochem. Pharm. Res., 2(2):261-268.
55. Singh, N.; Pandey, B. R.; Verma, P.; Bhalla, M. and Gilca, M. (2012). Phytopharmacotherapeutics of *Cyperus rotundus* linn. (Motha). Indian J. Nat. Prod. Resour., 3(4):467-476.
56. Mokkahasmit, M.; Ngarmwathana, W.; Sawasdimongkol, K.; (1971). Pharmacological evaluation of Thai medicinal plants. J. Med. Assoc. Thai., 54(7):490-503.
57. Soltan, M. M. and Zaki, A. K. (2009). Antiviral screening of forty-two Egyptian medicinal plants. J. Ethnopharmacol., 126(1):102-107.
58. Thebtaranonth C, Thebtaranonth Y, Wanauppathamkul S, Yuthavong Y. Antimalarial sesquiterpenes from tubers of *Cyperus rotundus*: Structure of 10,12-peroxycalamenene, a sesquiterpene endoperoxide. Phytochemistry 1995; 40:125-8.
59. Imam H, Sofi G, Seikh A, Lone A. The incredible benefits of Nagarmotha (*Cyperus rotundus*). International Journal of Nutrition, Pharmacology, Neurological Diseases. 2014 Jan 1;4(1):23-7.
60. Chandratre RS, Chandarana S, Mengi SA. Lipid lowering activity of alcoholic extract of *Cyperus rotundus*. Int J Res Pharm Chem 2011;1:1042-5.

61. Singh N, Kulshrestha VK, Gupta MB and Bhargava KP. A pharmacological study of *Cyperus rotundus*. Indian J Med Res 1970; 58: 103-109.
62. Ahmad M, Rookh M, Rehman AB, Muhammad N, Amber, Younus M and Wazir A. Assessment of antiinflammatory, anti-ulcer and neuro-pharmacological activities of *Cyperus rotundus*Linn. Pak J Pharm Sci 2014; 27(6-Special): 2241-2246.
63. Seo EJ, Lee DU, Kwak JH, Lee SM, Kim YS and Jung YS. Antiplatelet effects of *Cyperus rotundus*and its component (+)-nootkatone. Journal of Ethnopharmacology 2011; 135: 48-54.
64. Nagulendrankr, Velavan S, Mahesh R. *In Vitro* Antioxidant Activity and Total Polyphenolic Content of *Cyperus rotundus*Rhizomes, E-Journal of Chemistry 2007; 4(3): p 440-449.
65. Meena AK, Yadav AK, Niranjana US, Singh B, Nagariya AK, Verma M. Review on *Cyperus rotundus*-A potential herb. International Journal of Pharmaceutical and Clinical Research. 2010 Jan;2 (1):20-24.
66. Uddin SJ, Mondal K, Shilpi JA, Rahman MT. Anti-diarrhoeal activity of *Cyperus rotundus*, Fitoterapia 2006; 77; 134-136.

<p><b>Image</b> <b>Author -1</b></p>	<p>Author Name – Corresponding Author: <b>Manoj A</b></p> <p>Author Affiliation - <b>Department of Pharmacognosy, College of Pharmacy Madras Medical College, Chennai, India.</b></p> <p>Author Address/Institute Address – <b>ORCID ID: 0009-0008-1783-2854</b></p>
<p><b>Image</b> <b>Author -2</b></p>	<p>Author Name: <b>Kumudhaveni B</b></p> <p>Author Affiliation- <b>Department of Pharmacognosy, College of Pharmacy Madras Medical College, Chennai, India.</b></p> <p>Author Address/Institute Address – <b>ORCID ID: 0000-0002-358-2808</b></p>
<p><b>Image</b> <b>Author -3</b></p>	<p>Author Name: <b>Abirami R</b></p> <p>Author Affiliation - <b>Department of Pharmacognosy, College of Pharmacy Madras Medical College, Chennai, India.</b></p> <p>Author Address/Institute Address – <b>ORCID ID: 0009-0004-1958-3098</b></p>

<p><b>Image</b> <b>Author -4</b></p>	<p>Author Name: <b>Vanitha N</b></p> <p>Author Affiliation - <b>Department of Pharmacognosy, College of Pharmacy Madras Medical College, Chennai, India.</b></p> <p>Author Address/Institute Address – <b>ORCID ID: 0009-0007-0242-2505</b></p>
<p><b>Image</b> <b>Author -5</b></p>	<p>Author Name: <b>Pooja S</b></p> <p>Author Affiliation - <b>Department of Pharmacognosy, College of Pharmacy Madras Medical College, Chennai, India.</b></p> <p>Author Address/Institute Address – <b>ORCID ID: 0009-0009-1675-7052</b></p>
<p><b>Image</b> <b>Author -6</b></p>	<p>Author Name: <b>Radha R</b></p> <p>Author Affiliation - <b>Department of Pharmacognosy, College of Pharmacy Madras Medical College, Chennai, India.</b></p> <p>Author Address/Institute Address – <b>ORCID ID: 0000-0001-9631-8440</b></p>