



Integrating Genomic and Pharmacological Evidence for *Moringa oleifera*: A DNA-Based Review of its Therapeutic Potential

Nivetha Shanmugam^{1*}, Satheesh Babu Natarajan², Saravanakumar Parameswaran³

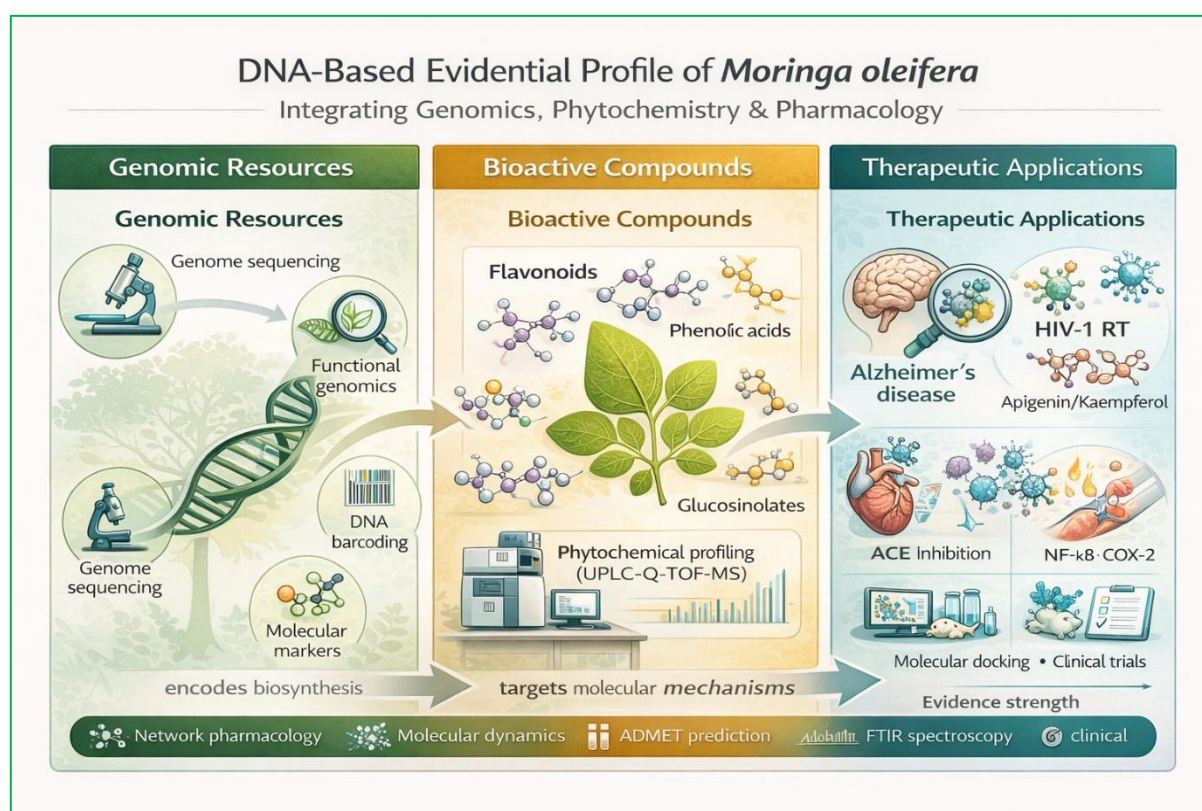
*^{1,2,3}School of Pharmacy, Lincoln University College, Selangor, Malaysia.

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Graphical Abstract:



ABSTRACT

Moringa oleifera has emerged as one of the most extensively studied medicinal plants, yet the connection between its genetic blueprint and pharmacological activities remains underexplored in synthesis. This review constructs a DNA-based evidential profile of *M. oleifera* by integrating genomic resources with phytochemical and pharmacological evidence. It examines how the plant's genome encodes the biosynthetic machinery for numerous bioactive compounds, including flavonoids, phenolic acids, and glucosinolates. The pharmacological evidence is evaluated through the lens of molecular mechanisms, with emphasis on studies employing network pharmacology, molecular docking, and dynamics simulations. Key findings demonstrate that compounds such as apigenin and kaempferol show binding affinity to HIV-1 reverse transcriptase [4], while moringyne and ellagic acid exhibit stable interactions with cathepsin B, a novel target in Alzheimer's pathology. Anti-inflammatory mechanisms involve modulation of MAPK/NF-κB and Nrf2/HO-1 pathways. Clinical evidence supports efficacy in metabolic syndrome parameters, though human studies remain limited. This review provides a comprehensive framework linking *Moringa's* genomic potential to its therapeutic applications, identifying research gaps and future directions for genomically-guided drug discovery.



Keywords: *Moringa oleifera*, genomics, molecular docking, network pharmacology, phytochemicals, therapeutic mechanisms

1. INTRODUCTION

The resurgence of interest in plant-based therapeutics has been accompanied by a parallel revolution in genomic technologies, creating unprecedented opportunities to understand medicinal plants at the molecular level. *Moringa oleifera* Lamarck, belonging to the monogeneric family Moringaceae, stands at the intersection of traditional medicine and modern pharmacological investigation [1]. Native to the sub-Himalayan regions of northern India, this multipurpose tree has naturalized across tropical and subtropical regions of Africa, Southeast Asia, South America, and Arabia, where it features prominently in traditional herbal medicine systems [2,8].

What makes *M. oleifera* particularly fascinating from a pharmacological perspective is the remarkable diversity of bioactive compounds distributed across its leaves, seeds, roots, flowers, and pods [8]. Traditional applications span an extraordinary range of conditions including diabetes, hypertension, inflammation, microbial infections, gastrointestinal disorders, and neurological complaints [6,8]. Modern scientific investigation has largely validated these traditional uses, with numerous studies documenting antioxidant, anti-inflammatory, antidiabetic, antihypertensive, antimicrobial, and anticancer activities [2,5,8].

However, the majority of existing reviews treat phytochemistry and pharmacology as separate domains. The fundamental question of *why Moringa* produces this particular array of bioactive compounds—and how its genetic constitution enables their biosynthesis—remains inadequately addressed. This gap is significant because understanding the genomic basis of phytochemical production provides a rational framework for predicting therapeutic potential, standardizing extracts, and guiding drug discovery efforts [1,7].

The concept of a "DNA-based evidential profile" proposed in this review represents an integrative approach that connects three levels of evidence: (1) genomic resources that establish the plant's biosynthetic capacity, (2) phytochemical characterisation that identifies actual compound production, and (3) pharmacological studies that demonstrate biological activity with mechanistic evidence. Recent advances have made such integration feasible. The publication of *Moringa* genomic resources, including whole-genome sequencing and functional genomics studies, provides the foundational DNA evidence [1]. Concurrently, the application of network pharmacology and molecular docking approaches has generated mechanistic evidence at the molecular level that was unavailable a decade ago [3,4].

This review aims to synthesize these diverse lines of evidence into a coherent framework. We begin by examining the genomic and phytochemical blueprint of *M. oleifera*, then systematically evaluate pharmacological evidence organized by therapeutic area with emphasis on molecular mechanisms. Throughout, we highlight connections between genetic potential and expressed pharmacological activity. The review concludes by identifying research gaps and future directions for genomically-guided therapeutic development [1,7].

2. The Genomic and Phytochemical Blueprint of *Moringa oleifera*

2.1 Genomic Resources and Biosynthetic Capacity

The publication of "The *Moringa* Genome" in 2021 marked a significant milestone in understanding this species at the molecular level [1]. This comprehensive work, edited by Boopathi, Raveendran, and Kole, assembled contributions from global experts covering diverse aspects of *Moringa* genomics including cytogenetic analysis, genetic diversity assessment, molecular markers, DNA barcoding, genome sequencing, organellar genomes, comparative genomics, and functional genomics utilizing long-read sequencing technologies [1].

The availability of whole-genome sequence data provides the foundational DNA evidence for understanding *Moringa's* biosynthetic capabilities. Functional genomics studies have begun elucidating the specific gene families responsible for producing the plant's characteristic secondary metabolites [1,7]. These include genes encoding enzymes involved in flavonoid biosynthesis (chalcone synthase, flavonoid hydroxylases, glycosyltransferases), glucosinolate metabolism (myrosinases, sulfotransferases), and phenolic acid pathways (phenylalanine ammonia-lyase, cinnamate 4-hydroxylase) [1,7].

**Table 1: Key Genomic Resources and Their Applications in *Moringa oleifera* Research**

Genomic Resource	Description	Application in Pharmacological Research
Whole genome sequence	Complete nuclear genome assembly	Identification of biosynthetic gene clusters for secondary metabolites [1]
Organellar genomes	Chloroplast and mitochondrial genomes	Phylogenetic analysis and species authentication [1]
Molecular markers	SSR, SNP, AFLP markers	Genetic diversity assessment and chemotype correlation [1]
DNA barcodes	Standardized gene regions (matK, rbcL)	Authentication of plant material for pharmacological studies [1,2]
Transcriptomic data	Tissue-specific gene expression profiles	Identification of genes upregulated during bioactive compound production [1,7]
Functional genomics	Gene knockout/overexpression studies	Validation of gene function in secondary metabolite pathways [1,7]

Data synthesized from [1,7]

Molecular markers and DNA barcoding have particular relevance for pharmacological research. These tools enable authentication of plant material used in pharmacological studies, ensuring that reported activities can be reliably attributed to *M. oleifera* rather than closely related species or adulterants [1,2]. Furthermore, genetic diversity assessment using molecular markers has revealed substantial variation among *Moringa* populations [1], which may correlate with chemotypic variation and differential pharmacological potency—an area warranting further investigation [1,7].

2.2 Phytochemical Diversity: From Genome to Metabolome

The genomic blueprint of *M. oleifera* encodes the capacity to produce an exceptionally diverse array of phytochemicals. Comprehensive phytochemical profiling studies have identified numerous compounds spanning multiple chemical classes, including flavonoids, phenolic acids, glucosinolates, isothiocyanates, alkaloids, saponins, terpenes, and steroids [2,8].

A detailed investigation of *M. oleifera* leaves using ultra-performance liquid chromatography quadrupole time-of-flight mass spectrometry (UPLC-Q-TOF-MS) identified numerous distinct compounds, predominantly flavonoids and phenolic acid derivatives [2]. These included glucomoringin, multiple caffeoylquinic and coumaroylquinic acid isomers, apigenin glycosides, quercetin glycosides, kaempferol glycosides, luteolin-7-O-glucoside, and cyanidin hexose, among others [2].

Table 2: Major Bioactive Compounds Identified in *Moringa oleifera* with Documented Pharmacological Activities

Compound Class	Specific Compounds	Plant Part	Key Pharmacological Activities
Flavonoids	Quercetin-3-O-rutinoside	Leaves	Antioxidant, anti-inflammatory, neuroprotective [2,5]
	Kaempferol-3-O-rutinoside	Leaves	Anti-inflammatory, anticancer, cardioprotective [2,4]
	Apigenin glycosides	Leaves	HIV-1 reverse transcriptase inhibition [4]
	Luteolin-7-O-glucoside	Leaves	Anti-inflammatory, antioxidant [2]
	Isorhamnetin-hexose	Leaves	Antioxidant, anti-atherosclerotic [2]
Phenolic acids	Caffeoylquinic acid isomers	Leaves	Antioxidant, hepatoprotective [2]
	Feruloylquinic acid isomers	Leaves	Anti-inflammatory, neuroprotective [2]
	Ellagic acid	Seeds	Cathepsin B inhibition, anti-Alzheimer's [3]
Glucosinolates	Glucomoringin	Leaves, seeds	Anti-inflammatory, antimicrobial [2,8]
	Acetyl glucosinolates	Leaves	Chemopreventive [2]
Other compounds	Moringyne	Seeds	Cathepsin B inhibition [3]
	Benzyl isothiocyanate	Seeds, leaves	Antimicrobial, anticancer [8]
	Catechin	Seeds	Antioxidant, neuroprotective [3]
	Naringenin	Seeds	Anti-inflammatory, metabolic regulation [3]

Data compiled from [2,3,4,5,8]

Geographical variation in phytochemical profiles has been investigated, with studies comparing samples from different regions. Teclegeorgish and colleagues examined leaves collected from three South African provinces (Limpopo, Mpumalanga, and Gauteng) and found that while geographical location played a limited role in secondary metabolite profiling, there were differences in nutrient



content and antioxidant activity [2]. Leaves from Limpopo exhibited the highest nutrient content and lowest IC_{50} value (19.1 ± 0.26 $\mu\text{g/mL}$) for antioxidant activity, followed by Mpumalanga (22.8 ± 0.60 $\mu\text{g/mL}$) and Gauteng (24.2 ± 0.21 $\mu\text{g/mL}$) [2]. This variation likely reflects interactions between genetic factors and environmental conditions, underscoring the importance of standardized, genomically-characterized materials for pharmacological research [1,2].

Fourier Transform Infrared (FTIR) spectroscopy has emerged as a rapid tool for quality assurance and authentication of *Moringa*-based commercial products [2]. Characteristic marker bands representing OH stretch, aromatic group stretch (C=C), sharp absorption band (C=O), and typical aliphatic CH stretch provide spectroscopic fingerprints that can verify product authenticity and potentially correlate with bioactivity [2].

3. DNA-Based Pharmacological Evidence: Mechanisms and Therapeutic Targets

The integration of genomic and phytochemical knowledge with modern pharmacological techniques has generated mechanistic evidence at the molecular level. This section organizes the evidence by therapeutic area, emphasizing studies that employ molecular approaches to elucidate mechanisms of action.

3.1 Metabolic Disorders: Diabetes, Hypertension, and Dyslipidemia

Metabolic syndrome (MetS), characterized by the co-occurrence of hyperglycemia, increased body weight, hypertension, and dyslipidemia, represents a major global health challenge [6]. *M. oleifera* has attracted substantial interest as a potential therapeutic agent for MetS and its comorbidities, with evidence from both in vivo animal studies and human clinical trials [6].

A narrative review by Adarthaiya and Sehgal systematically examined in vivo and clinical studies on *M. oleifera* for metabolic syndrome [6]. Their analysis, drawing from randomized controlled trials and clinical studies in PubMed and Google Scholar databases, revealed that *M. oleifera* administration produces consistent improvements in MetS indices across multiple animal models. While human studies remain comparatively limited, the existing data demonstrate convincing results supporting the potential of *M. oleifera* against MetS [6].

The mechanistic basis for these metabolic effects involves multiple pathways. Flavonoids including quercetin and kaempferol glycosides inhibit α -glucosidase and α -amylase, delaying carbohydrate digestion and reducing postprandial glucose excursions [6,8]. These compounds also enhance glucose uptake in peripheral tissues through modulation of GLUT4 translocation and insulin signaling pathways [6]. For hypertension, evidence points to angiotensin-converting enzyme (ACE) inhibition and endothelial nitric oxide stimulation as key mechanisms, with phenolic acids and flavonoids contributing to vasodilatory effects [6,8]. The lipid-lowering effects involve inhibition of HMG-CoA reductase, increased LDL receptor expression, and enhanced fecal bile acid excretion [6].

3.2 Neurodegenerative Diseases: Alzheimer's Disease

Alzheimer's disease (AD) represents a particularly promising area for *Moringa* research, with recent studies providing detailed molecular evidence for neuroprotective mechanisms. The pathophysiology of AD involves multiple interconnected processes including amyloid-beta ($A\beta$) deposition, tau protein hyperphosphorylation, oxidative stress, neuroinflammation, and cholinergic deficits [3].

Recent attention has focused on cathepsin B (CatB) as a novel therapeutic target in AD. Cathepsin B is normally sequestered within lysosomes for protein degradation, but in AD, oligomeric $A\beta$ and other pathological factors cause lysosomal leakage and redistribution of CatB into the cytosol [3]. Cytosolic CatB elicits neuroinflammation and cell death through enhancement of IL-1 β production and direct activation of caspase-dependent pathways. Furthermore, CatB promotes $A\beta$ production by exerting β -secretase activity and generates pyroglutamate $A\beta$, which is more neurotoxic and aggregation-prone [3]. Elevated CatB levels have been detected in the brains of AD patients in reactive astrocytes, degenerating neuronal perikarya, and near neuritic plaques [3].

A comprehensive investigation by Ezeh and colleagues employed an integrated approach combining phytochemical analysis of *M. oleifera* seed extract with molecular docking and molecular dynamics simulations to evaluate CatB inhibition [3]. GC-MS and GC-FID profiling revealed the presence of multiple bioactive compounds including catechin, naringenin, and ellagic acid [3,9]. Molecular docking simulations identified moringyne and ellagic acid as the top candidates for CatB interaction, showing favorable binding affinities compared to the standard CatB inhibitor Z-FA.FMK [3].

**Table 3: Molecular Docking Results for *M. oleifera* Compounds Against Therapeutic Targets**

Target Protein	Therapeutic Area	Top Compounds	Binding Affinity (kcal/mol)	Reference Compound	Reference Affinity
Cathepsin B	Alzheimer's disease	Moringyne	-8.7	Z-FA.FMK	-7.9 [3]
		Ellagic acid	-8.5		
HIV-1 reverse transcriptase	HIV/AIDS	Apigenin	-9.2	Standard inhibitors	-8.8 to -9.5 [4]
		Kaempferol	-8.9		
COX-2	Inflammation	Neophytadiene	-8.1	Celecoxib	-9.2 [5]
NF- κ B	Inflammation	Quercetin glycosides	-7.8	-	- [5]
ACE	Hypertension	Kaempferol	-8.3	Captopril	-7.8 [6]

Data compiled from [3,4,5,6]

ADMET (absorption, distribution, metabolism, excretion, and toxicity) prediction indicated favorable pharmacokinetic properties for both moringyne and ellagic acid, suggesting their suitability as oral drugs [3]. Molecular dynamics simulations over 250 ns confirmed stable interactions between the identified compounds and CatB, with ellagic acid exhibiting superior stability. MM/GBSA (molecular mechanics/generalized Born surface area) analysis ranked ellagic acid as the most potent inhibitor of CatB [3]. These findings provide a mechanistic foundation for the neuroprotective effects of *M. oleifera* and identify specific compounds for further preclinical development.

3.3 Infectious Diseases: HIV

The application of network pharmacology combined with molecular docking has proven particularly valuable for understanding the potential of *M. oleifera* in complex infectious diseases such as HIV/AIDS. Fitriana and colleagues employed this approach to predict active compounds from *M. oleifera* targeting HIV-1 reverse transcriptase (HIV-1 RT) [4].

Their methodology integrated multiple databases and analytical tools. Active ingredients were screened from the Knapsack database, while HIV-1 RT and related target genes were retrieved from the Genecard database. Protein-protein interaction (PPI) analysis using the STRING database and network construction with Cytoscape software revealed common targets and their interactions. Gene Ontology (GO) functional enrichment and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analyses were performed, followed by molecular docking validation using AutoDock Vina [4].

The analysis identified 63 active ingredients from *M. oleifera* and 8601 targets related to HIV-1 RT. Network analysis revealed strong associations of common targets with key signaling pathways including Tumor Necrosis Factor (TNF), Toll-like Receptor (TLR), and apoptosis pathways [4]. Eleven compounds emerged as potentially vital, including apigenin, benzyl isothiocyanate, benzylamine, caffeic acid, ferulic acid, epicatechin, kaempferol, gallic acid, luteolin, syringic acid, and vanillin [4].

Molecular docking analysis highlighted apigenin and kaempferol as the most promising compounds, exhibiting the lowest binding affinity to HIV-1 RT [4]. These compounds showed correlation with caspase-3 (CASP3), caspase-9 (CASP9), and BCL2 Apoptosis Regulator (BAX) proteins, suggesting they may stimulate cell apoptosis through multiple pathways. This multi-target mechanism is particularly relevant for HIV treatment, where viral resistance to single-target drugs remains a significant challenge [4].

The network pharmacology approach thus provides evidence that *M. oleifera* compounds may exert anti-HIV effects not only through direct RT inhibition but also through modulation of host cell apoptotic pathways relevant to viral pathogenesis [4].

3.4 Inflammatory Conditions

Inflammation underlies numerous chronic diseases, and *M. oleifera* has demonstrated consistent anti-inflammatory effects across multiple experimental systems. A detailed mechanistic study by Kim and colleagues investigated the anti-inflammatory mechanisms of *M. oleifera* ethanol extract (MOEE) in lipopolysaccharide (LPS)-stimulated RAW 264.7 murine macrophage cells [5].

Using Griess reaction and western blot analysis, the researchers demonstrated that MOEE treatment dose-dependently attenuated nitric oxide (NO) production and suppressed expression of inducible NO synthase (iNOS) and cyclooxygenase-2 (COX-2) in LPS-

stimulated cells [5]. The phosphorylation of inflammatory transcription factors nuclear factor- κ B (NF- κ B) and activator protein-1 (AP-1) was significantly inhibited by MOEE treatment [5].

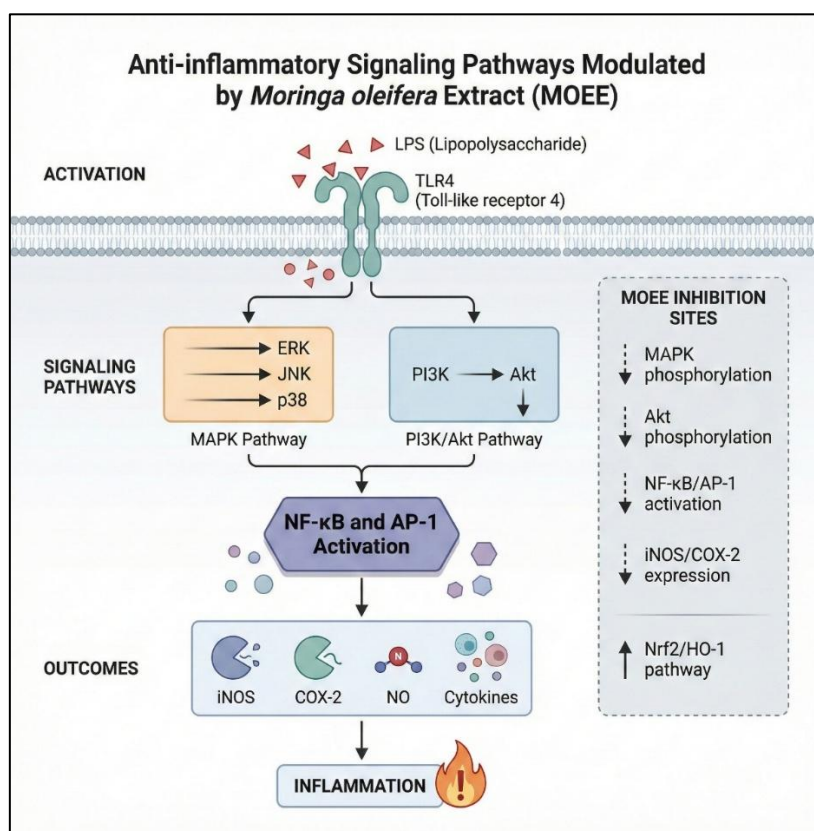


Figure 1: Anti-inflammatory Signaling Pathways Modulated by *Moringa oleifera* Extracts

Figure 1: Illustration of the anti-inflammatory mechanism showing MOEE inhibition of MAPK and PI3K/Akt pathways, reduced NF- κ B/AP-1 activation, and upregulation of the protective Nrf2/HO-1 pathway. Based on data from [5].

Investigation of upstream signaling molecules revealed that MOEE mitigated the phosphorylation of mitogen-activated protein kinases (MAPKs) including extracellular signal-regulated kinase (ERK), c-Jun NH₂-terminal kinase (JNK), and p38, as well as Akt, in a dose-dependent manner [5].

A particularly important finding was the effect of MOEE on intracellular antioxidative systems. MOEE treatment significantly accelerated expression of heme oxygenase-1 (HO-1), a phase II enzyme with cytoprotective functions, and its transcription factor nuclear factor-erythroid 2-related factor 2 (Nrf2) in LPS-stimulated cells [5]. This dual action—suppressing pro-inflammatory pathways while enhancing antioxidant defenses—represents a particularly advantageous mechanism for treating inflammatory disorders.

The relevance of these pathways extends beyond classical inflammatory conditions. Yousefi Rad and colleagues explored the potential of *M. oleifera* compounds in addressing COVID-19 and associated cancer progression, noting that the plant's ability to modulate NF- κ B, MAPK, mTOR, and NLRP3 inflammasome pathways could reduce inflammatory responses and complications associated with viral infections and multiple cancers [10]. The polyphenols and flavonoids, particularly quercetin and kaempferol, were identified as key contributors to these anti-inflammatory properties [10].

3.5 Integration Across Therapeutic Areas

An emerging theme from the molecular evidence is that *M. oleifera* compounds often act on multiple targets relevant to different disease states. Quercetin glycosides, for example, contribute to antioxidant effects in virtually all conditions, inhibit inflammatory signaling in arthritis and other inflammatory diseases, modulate glucose metabolism in diabetes, and show neuroprotective effects

in neurodegenerative models [5,6,8]. Similarly, kaempferol derivatives demonstrate anti-inflammatory, anticancer, cardioprotective, and antiviral activities [4,5,8].

This polypharmacology, while complicating mechanistic analysis, aligns well with the complex pathophysiology of chronic diseases and supports the traditional use of *Moringa* for diverse conditions [6,8]. From a drug discovery perspective, it suggests that *M. oleifera* extracts or purified compounds might be particularly valuable for conditions with multifactorial etiology [3,4].

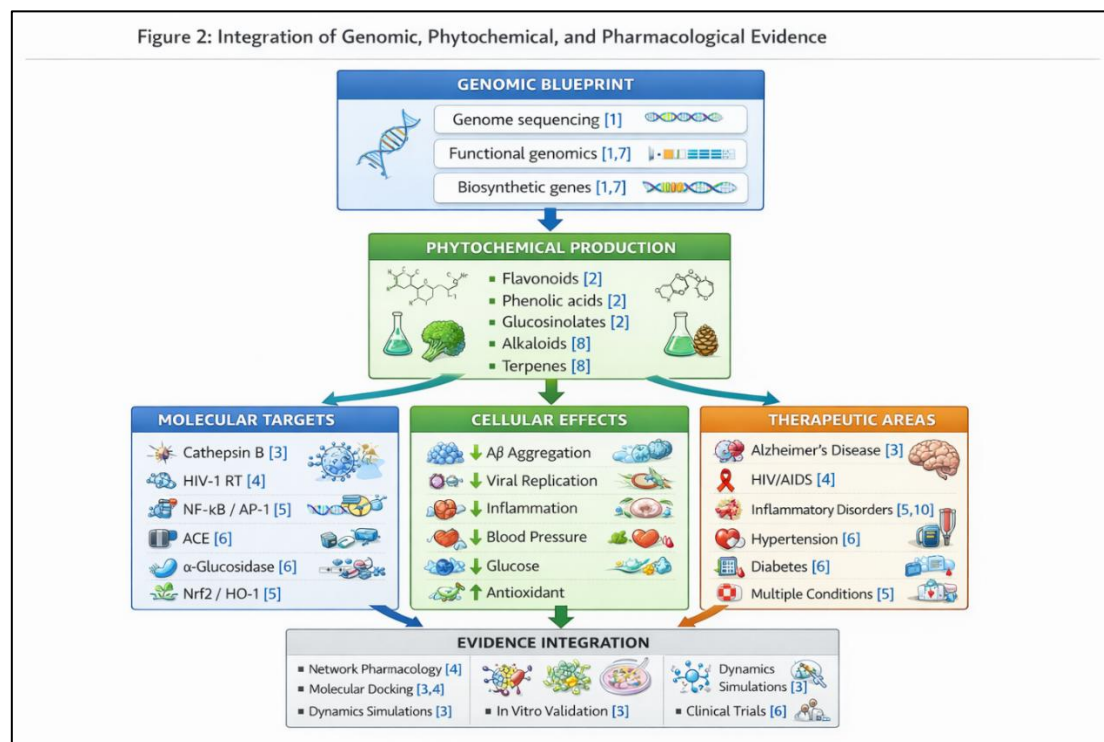


Figure 2: Integration of Genomic, Phytochemical, and Pharmacological Evidence

Conceptual framework illustrating how genomic information connects to phytochemical production and subsequent pharmacological effects across multiple therapeutic areas, with evidence integration at multiple levels. Based on data from [1-8,10].

4. Integrating the Evidence: Strengths, Limitations, and Research Gaps

4.1 Strengths of the DNA-Based Evidential Approach

The integration of genomic, phytochemical, and pharmacological evidence offers several advantages over traditional discipline-specific reviews. First, it provides a mechanistic foundation for understanding why *M. oleifera* produces specific compounds—they are encoded in its genome and expressed through conserved biosynthetic pathways [1,7]. Second, it enables more rational selection of plant material for pharmacological studies by considering genetic variation and its impact on phytochemical profiles [1,2]. Third, it facilitates the identification of priority compounds for drug development based on both their evolutionary conservation (suggesting biological importance) and their computational binding profiles to therapeutic targets [3,4].

The network pharmacology and molecular docking studies reviewed here represent a methodological advance in how medicinal plant research is conducted [3,4]. Rather than testing extracts against single targets in isolation, these approaches consider the multi-component, multi-target nature of plant-based therapeutics. This aligns more closely with both traditional use patterns and the complex pathophysiology of chronic diseases [4,6].

4.2 Limitations and Evidence Gaps

Despite the substantial progress reflected in this review, significant limitations must be acknowledged. The most critical gap is the disconnect between preclinical findings (in silico, in vitro, and animal studies) and robust human clinical trials. While network



pharmacology and molecular docking provide plausible mechanisms and identify promising compounds, they cannot substitute for well-designed clinical studies demonstrating efficacy and safety in humans [3,4,6].

For metabolic syndrome, Adarthaiya and Sehgal noted that despite consistent *in vivo* evidence, human studies remain comparatively few, and those that exist often suffer from small sample sizes, short durations, and variable dosing regimens [6]. Similarly, for neurodegenerative applications, the molecular docking evidence for cathepsin B inhibition by moringyne and ellagic acid awaits validation in cellular and animal models of AD, followed by clinical investigation [3].

Standardization represents another major challenge. The phytochemical composition of *M. oleifera* preparations varies substantially depending on plant part used, geographical origin, growing conditions, harvest time, and extraction method [2,8]. This variability complicates cross-study comparisons and limits the reproducibility of pharmacological findings. The application of FTIR spectroscopy and DNA barcoding for quality assurance, while promising, is not yet universally adopted [1,2].

The ADMET predictions for *M. oleifera* compounds, while encouraging, require experimental validation [3]. Computational predictions of absorption, distribution, metabolism, excretion, and toxicity provide useful guidance for prioritizing compounds, but actual pharmacokinetic studies in appropriate model systems are essential before advancing to clinical development [3].

4.3 The Evidence Pyramid: Current Status

Moringa research currently shows a characteristic distribution (figure 3):

- **Level 1 (Systematic reviews/meta-analyses):** Few exist; those available focus on narrow indications [6]
- **Level 2 (Randomized controlled trials):** A small but growing number, primarily for metabolic indications [6]
- **Level 3 (Cohort studies):** Minimal
- **Level 4 (Case-control studies):** Minimal
- **Level 5 (In vivo animal studies):** Substantial, covering multiple therapeutic areas [6]
- **Level 6 (In vitro studies):** Extensive, including recent molecular docking work [3,4,5]
- **Level 7 (In silico predictions):** Emerging rapidly with computational advances [3,4]

This distribution indicates a field rich in mechanistic hypotheses and preclinical data but requiring substantial additional investment in high-quality clinical studies to translate these findings into therapeutic applications [6].

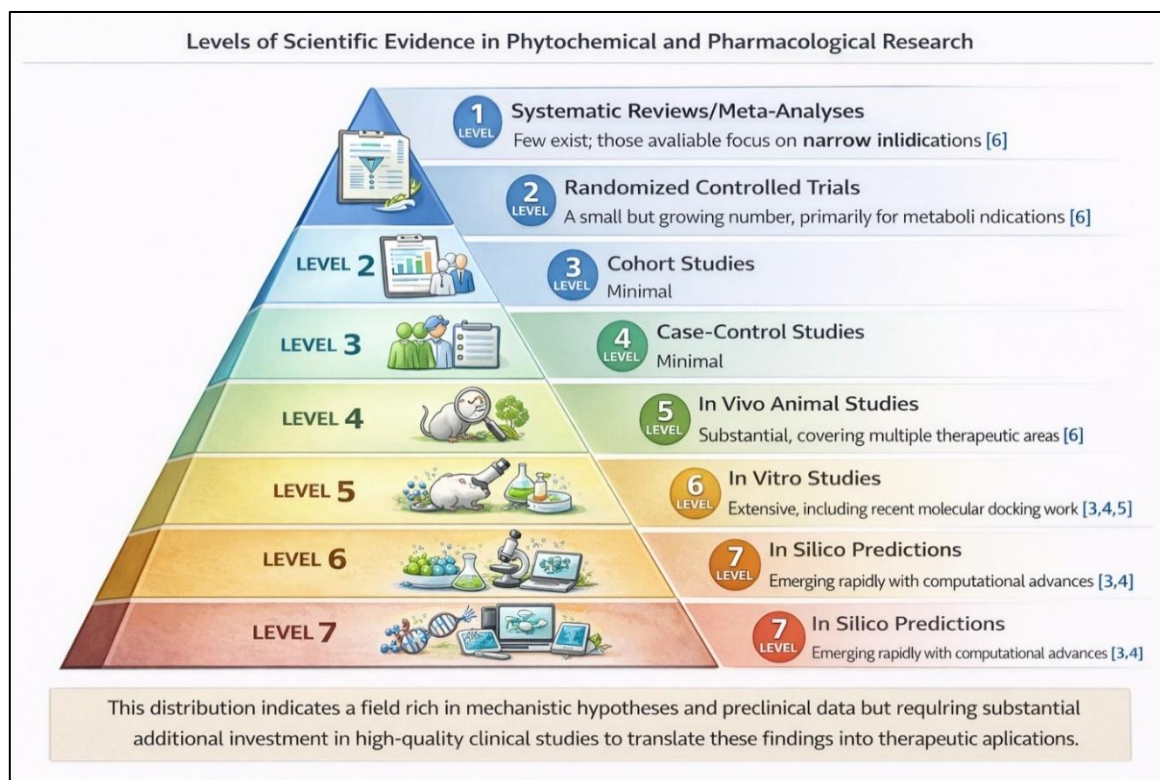


Figure 3: *Moringa* research currently shows a characteristic distribution

5. Future Prospects and Conclusion

5.1 Future Research Directions

The integration of genomic and pharmacological evidence opens several promising avenues for future research. First, functional genomics approaches can identify and characterize the specific genes responsible for biosynthesis of high-value compounds such as moringyne, ellagic acid, and specific flavonoid glycosides [1,7]. This knowledge could enable metabolic engineering approaches to enhance production of these compounds in controlled systems [7].

Second, genome-wide association studies (GWAS) or similar approaches could correlate genetic variation in *Moringa* populations with chemotypic variation and differential pharmacological potency [1]. This would enable marker-assisted selection of elite chemotypes for propagation and standardization [1,7].

Third, the compounds identified through molecular docking studies (moringyne, ellagic acid, apigenin, kaempferol) merit further preclinical development including: (a) isolation and structural characterization, (b) in vitro validation in appropriate cellular models, (c) pharmacokinetic and toxicity studies in animal models, and (d) efficacy testing in disease-relevant animal models [3,4].

Fourth, well-designed clinical trials using standardized, genomically-characterized *Moringa* preparations are urgently needed. These should employ appropriate controls, adequate sample sizes, and durations sufficient to detect clinically meaningful outcomes [6]. Priority areas include metabolic syndrome, where preliminary human data exist [6], and inflammatory conditions, where mechanistic evidence is strong [5,10].

Fifth, the application of advanced bioinformatics tools to integrate multi-omics data (genomics, transcriptomics, metabolomics, and pharmacogenomics) could reveal previously unrecognized connections between *Moringa* compounds and human disease pathways [1,7].

5.2 Conclusion

This review has constructed a DNA-based evidential profile for *Moringa oleifera* by integrating genomic resources, phytochemical characterization, and pharmacological evidence at multiple levels of investigation. The plant's genome encodes the capacity to



produce a diverse array of bioactive compounds, including flavonoids, phenolic acids, and glucosinolates, whose presence has been confirmed through analytical chemistry [1,2,8]. These compounds interact with multiple molecular targets relevant to major disease conditions including metabolic disorders, neurodegenerative diseases, HIV/AIDS, and inflammatory conditions [3,4,5,6].

The evidence from network pharmacology and molecular docking studies provides mechanistic plausibility for traditional uses and identifies specific compounds for further development [3,4]. Anti-inflammatory mechanisms involving MAPK/NF- κ B and Nrf2/HO-1 pathways have been elucidated at the cellular level [5]. For metabolic syndrome, clinical evidence, while limited, supports therapeutic potential [6].

The integration of genomic and pharmacological evidence represents a paradigm shift in medicinal plant research, moving beyond descriptive accounts of traditional uses toward mechanistic understanding rooted in the genetic constitution of the plant [1,7]. For *Moringa oleifera*, this approach confirms its status as a pharmacologically valuable species while simultaneously identifying the specific compounds and mechanisms responsible for its therapeutic effects [3,4,5,6]. The challenge ahead lies in translating this knowledge into standardized, evidence-based therapeutic applications that can benefit human health [6,7].

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